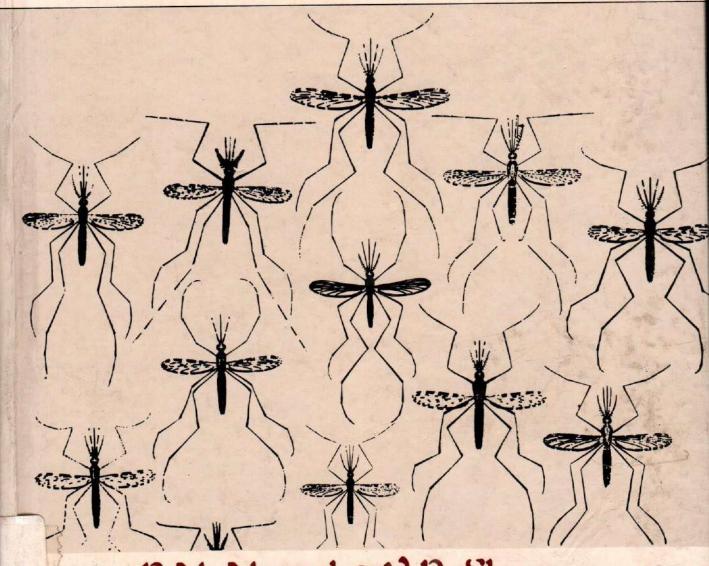
INDIAN ANOPHELINES



B.N. Nagpal • V.P. Sharma

A. CC NO - 4580

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B.N. Nagpal V.P. Sharma





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PREFACE

Sometime around 1679, appeared in Italy the first book on mosquitoes. It took another 75 years or so before the first description of anopheline larva was made in 1754. Again it was not until 1897 when Sir Ronald Ross in India incriminated mosquitoes in malaria transmission that there was any taxonomic study of mosquitoes. The turn of the century however, marked a watershed in malarial studies: there was a tremendous upsurge of interest throughout the world on mosquito systematics. In 1933 an excellent publication on "anophelines" by Sir S.R. Christophers in the Fauna of British India series laid the foundation of taxonomic research. There have since been contributions that deserve mention, for example, the 'Vectors of Malaria in India' published by the National Society for Malaria and Other Communicable Diseases in 1957 and 'Anophelines of India' by T. Ramachandra Rao in 1984. Both these books were useful additions to our mosquito systematics, biology and control. These are classic works to doubt, but unfortunately they have become outdated. There was therefore a let need for a comprehensive book on the Indian Anopheles mosquitoes.

We therefore took up a study of Indian anophelines, spread over a decade and spanning both laboratory and field investigations. Embodying the results of our studies, this book provides keys to the identification of all *Anopheles* mosquitoes reported from India with detailed drawings of the species, geographical distribution and their role in disease transmission. We hope this book would be a useful manual to the students, researchers and those associated with malaria control. We also expect that this compilation would go a long way in our fight against vector borne diseases.

We thank Dr. R.A. Ward, Editor of the American Mosquito Control Association, Washington, USA for reviewing the manuscript and for making many useful suggestions. The art work was done by late (Mrs.) Meenu Talwar, who worked treessly in preparing the plates. Thanks are also due to Dr.(Mrs.) Aruna Srivastava, technical staff of the computer section, MRC and field staff for their help and cooperation.

December 1994

B.N. NAGPAL V.P. SHARMA

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INTRODUCTION

Among insects, mosquitoes are the most important group; they transmit diseases such as malaria, filaria, dengue, Japanese encephalitis, and yellow fever. Knowledge of mosquitoes in India dates back to the second century. Susruta, in his famous work *Susruta Samhita*, described 12 kinds of life-destroying insects including mosquitoes and grouped them into Five categories — Samudrah, Parimandala, Hastimashaka, Krishna, and Parvatiya, on the basis of ecological, morphological, and causative characters. Sir Ronald Ross's discovery of malaria parasite oocystes in the gut of *Anopheles* mosquitoes in 1897 stimulated a great deal of interest leading to an upsurge of contributions on various aspects of mosquitoes.

Mosquitoes are characterized by a slender elongated body covered with scales and the possession of piercing and sucking mouth parts. The insects belong to the phylum Arthropoda, order Diptera, suborder Nematocera, family Culicidae, and subfamilies Anophelinae, Culicinae, and Toxorhynchitinae. Table 1 shows the systematic position of anopheline mosquitoes with specific and distinctive characters of phylum, class, subclass, order, division, suborder, family, subfamily, and genus. The mosquitoes of Toxorhynchitinae do not suck blood nor do they transmit any disease. The subfamilies Anophelinae and Culicinae are the ones which transmit disease.

The subfamily Anophelinae consists of Three genera — Anopheles, Bironella, and Chagasia—the subfamily Culicinae 33 genera, and Toxorhynchitinae only one genus. Worldwide there are over 3200 species of mosquitoes under 37 genera. In India the genera Anopheles, Aedes, Mansonia, and Culex are important inasmuch as they transmit malaria, dengue, filaria, and Japanese encephalitis. The genus Anopheles consists of about 420 species, of which 50 are well-known vectors of malaria. Table 2 gives the distribution of the anophelines in 12 epidemiological zones of the world. The subgenus Anopheles is found in all the 12 zones with a maximum distribution in Malaysian (56 species) and a minimum from Afrotropical zones (11 spp.). The subgenus Cellia is found in 10 zones, and a maximum number of species is recorded from Afrotropical zone (115 spp.) and a minimum from South American zone (1 sp. only). The subgenus Kerteszia is reported from only three American zones, with a maximum from South America (11 spp.), followed by Central America (5 spp.) and North America (1 sp. only). The subgenus Lophopodomyia is reported from two zones—South American (6 spp.) and Central American (1 sp. only). The subgenus Nyssorhynchus is reported only from four epidemiological zones with a maximum distribution in South America (30 spp.), followed by Central American (9 spp.), North American (4 spp.), and Chinese zone (1 sp.). The subgenus Stethomyia is reported from two regions only, South American (5 spp.) and Central American (2 spp.). Table 3 lists the countries with code numbers under 12 epidemiological zones of malaria to help locate the country in Fig. 1. In India, 58 species of anophelines are found, of which 6 are primary vectors - An. culicifacies, An. stephensi, An. fluviatilis, An. dirus,

An. sundaicus, and An. minimus — and 4 are secondary vectors, — An. annularis, An. philippinensis, An. jeyporiensis and An. varuna.

The genus Anopheles differs morphologically from other genera by virtue of an elongated slender palpus of length equal to that of the proboscis. Male palpi are club-shaped. Table 4 and Fig. 2 give the differences between the genera Anopheles and Culex in the egg, larva, pupa and adult stages. The diversity in characters and their ability to transmit the disease have prompted entomologists to conduct faunistic, ecological, vector incrimination and other studies.

Giles (1900, 1902) and Theobald (1901a, 1910) were the pioneers in the field of taxonomy and their contributions to mosquito systematics are unparalleled. Giles, in his book on *Gnats or Mosquitoes* published in 1900, gave the description of 242 species belonging to 8 genera: *Megarhinus* (18 spp.), *Anopheles* (30 spp.), *Psorophora* (3 spp.), *Sabethes* (3 spp.), *Culex* (160 spp.), *Aedes* (13 spp.), *Corethra* (12 spp.), and *Mochloynyx* (3 spp.). He grouped 20 species under continental Asia including three species under the genus *Megarhina*, six species under the genus *Anopheles*, and 11 species under the genus *Culex*. The anophelines listed were, *An. lindesayii* Giles, *An. rossii* Mittri, and *An. sinensis* Wied A and B.

In the second edition published in 1902, Giles described 252 species under 24 genera which included the redescribed species and new ones. A total of 47 species was described from India (excluding 2 spp. from Sri Lanka), of which 14 were anophelines which comprised 11 species and 3 subspecies, *Megarhina* (2 spp.), *Mucidus* (1 sp. only), *Stegomyia* (8 spp.), *Armigeres* (2 spp.), *Culex* (14 spp.), *Taeniorhynchus* (1 sp.), *Panoplites* (2 spp.), *Aedes* (1 sp.), *Aedeomyia* (1 sp.), and *Corethra* (1 sp.).

Of the anophelines the following species were described from India: An. niger-rimus Giles; An. jamesii Theobald; An. sinensis Wiedemann, (a) ssp. An. fuliginosus Giles, (b) ssp. An. indiensis Theobald and (c) ssp. An. annularis Van der Wulp; An. barbirostris Van der Wulp; An. indicus Theobald; An. rossii Giles; An. lindesayi Giles; An. culicifàcies Giles; An. gigas sp.n.; An. listoni sp.n.; and An. theobaldi sp.n.

Theobald, in his monograph on the Culicidae published originally in 1901 (Vol. 1), grouped mosquitoes of the world under 22 genera, of which 9 were from India: — Anopheles (6 spp.), Megarhinus (2 spp.), Stegomyia (4 spp.), Armigeres (2 spp.), Culex (14 spp.), Taeniorhynchus (1 sp.), Panoplites (2 spp.), Aedeomyia (1 sp.) and Corethra (1 sp.).

Theobald published two more volumes in 1903 and 1907, in which he classified mosquitoes under 10 subfamilies: Anophelinae, Megarhiniae, Culicinae, Aedinae, Uranotaeninae, Deinoceratinae, Heptaphlebomyinae, Trichoprosoponinae, Dendromyinae and Limatinae. The subfamily Anophelinae contained 18 genera, the subfamily Culicinae 58, and the subfamily Aedinae 9 genera. Theobald (1907) described 107 species of anophelines under 16 genera. A supplement published in 1910 contained the description of 21 genera and 392 species of Culicidae of the world. The family Anophelinae consisted of 21 genera with 122 species (except the genus *Manguinhosia*).

Table 1: SYSTEMATIC POSITION OF ANOPHELINE MOSQUITOES

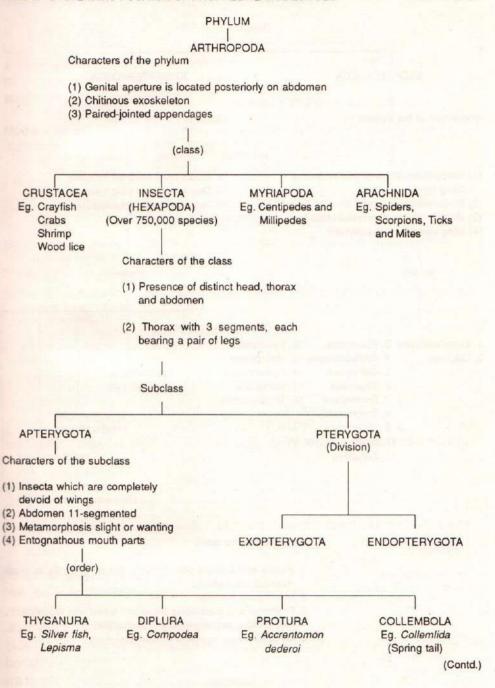


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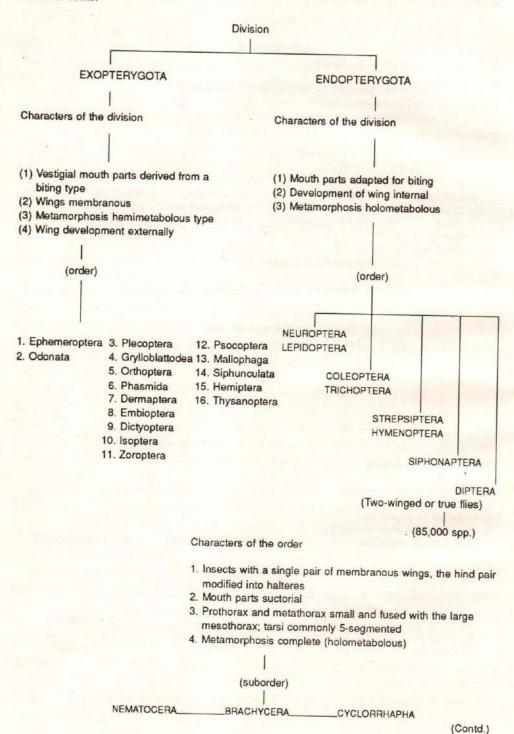


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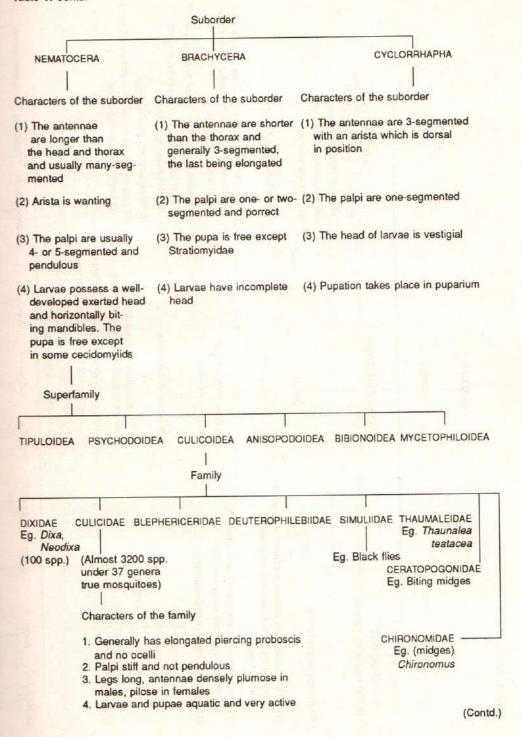
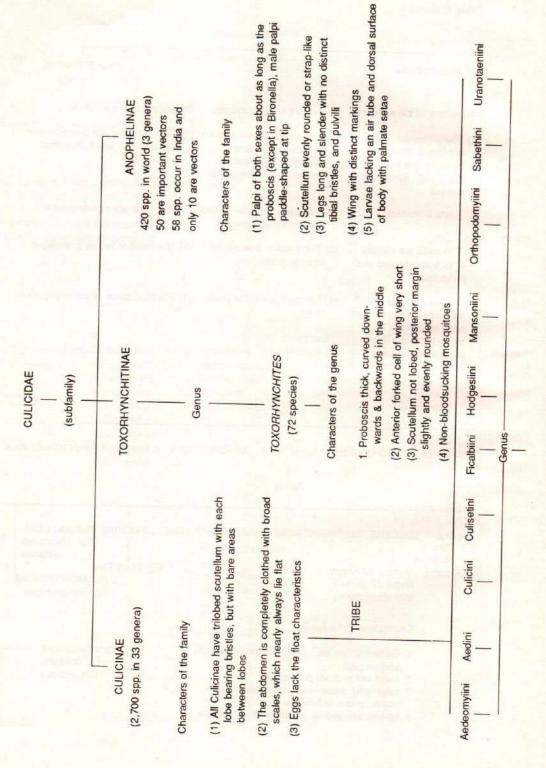


Table 1; Contd.



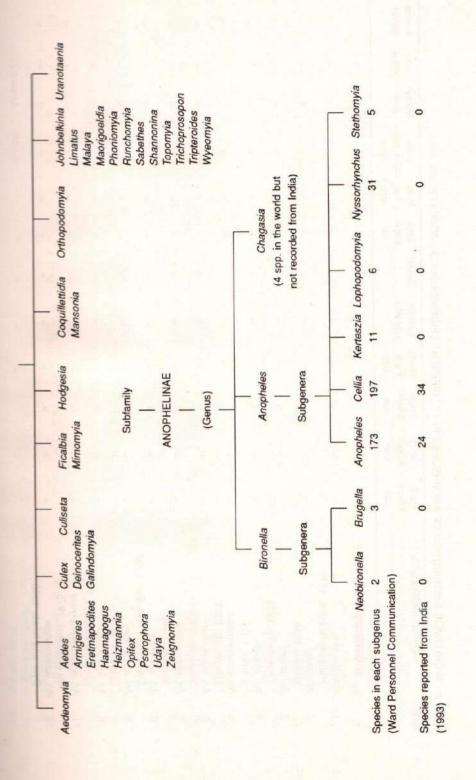


Table 2: WORLDWIDE DISTRIBUTION OF ANOPHELINES IN 12 DIFFERENT EPIDEMIOLOGICAL ZONES OF MALARIA

18.		North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert Afro- tropi- cal	Afro- tropi- cal	Indo- Ira nian	Indo- Chinese Hills	Malay- sian	Chi-	Austra- lasian
	SUBGENUS ANOPHELES						Boyn 188	1					
-	An. aberrans	ı	į	ı	į								
3	Harrison Scanlon, 1975				ı	ı	1	ı	1	+	I	1	1
ci	An. acaci Raicae 1046	Į.	1	1	Ţ	1	ı	1	ı	ı	+	1	1
3	An. ahomi	1	1	ı	, l								
-	Chowdhury, 1929				ı	ı	ı	1	+	+	1	1	1
d'	An. aitkenii James, 1903	ı	ı	1	ſ	ı	1	1	+	+	1	1.	ı
ıo.	An. albotaeniatus (Theobald), 1903	1	ı	1	ı	ı	1	1	1	+	+	1	1
9	An. algeriensis Theobald, 1903	1	1	1	+	+	+	1	+	_1	1	1	1
	An. alongensis Venhuis, 1940	1	ı	1	1	1	ı	1	1	+	ı	- 1	1
8	An. anchietai Correa and Ramalho, 1968	1	ı	+	1	1	1	1	1	ı	1	ı	1
oi	An. annandalei Prashad, 1918	ı	1	ı	1	1	1	1	+	+	+	1	1
10.	An. annulipalpis Lynch Arribalzaga 1878	1	1	+	ı	1	1	r	1	1	1	1	1
1.		ĩ	1	1	1	-1	1	1	ı	1	1	+	1
12.	An. apicimacula Dyar and Knab. 1906	+	+	+	1	1	+	1	1	1	ı	1	- 1

13.	14.	15.	16.	17.	18	19.	20.	21.	22.		23.	24.	25.	26.	
An. arbonicola Zavortink, 1970	An. argyropus (Swellengrebel) 1914		An. atratipes Skuse, 1889	An. atroparvus Van Theill, 1927	An. atropos Dyar and Knab, 1906		An. baezai Gater, 1933	An. baileyi	An. balerensis	Mendoza, 1947	An. bancroftii Giles, 1902	An. barberi	An. barbirostris	An. barbumbrosus	Strickland and Chowdhury, 1927
1	1	1	1	f	+	+	i	1	1		I	+	1	1	
+	.1	1	1	1	+	ı	1	Ī	- 1		1	1	J.	1	2
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1	1	1	1	1	1	1	1	1	1		1	ı	1	-1	
T	1	i	Ů	Î	1	1	1	1	- 1		1	ı	1	- 1	
1	+	1	Ť	1	1	1	1	+	- 1		1	ı	+	+	
1	+	+	1	1	1	+	+	+	1		Ě	1	+	+	
1	+	+	1	1	1	+	+	1	+		ı	ı	+	+	
1	1	1	1	T	1	1	-1	+	ı		1	ı	1	1	
I	1	1	+	1	1	1	1	1			+	1	1	1	

* Zones shown in figure and countries under each zone are given in Table.

Table 2: Contd.

		Ameri- can	Sentral Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
27.	An. barianensis	1	1	1	1								
	James, 1911				+	1	1	1	+	ı	ı	1	1
28.	An. barianensis	ı	1	1	4								
	James, 1911			ı	ł.	1	1	Ĺ	ı	1	1	1	1
29.	An. bengalensis	ı	ı	1	ĵ								
	Puri, 1930				ı	1	ı	l,	+	+	+	+	1
30	An. bonnei	1	ì	4									
	Da Fonseca and		i	+	ı	ı	ı	ı	ı	1	ı	1	1
	Da Silva Ramos, 1939												
31.	An. borneensis	1											
	McArthur, 1949			ı	ı	1	1	1	Ĺ	ı	+	1	1
32.	An. bradleyi	+											
	King, 1939			1	1	1	1	ı	+	į.	1	1	1
33.	An. brevipalpis	ı	-	· ·									
	Roper, 1914			ı	ı	I	ı	1	1	1	+	1	1
34.	An. brevirostris	ı	1										
	Reid, 1950				ı	1	ı	ı	1	ı	+	1	1
35.	An. bulkleyi	ı	,										
	Causey, 1937				1	1	ī	1	ī	+	1	1	1
36.	An. bustamentei	1	1	+	3								
	Galvao, 1955				i	1	ı	i	1	ı	1	1	1
37.	An. calderoni	1	1	+									
	Wilkerson, 1991				1	1	1	ı	1	1	ı	ı	1
38.	An. caliginosus	1	- 1										
	De Meillon, 1943				ı	1	ī	+	1	1	1	ı	ı
39.	An. campestris	1	-1										
	Reid, 1962				ı	1	1	ı	1	+	+	1	1
40	An. chiriquierensis	í	+	1	,	1							
	Komp, 1936						1		1	ı	1	1	1
41.	An. chodukini	1	1	,	+	1	-	,					
	Martini, 1929							ı	ı	ı	1	1	1

42. A	43. A	44. A	45. A	46. A	47. A	48. A	49. A	50. A		51. A	52. A		53. A	54. A		55. 4		56. 4		2/.	
An. claviger	(Meigen), 1804 An. colledgei	Marks, 1956 An. collessi Reid 1963	An. concolor Edwards 1938	An. corethroides Theobald, 1907	An. coustani	An. crawfordi Reid, 1953	An. crucians Wiedemann, 1828	An. cucphuongensis Phan Manh, Hinh and	Vien, 1991	An. culiciformis Cogill, 1903	An. donaldi	Reid, 1962	An. earlei Vargas, 1943	An. eiseni	Coquillett, 1902	An. ejercitoi	Mendoza, 1947		Kanda and Oguma, 1978	An. evandrol Da Costa Lima. 1937	THE WOLKETTE
i	1	1	1	1	1	t	+	1		1	1		+	+		į		I		(Day.
1	1	1	1	i	ı	Î.	+	ı		1	4		ı	+		1		I		1	
1	1	1	1	1	1	1	1	ı		1	1		1	+		t		1		٠	
+	ı	1	ı	1	1	1	1	1		1	1		I.	1		ı		ı			
+	1	1	1	1	+	1	1	1		1	1		ı	1		1		ı		i	
1	1	ı	1	Ĭ.	+	1	1	1		1	1		1 = 5	1		1		ı			
1	1	ı	+	1	+	1		1		1			ı	1		1		ı			
+		1	ĺ	1	1	1	1	1		+	1		ı	1		1		1			
		1		1	1	+	1	+		1	ı		ı	Ĺ		1			1		
1	1	+	1	1	ı	+	ı	ľ		1	+	- 1	ı	i		+	- 1		1		
,	1	1	1	1	ı	r	ı	1		1	ı		ř.	ı		1	+		1		
	+	+	1	+	ı	t	1	1		1	+	1		1		ı	1		1		(Contd.)

58. A 60. A 61. A 63. A 64. A		Ameri-	Ameri-	Ameri-	Eura-	Medite- rranean	Desert	tropi-	lra- nian	Chinese	sian	nese	asian
 		Eg.	, and a	100							-	1	'
र र र र र र	An. fausti	+	1	1	1	1	1	ı	ı	ı			
 	Vargas, 1943										1	ı	1
4 4 4 4	An. fluminensis	1	1	+	1	1	1	1	1	1	8		
* * * * *	Root, 1927									To H	+	1	1
4 4 4 4	An. fragilis	1	1	1	1	I	1	l.	1				
4 4 4 4	(Theobald), 1903										,	- 1	-1
	An. franciscanus	+	1	1	1	1	1	1	ı	ı	ľ.		
	McCracken, 1904										4	1	
	An. franciscoi	1	1	1	1	1	1	1	ı	1			
	Reid, 1962											-	,
	An. freeborni	+	1	1	1	t.	1	1	i	ı	1		
	Aitken, 1939										-	1	
	An. freyi	1	ļ	1	+	t	1	ı	1	1			
	Meng, 1957							-			1		
65. A		1	1	1	Î	1	i	+	1	1			
	Van Someren, 1947								- 41	1	1	1	
66. A	An. gabaldoni	+	+	1	1	ı	1	ı	1				
	Vargas, 1941							1	-	1	1	1	-
67. A	An. georgianus King, 1939	+	1	1	ı	ſ	1				4	1	
68. 4	An. gigas	1	1	1	1	1	1	1	+				
	Giles, 1901						1	1	1	1	1	1	1
69.	Ar. grabhamii	ľ	+	1	1								
	Theobald, 1901								1	1	1	1	
70. 4	An. guarao	1	1	+	1	1	1	1					
	Anduze and												
	Capdevielle, 1949								4	1	1	1	1
71. /	An. habibi	1	1	1	1	1	1						
	Mulligan and Puri, 1936									1	1	1	-1
72. /	An. hectoris	+	+	1	1	1	1	ı					
	Giaquinto-Mira, 1931												

An. helheurale		1 1	1	(Strickland), 1916	71	1		and	raaf), 1919	1	806	. 1	1 +		1	1		1	, 1985	1	1	17	1	Falleroni, 1926	
	1	1						1		1		1	ı			1		1		1	1		+		
1		1						1							1			1		1	1		1		
	1	+	+			1				1		+		- 1		1		+		1	+		1		
	1		1			1						+	1	4		+		1		ı	1		1		9)
	1	1	1	1		1	,			1		ı	ı	1		1		1		1	1		1	1	(Contd.)

Table 2: Contd.

S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
.68	An. lesteri	1	1	1	ı	1	ı	1	1	1	+	+	1
	Baisas and Hu, 1936									7	7	·	1
90	An. letifer	ı	1.	1	1	1	1	ı	ı	+	۲		
	Sandosham, 1944												
91.	An. lewisi	1	1	1	+	1	1	1	ı	1	1	ı	1
	Ludlow, 1920												
92.	An. liangshanensis	1	1	1	1	1	1	1	1	1	ţ	+	Ŀ
	Yeng and Huang, 1984												
93	An. lindesayi	1	1	1	+	1	1	ı	+	+	+	+	1
	Giles, 1900												
76	An maculipennis	1	1	ı	+	+	1	1	+	1	ı	1	1
	Meigen, 1818												
95.	An. maculipes	1	1	+	1	1	1	1	1	1	l.	1	1
	(Theobald), 1903												
96	An. manalangi	1	1	1	1	1	1	1	1	1	+	ı	1
	Mendoza, 1940												
97.	An. marteri	ı	L	1	+	+	+	1	1	1	1	1	1
	Senevet and Prunnelle, 1927												
98	An. martinius	1	Į.	1	+	1	1	1	1	1	1	1	1
	(Shingarev), 1926												
66	An. mattogrossensis	1	1	+	1	1	1	ı	1	1	1	1	1
	Lutz and Neiva, 1911												
100	An. mediopunctatus	1	+	+	1	1	1	1	ı	ı	Ē	1	1
	(Theobald), 1903												
101	An. melanoon	1	1	1	+	+	1	Ĺ	+	ı	1	ı	ı
	Hacket, 1934											3.5	
102.	An. mengalangensis	1	1	1	1	+	1	1	1	Ī	1	+	ı
	Ma, 1981												1
103	An. messeae	1	ı	ı	+	1	+	ı	ı	ı	ĺ	ı	E .
	Falleroni, 1926												

	+																											1		1	
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1		1		1		+		ı		1		ı			1		1		1		1		ı		1		1		1		+
1		1		1		+		1		ľ		ļ			1		1		+		1		t		1		1		1		+
A. minor	Da Costa Lima, 1929	An. montanus	Stanton and Hacker, 1917	An. nambiensis	Coetzee, 1984	An. neomaculipalpus	Curry, 1931	An. nigerrimus	Giles, 1900	An. nilgericus	Christophers, 1924	An. nitidus	Harrison, Scanlon and	Reid, 1973	An. noniae	Reid, 1963	An. obscurus	(Gruenberg), 1905	An. occidentalis	Dyar and Knab, 1906	An. omorii	Sakakibara, 1959	An. palmatus	(Rodenwaldt), 1926	An. paludis	Theobald, 1900	An. papuensis	Dobrotworsky, 1957	An. paraliae	Sndoshom, 1959	An. parapunctipennis
	104.	105.		106.		107.		108		109.		110			111.		112		113.		114		115		116.		117.		118.		119.

Table 2: Contd.

120. An peditaeniatus 121. (Leiosester), 1908 121. Ludlow, 1907 122. An perpasasi 123. An perpasasi 124. An perpasasi 125. An perparasing le 1938 126. An perparasing le 1938 127. An perparasing le 1938 128. An politicaris Barraud, 1932 127. An politicaris Barraud, 1932 127. An politicaris Barraud, 1932 128. An politicaris 129. An powderlix 130. An powderlix 131. An pseudomaculipes 131. An pseudomaculipes 132. An powderlix 133. An pseudomaculipes 133. An pseudomaculipes 14. An pseudomaculipes 15. An powderlix 15. An pseudomaculipes 15. An powderlix 15. An pseudomaculipes 15. An pseudomaculipes 16. An pseudomaculipes 17. An pseudomaculipes 18. An pseudomacul		An. peditaeniatus (Leicester), 1908 An. perplexens Ludlow, 1907 An. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. perytoni		can	can	sian	rranean		tropi- cal	Ira- nian	Chinese	sian	C	nese
An peryassui An peryanani Amerasiophe, 1938 An pertagnani Amerasiophe, 1938 An pinioum Barraud, 1932 An pinioum Sis Barraud, 1932 An powelli An powelli An An pesudomaculipes An pesudomaculipes An pseudopuncipennis + + + + + + + + + + + + + + + + + + +		(Leicester), 1908 An. perplexens Ludlow, 1907 An. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. petroni	1	1	1	1	-					183		
An. perplexens + -		An. perplexens Ludlow, 1907 An. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. pertoni						1	ı	+	+	+	1	
Ludlow, 1907 An. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. petragnani Kulasekera, Harrison &		Ludlow, 1907 An. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. pertoni	+	ı	1	1	1	j						
An peryassui An petragnani Del Vecchio, 1939 An petragnani Del Vecchio, 1939 An petragnani Del Vecchio, 1939 An petragnani An petragnani An petragnani Barraud, 1932 An pinaurensis An pinaurensis An powelli Barraud, 1932 An powelli Ludlow, 1902 An peeudoparbirostris Ludlow, 1908 An pseudopunctipennis		4n. peryassui Dyar and Knab, 1908 An. petragnani Del Vecchio, 1939 An. pertoni							1	1	1	1	1	
An. petragnani An. pe		Dyar and Knab, 1908 4n. petragnani Del Vecchio, 1939 4n. peytoni	Î	1	+	1	- 1							
An. petragnani An. petragnani Del Vecchio, 1939 An. peytoni Kulasekera, Harrison & Amerasinghe, 1988 An. pinjaurensis Barraud, 1932 An. pinjaurensis Barraud, 1932 An. powderi Zavortink, 1970 An. powderi Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis		An petragnani Del Vecchio, 1939 An pevtoni						1	1	ı	ı	ı	1	
An. pseudopunctipennis		Del Vecchio, 1939 4n. pevtoni	1	1	1	1	Н							
An. peytoni Kulasekera, Harrison & Amerasinghe, 1988 An. pilinotum Barraud, 1932 An. pilinotum Barraud, 1932 An. powderi An. powdorink, 1970 An. powdorazulipes Chen 1902 An. pseudobarbirostris Ludlow, 1902 An. pseudopunctipennis		An. pevtoni						ı	1	F	1	ı	1	
Kulasekera, Harrison & Amerasinghe, 1988 An. pilinotum An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis			1	1	1									
Amerasinghe, 1988 An pilinotum Barraud, 1932 An pseudopunctipennis		Kulasekera, Harrison &				ř.	ľ.	ı	1	+	1	ı	1	
An. pilinotum Barraud, 1932 An. pinjaurensis Barraud, 1932 An. plumbeus Stephens, 1828 An. pollicaris Reid, 1962 An. powderi Zavortink, 1970 An. powelli Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudoparaculipes (Peryassu), 1908 An. pseudopunctipennis + + +		Amerasinghe, 1988												
An production is a control of the following section is a control of th		4n. pilinotum	1	1	1									
An pinjaurensis Barraud, 1932 An plumbeus Stephens, 1828 An pollicaris Reid, 1962 An powderi Zavortink, 1970 An pseudobarbirostris Ludlow, 1902 An pseudopunctipennis		Barraud, 1932					1	1	1	1	ı	+	1	
An pseudopunctipennis + + + + + + + + + + + + + + + + + + +		An. pinjaurensis	ı	1	1	1								
An plumbeus Stephens, 1828 An Pollicaris Reid, 1962 An powderi Zavortink, 1970 An pseudobarbirostris Ludlow, 1902 An pseudopunctipennis		Barraud, 1932						ı	ı	+	1	ı	1	
An pseudopunctipennis + + + + + + + + + + + + + + + + + + +		An. plumbeus	1	1	1	+	+	+						
An. pollicaris Reid, 1962 An. powderi Zavortink, 1970 An. powelli Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudopunctipennis		Stephens, 1828								+	1	ı	I	
An. powderi Zavortink, 1970 An. powelli Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis		In. pollicaris	1	1	1	1								
An. powderi Zavortink, 1970 An. powelli Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudomaculipes (Peryassu), 1908 An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis		Reid, 1962					ď.	1	1		+	+	1	
An. powelli Lee, 1944 An. pseudobarbirostris Ludlow, 1902 An. pseudoparculipes (Peryassu), 1908 An. pseudopunctipennis An. pseudopunctipennis An. pseudopunctipennis	V	In. powderi	1	+		ı	1	1						
An. powelli Lee, 1944 An. pseudobarbirostris - Ludlow, 1902 An. pseudomaculipes (Peryassu), 1908 An. pseudopunctipennis + + + +		Zavortink, 1970							i	ı	ı	1	1	
An pseudobarbirostris - Ludlow, 1902 An pseudomaculipes +	A	In. powelli	1	1	1	1								
An pseudobarbirostris Ludlow, 1902 An pseudomaculipes (Peryassu), 1908 An pseudopunctipennis An pseudopunctipennis		Lee, 1944						1	1		1	1	1	
An. pseudomaculipes		in. pseudobarbirostris	1	1	-									
An pseudomaculipes – – + – – – (Peryassu), 1908 An pseudopunctipennis + + + – – – – – – – – – – – – – – – –		Ludlow, 1902				1)	1	1	1	1	+	+	1	
(Peryassu), 1908 An. pseudopunctipennis + + + +		n. pseudomaculipes	1	1	+	1			1					
An pseudopunctipennis + + + +		(Peryassu), 1908								1	1	1	1	
		n. pseudopunctipennis	+	+	+	1	-							

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sis	nticus 1957			1906				tus											-		am, 1946		8				
dosinens	dostigme	S	Yamada, 1937 n. punctimacula	Dyar and Knab, 1	tipennis 1823	ati	Laveran, 1902	frimacula	824	inc	1, 1952		Harrison, 1973	1	950	arovi	1903	rensis	Rozeboom, 1951	roi	Bohart and Ingram,	ratus	(Leicester), 1908	noni	1931	ılti	Rouband, 1935
An. pseudosinensis	An. pseudostigmaticus Dobrotworsky 1957	An. pullus	Yamada, 1937 An. punctimacula	Dyar a	An. punctipennis (Say), 1823	An. pursati	Lavera	An. quadrimaculatus	Say, 1824	An. rachour	Galvao, 1952	An. reidi	Harrisc	An. roperi	Reid, 1950	An. sacharovi	Favre, 1903	An. samarensis	Rozebo	An. saperoi	Bohart	An. separatus	(Leices	An. shannoni	Davis, 1931	An. sicaulti	Roubai
134.	135.	136.	137.		138.	139.		140		141		142.		143.		144		145.		146.		147.		148.		149.	

Table 2: Contd.

150. An similissimus Strickland and Chowdhury, 1927	S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi-	Austral- asian
Strickland and Chowdhury, 1927 An. sinensis Wiedemann, 1828 An. sinenoides An. sintonoides An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1988 An. stigmaticus Skuse, 1966 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis An. tasmaniensis Cheebald, 1902 An. tasmaniensis An. tasmaniensis An. tasmaniensis Cheebald, 1903 An. tasmaniensis An. tasmani	150.	An. similissimus	1	1	1	1	.1	1	1	T	1	+	1	.1
An. sinensis Wiedemann, 1828 An. sineroides Yamada, 1924 An. sintoni Puri, 1929 An. sintonoides Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Dobrotworsky, 1966 An. tibiamaculatus (Neiva), 1906 An. tibiamaculatus (Neiva), 1908 An. vanus Walker, 1859		Strickland and Chowdhury 1927												
Wiedemann, 1828 An. sineroides Yamada, 1924 An. sintoni Puri, 1929 An. sintonoides Ho, 1938 An. subalpinus Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. subalpinus An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1889 An. stigmaticus An. stigmaticus Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tibiamaculatus (Neival, 1906 An. tibiamaculatus (Neival, 1906 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	151	An. sinensis	1	1	1	+	1	1	ı	+	+	+	+	ı
An. sineroides Yamada, 1924 An. sintoni Puri, 1929 An. sintonoides Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1928 An. stigmaticus Dobrotworsky, 1966 An. tenebrosus Dobrotworsky, 1966 An. tenebrosus Dobrotworsky, 1966 An. tibiamaculatus (Neiva), 1906 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Wiedemann, 1828												
Yamada, 1924 An. sintoni Puri, 1929 An. sintonoides Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1889 An. stigmaticus Skuse, 1889 An. stigmaticus Edwards, 1928 An. tameniensis Dobrotworsky, 1966 An. tamenosus Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tibiamaculatus (Neiva), 1906 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	52.	An. sineroides	1	1	J	1	1	1	1	1	1	1	+	1
An. sintoni Puri, 1929 An. sintonoides An. subalpinus Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tibiamaculatus (Neiva), 1908 An. vanus		Yamada, 1924												
An. sintonoides An. sintonoides An. subalpinus An. subalpinus An. sigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tibiamaculatus Coenitz, 1906 An. tibiamaculatus (Neiva), 1908 An. vanus Walker, 1859	53	An. sintoni	1	1	1	1	1	1	1	+	i	1	1	1
An. sintonoides Ho, 1938 An. subalpinus Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis An. tasmaculatus (Neiva), 1906 An. tibiamaculatus (Neiva), 1906 An. tumbrosus (Theobald), 1903 An. vanus Walker, 1859		Puri, 1929												
Ho, 1938 An. subalpinus — Hackett and Lewis, 1935 — An. stigmaticus — Skuse, 1889 — An. stricklandi — Reid, 1965 — An. stricklandi — Reid, 1965 — An. tasmaniensis — Dobrotworsky, 1966 — An. tasmaniensis — Doenitz, 1902 — An. tibiamaculatus — (Neiva), 1906 — An. tibiamaculatus — (Neiva), 1906 — An. tibiamaculatus — (Neiva), 1906 — An. tibertti — Scanlon and Peyton, 1967 — An. umbrosus — (Theobald), 1903 — An. vanus — Walker, 1859 —	54.	An. sintonoides	1	1)	1	1	1	1	1	+	+	+	1
An. subalpinus An. subalpinus An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Doenitz, 1902 An. thiamaculatus (Neiva), 1906 An. tigarnia and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Ho, 1938												
Hackett and Lewis, 1935 An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Doenitz, 1902 An. tiniamaculatus (Neiva), 1906 An. tipiamaculatus (Neiva), 1906 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	55.	An. subalpinus	1	1	1	+	+	1	1	1	1	1	1	1
An. stigmaticus Skuse, 1889 An. stricklandi Reid, 1965 An. symesi Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tasmaniensis Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scandan and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Hackett and Lewis, 1935												
Skuse, 1889 An. stricklandi Reid, 1965 An. symesi Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tanebrosus Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scand and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	.99	An. stigmaticus	1	1	1	1	1	1	1	1	1	+	1	+
An. stricklandi Reid, 1965 An. symesi Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tanebrosus Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scandon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Skuse, 1889												
An. symesi Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tenebrosus Doenitz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	57.	An. stricklandi	ı	1	1	1	1	1	1	1	+	+	1	1
An. symesi Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tenebrosus Doeniz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Reid, 1965												
Edwards, 1928 An. tasmaniensis Dobrotworsky, 1966 An. tenebrosus Openiz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	58.	An. symesi	ı	ı	1	1	-1	1	+	1	1	1	ı	1
An. tasmaniensis Dobrotworsky, 1966 An. tenebrosus Doeniz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Edwards, 1928												
Dobrotworsky, 1966 An. tenebrosus Doeniz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	59.	An. tasmaniensis	ı	1	1	1	1.	1	1	1	ı	1	1	+
An. tenebrosus Doeniz, 1902 An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Dobrotworsky, 1966												
An tibiamaculatus (Neiva), 1906 An tigertti Scanlon and Peyton, 1967 An umbrosus (Theobald), 1903 An vanus Walker, 1859	.09	An. tenebrosus	1	į	1	1	1	+	+	1	1	1	1	1
An. tibiamaculatus (Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		Doenitz, 1902												
(Neiva), 1906 An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	61.	An. tibiamaculatus	1	1	+	1	1	1	1	1	ı	1	1	ī
An. tigertti Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859		(Neiva), 1906												
Scanlon and Peyton, 1967 An. umbrosus (Theobald), 1903 An. vanus Walker, 1859	62.	An. tigertti	1	1	1	1	1	1	1	1	+	1	1	1
An. umbrosus (Theobald), 1903 An. vanus – – – Walker, 1859		Scanlon and Peyton, 1967												
(Theobald), 1903 An. vanus Walker, 1859	63	An. umbrosus	1	1	1	1	1	1	1	+	+	+	ı	1
An. vanus — — — — — — — — — — — — Walker, 1859		(Theobald), 1903												
Walker, 1859	64	An. vanus	1	1	1	1	1	1	1	1	1	+	1	1
		Walker, 1859												

Table 2: Contd.

S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
180	An. arabiensis	1	1	1	1	1	+	+	,				-
	(Patton), 1905									(la la	1	ı
181	An. ardensis	1	1	1	1	1	1	+	ı	- 1	1	1	
	(Theobald), 1905											,	1
182.	¥	1	1	1	1	1	1	+	ı	- 1		1	
	(Gough), 1910											1	1
183	An. amoulti	1	ı	ı	1	1	1	4					
	Grjebine, 1966									ı	1	1	1
184	An. aruni	1	1	-1	ļ	-	1	4					
	Sobti, 1968									ı	ı	ı	1
185.	An. aurirostris	1	1	1	ı	1		į.			100		
	(Watson), 1910									ı	+	1	ı
186.	An. austenii	1	1	-	1								
	(Theobald), 1905						1	+	1	ı	1	1	1
187	An. azaniae	1	ı	1	1	,		7	9				
	Bailly-Choumara, 1960								1	1	1	ı	1
188	An. azevedoi	ı	1	1	1	1	1	+					
	Ribeiro, 1969								1	1	1	ı	1
189.	An. balabacensis	1	1	1	1	1	,		+		3		
	Baisas, 1936									+	+	1	1
190	An. barberellus	1	1	1	1	1	+	+	1	,			
	Evans, 1932											1	1
191.	An. berghei	1	1	1	1	,	1	4	-				
	Vincke and Leleup, 1949											1	1
192	An. bervoetsi	1	1	1	1	1	1	4	J				
	D'Haenans, 1961				V.	ļķ.	F	F	ı	ı	ı	1	1
193.	An. brohieri	1	1	-1	ı	,		+					
	Edwards, 1929								1	ı		1	L
194.	An. brucei	1	1	1	j	1	1	,					
	Service, 1960							+	1	1	1	1	1

	An. brumpti Hamon and Rickenbach, 1955				1	1	1	1	+	+	1 1 1 1 1	1 1 1 1 1	1 1 1 1 1
196.	An. brunnipes (Theobald), 1910	1		1		1	1	1	+ 1 1	+ 1 1	1 1 1	1 1 1 1 1	1 1 1 1 1 1
197.	An. buxtoni	1		ı		1	1	1	+ 1 1 1 1	+ 1 1 1 1	1 + 1 1 1 1 1	1 + 1 1 1	1 + 1 1
198.	An. bwambae White, 1985	t	17.1	1	l I		ı	l l	+ 1 1	1. 1. 1.	+ 1 1	1 + 1 - 1 - 1 - 1	1 + 1 + 1 + 1 + 1 + 1 + 1 + 1 + 1 + 1 +
199.	An. cameroni De Meillon and Evans, 1935		1			ı	1	1	1	1	1 1 1	1 + 1 - 1	1 + 1 - 1
200.			1			i	1	1	1 1 1	1 1 1	1 1 1	1 + 1	1 1 1 1 1 1
	Adam, 1961												
201.	An. carteri		1		1		ī	1	+	+	+ 1	1 + 1	1 + 1
200	Evans and De Meillon, 1933				1			1	+	+	1	1 +	1 +
	Mountained and Carter 1011		J										
203	(Newstead and carter), 1911		1		1		ı	1	-1	-1	1 1	1 +	1 + 1
	M. cincius		1		ı		ı	1					
	(Newstead and Carter), 1910								4	4	+	1 4	1 1
204.	An. cinereus		1		1		1	1	+ +	+ +	+ +	1 + + 1	1 + + 1
	Theobald, 1901												
205.	An. clowi		1		1		1	1	1	1	1 1	1 1 1 1	1 1 1 1 1
	Rozeboom and Knight, 1946												
506.			ı		ı		1	1	+	+	1 + 1	1 + 1 1 1	1 + 1
	Evans and Lesson, 1935												
207.	An. cordurieri		1		1		1	L	+	+	1 + 1	1 + 1 1	1 1 + 1
	Grjebine, 1966												
208.	An. cristatus		1		1		1	1	1	1	1 1	1 1 1	+ 1 1 1 1 1
	King and Baisas, 1936												
209.	An. cristipalpis		1		1		1	1	+	+	1 + 1	1 + 1	1 + 1
	Service, 1977												
210.	An. culicifacies		1		1		+	+	+ + + +	+ + + +	+ + + + +	+ + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
	Giles, 1901												
211.	An. cydippis		1		1		1	1	+	+	1 + 1	1 + 1 1	1 + 1 1
	De Meillon 1931												

Table 2: Contd.

212. An dencalicus Coluzzi 1958 213. An dencalicus Coluzzi 1958 214. An dennigion 215. An dennigion 216. An dennigion 217. An dennigion 218. An dennigion 219. An dennigion 220. An dennigion 22	S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi-	Austral- asian
An derized til. 1939 An charactetti, 1939 An derized til. 1939	212.	An. dancalicus	Ī	1	1	1	1	1	+	1	1	J	1	1
An deaming Service, 1958 An deaming Service, 1970 Service, 1970 An demingioni Evans, 1933 An dirus An disparate and Carter), 1911 An official service		Corradetti, 1939										92		
Coluzzi 1958 An deemingi Service, 1970 An deminginal Evans, 1939 An dermeliani An dispicus An dispicus An dispicus An erepens Gillies and Coetzee, 1987 An deminoid Button, 1939 An deminoid Button, 1939 An erepens Gillies and Coetzee, 1987 An deminoid Button, 1958 An erepens Gillies and Coetzee, 1987 An elephony Gillies and Coetzee, 1987	213.	An. daudi ·	1	1	1	1	1	1	+	1	1	1	1	1
An deemingi An deemingi Service, 1970 An derivalioni Evans, 1933 An dispar Ratinanthikul and Harbach, 1991 An dispar An dispar An dispar An dispar Christophers, 1924 An dureni Edwards, 1938 An elegans An elegans An elegans An erpens Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus		Coluzzi, 1958												
An derival in the service in 1970 An derival in the service in 1970 An direction and Harrison, 1979 An displactus An eigens An eigens An eigens An eigens Gillies and Coertee, 1987 An ethiopicus Gillies and Coertee, 1987 An ethiopicus Gillies and Coertee, 1987	214.	An. deemingi	1	1	1	1	1	1	+	1	1	ı	1	1
An demeillon! Evans, 1933 An diras Peyton and Harrison, 1979 An dispar Rattanatritikul and Harbach, 1991 An disportant and Carter), 1911 An disportant and Carter), 1911 An disportant and Coetzee, 1991 An disportant and Carter), 1911 An disportant and Coetzee, 1987 An ethiopicus Contracts, 1933 An erpens Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus Cologia and Coetzee, 1987		Service, 1970												
Evans, 1933 An dirus An dirus Patron and Harrison, 1979 An dispar Rattanarithikul and Harbach, 1991 An dispara Control of Cortoe, 1911 An dispara Control of Cortoe, 1938 An erepens Control of Cortoe, 1987 An ethic picus Coltoe, 1987	215.	An. demeilloni	1	1	1	1	ı	1	+	1	1	1	1	
An dirus An dirus An distinct and Harrison, 1979 An distinctus (Newstead and Carter), 1911 An distinctus Christophers, 1924 An diveni Edwards, 1938 An diveni Edwards, 1938 An erepens Cortadett, 1939 An erepens Gillies and Coetzee, 1987 An ethickus Gillies and Coetzee, 1987 An ethickus Gillies and Coetzee, 1987 An ethickus Gillies and Coetzee, 1987		Evans, 1933												
An displanative land Harbach, 1979 An displanative land Harbach, 1991 An displanative land Carter), 1911 An displanative land Carter), 1911 An domicola (Newstead and Carter), 1911 An domicola (An displanative land) An displanative land An elegans Corradetti, 1939 An erepens Coloccee, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987	216.	An. dirus	1	1	1	1	1	ı	1	1	+	-1	1	1
An dispar An dispar An dispar An dispar An disparathikul and Harbach, 1991 An distinctus (Newtodicus) Edwards, 1916 An derviedicus Christophers, 1924 An durani Edwards, 1938 An elegans Corradetti, 1939 An erspens Gillies and Coetzee, 1987 An elispicus Gillies and Coetzee, 1987 An elispicus Corradetes, 1987		Peyton and Harrison, 1979												
An distinctus An distinctus (Newstead and Carter), 1911 An distinctus (Newstead and Carter), 1911 An distinctus Christophers, 1924 An distinctus Christophers, 1924 An distinctus Christophers, 1938 An distinctus Christophers, 1938 An elegans Corradetti, 1939 An erepens Colares, 1987 An ethiopicus Gillies and Coetzee, 1987 An ethiopicus Gillies and Coetzee, 1987	217.	An. dispar	J	1	1	1	ı	1	1	1	1	+	1	1
An distinctus (Newstead and Carter), 1911 An domicola Edwards, 1916 An diversity Patton, 1905 An elepens Corradetti, 1939 An ethiopicus Gillies and Coetzee, 1987 An distinctus (Newstead and Carter), 1911		ithikul and												
An. domicola Edwards, 1916 An. draviedicus Christophers, 1924 An. draviedicus Christophers, 1924 An. drahali Paton, 1905 An. dureni Edwards, 1938 An. elegans (James), 1903 An. erepens Corradetti, 1939 An. erepens Gliles and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987	218.		1	ı	1	1	1	-1	+	ı	ı	ı	1	1
An. domicola Edwards, 1916 An. draviedicus Christophers, 1924 An. draviedicus Corradetti, 1938 An. ergens Gillies and Coetzee, 1987 An. demicola Edwards, 1938 An. ergens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		(Newstead and Carter), 1911												
Edwards, 1916 An draviedicus Christophers, 1924 An drhali Patton, 1905 An elegans Coradetti, 1939 An erlipopicus Gillies and Coetzee, 1987 An devende, 1987 An devende, 1987 An elegans Gillies and Coetzee, 1987 An elegans Coradetti, 1939 An ethiopicus Gillies and Coetzee, 1987	219.	An. domicola	1	1	Ī	1	1	1	+	1	1	1	1	ı
An. draviedicus Christophers, 1924 An. dureni Fatton, 1905 An. everyeans Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987 An. draviedicus Gillies and Coetzee, 1987		Edwards, 1916												
Christophers, 1924 An. dthali An. dthali Patton, 1905 An. dureni Edwards, 1938 An. elegans + + + + + +	220.	An. draviedicus	1	1	1	1	1	1	1	1	+	+	1	ı
An. dthali Patton, 1905 An. dureni Edwards, 1938 An. elegans (James), 1903 An. erythraeus Corradetti, 1939 An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987 Gillies and Coetzee, 1987		Christophers, 1924									e:	20		
Patton, 1905 An. dureni Edwards, 1938 An. elegans + + +	221.	An. dthali	1	1	1	1	1	+	+	+	1	ı	1	1
An. dureni Edwards, 1938 An. elegans (James), 1903 An. erythraeus Corradetti, 1939 An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		Patton, 1905												
Edwards, 1938 An. elegans (James), 1903 An. erythraeus Corradetti, 1939 An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987	222.	An. dureni	1	1	1	1	1	1	+	1	1	1	1	į
An. elegans (James), 1903 An. erythraeus Coradetti, 1939 An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		Edwards, 1938												
(James), 1903 An. erythraeus +	223.	An. elegans	1	1	1	1	1	1	1	+	1	1	1	ı
An. erythraeus Coradetti, 1939 An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		(James), 1903		13										
Corradetti, 1939 -	224.	An. erythraeus	1	1	1	1	1	1	+	1	ı	,	ı	ı
An. erepens Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		Corradetti, 1939												
Gillies and Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987	225.	An. erepens	1	1	1	1	1	1	+	ı		1	1	1
Coetzee, 1987 An. ethiopicus Gillies and Coetzee, 1987		Gillies and												
An ethiopicus + + Gillies and Coetzee, 1987		Coetzee, 1987												
1987	226.		1	1	1	1	1	1	+	1	3	1	1	1
		Coetzee,												

177	An rain												
	Leleup, 1952												
228.	An. farauti	1	1	1	1	ı	1	ļ	1	ı	+	1	
	Laveran, 1902												
229.	An. filipinae	ī	ì	1	1	1	1	Î	1	+	+	i.	
	Manalang, 1930												
230.	An. flavicosta	1	1	1	1	1	1	+	1	1	1	t	
	Edwards, 1911												
231.	An. flavirostris	1	1	1	1	1	ı	1	1	1	+	1	
	(Ludlow), 1914												
232.	An. fluviatilis	1	1	1	+	1	+	1	+	+	+	+	
	James, 1902												
233.	An. fontinalis	1	1	1	1	1	1	+	1	1	ı	1	
	Gillies and De Meillon, 1968												
234.	An. freetownensis	ı	1	1	Í	1	1	+	1	1	1	1	
	Evans, 1925												
235.	An. funestus	1	1	1	1	1	1	+	1	1	ı	1	
	Giles, 1900												
236.	An. fuscivenosus	Į	ī	1	1.	1	1	+	I	1	1	ı	
	Leeson, 1930												
237.	An. gambiae	1	1	1	1	1	1	+	1	1	ľ	1	
	Giles, 1902												
238.	An. garnhami	1	1	1	1	1	1	+	1	1_	ı	ı	
	Edwards, 1930												
239.	An. gibbinsi	1	1	1	1	1	1	+	Ĭ	Î	ı	1	
	Evans, 1935												
240.	An. grassei	1	1	1	1	1	1	+	1	1	1	I	
	Grjebine, 1953												
241.	An. grenieri	1	1	1	1	1	1	+	1	ı	ı	ı	
	Grjebine, 1964												
242.	An. greeni	1	1	1	1	1	1	1	1	ı	+	1	
	Rattanarithikul and												
	Harbach, 1991							*		Daniel Control			

Table 2: Contd.

S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
243.	An. griveaudi	1	1										
	Grjebine, 1960			ı.	ı	1	1	+	1	1	1	1	1
244.	An. hackeri	1											
	Edwards, 1921		I,	ı	ŀ	ı	1	1	1	+	+	1	1
245.	An. hamoni												
	Adam, 1962		ı	1	1	1	1	+	1	t	1	1	1
246.	An. hancocki	1											
	Edwards, 1929	ı	ľ	1	1	1	1	+	1	ı	1	1	ı
247.	An. hargreavesi												
	Evans, 1927	ı	ı	1	1.	1	+	+	1	1	1	1	1
248.	An. harperi												
	Evans, 1936	ı	1	1	ı	1	1	+	1	1	1	1	1
249.	An. hilly												
	Woodhill and Lee, 1944	ı	i	ı	1	ī	1	1	I	1	1	ı	+
250.	An. hughi												
	Lambert and Coetzee, 1982		ı	1	1	1	ı	+	1	1	ı	1	1
251.	An. incognitus												
	Brug, 1931			,	1	ı	1	ı	1	1	1	1	+
252.	An. indefinitus	1	3)										
	(Ludlow), 1904			1	F	į.	1	1	1	+	+	+	1
253.	An. introlatus	1											
	Colless, 1957			ı	1	1	1	1	1	L	+	1	1
254.	An. jamesii	- ()											
	Theobald, 1901		ı	ľ.	1	1	1	1	+	+	+	+	1
255.	An. jebudensis	1											
	Froud, 1944		ı	1	ı	1	1	+	1	1	1	1	1
256.	An. jeyporiensis	1											
	James, 1902			1	ı	1	1	L	+	+	+	1	1
257.	An. karwari	1											
	(James), 1902			ı	1	1	F	1	+	+	+	1	+

258.		259.		260.		261.		262.		263.			264.		265.		266.		267.		268.		269.		270.		271.		272.		273.		274.	TO THE
An. Keniensis	Evans, 1931	An. kingi	Christophers, 1923	An. kochi	Doenitz, 1901	An. kolambuganensis	Baisas, 1932	An. koliensis	Owen, 1945	An. kosiensis	Coetzee, Segerman	and Hunt, 1987	An. Iacani	Grjebine, 1953	An. latabensis	Lambert and Coetzee, 1982	An. leesoni	Evans, 1931	An. Ieucosphyrus	Doenitz, 1901	An. limosus	King, 1932	An. listeri	De Meillon, 1931	An. litoralis	King, 1932	An. Iloreti	Gil Collado, 1936	An. longipalpis	(Theobald), 1903	An. longirostris	Brug, 1928		Gillies and Coetzee, 1987
- 1		1		1		1		1		1			1		1		1		1		1		1		1		I		1		1		1	
1		1		1		1		1		1			1		1		1		1		1		1		1		1		1		1		-	
1		1		1		1		1		1			1		1		1		1		1		1		ı		1		1		1	1000	1	
1		1		,		1		1		ı			1		1		1		1		1		1		1		1		1		1		1	
1		1		1		1		1		1			1		1		1		1		1		1		1		1		1		1		1	
1		1		1		1		1		1			1		1		1																	
+		+		+						+			+		+		+																+	
1				+																													1	
1																																	1	
1		1		1		1		ı		ı			ı		1		i		1		1		1		1		1		1		ı		1	
1		-1		+		ı		+		+			1		1		ı		1		1		1		1		1		I		+	4	1	

Table 2: Contd.

275. An lovestae Evans, 1934 276. An ludiowae 277. An ludiowae 278. An macarburi 278. An macarburi 279. An macarburi 279. An macarburi 270. An macarburi 270. An macarburi 270. An macarburi 270. An macarburi 271. An machigalpis 270. An macarburi 270. An macarburi 271. An machigalpis 270. An machigalpis 270. An machigalpis 270. An machigalpis 281. An machigalpis 282. An majori 383. An majori 384. An machigalpis 385. An macarburi 386. An macarburi 387. An macarburi 388. An macarburi 388. An macarburi 389. An macarburi 389. An macarburi 380. An macarburi 380. An macarburi 381. An macarburi 382. An macarburi 383. An macarburi 384. An macarburi 385. An macarburi 386. An macarburi 387. An macarburi 388. An macarburi 389. An macar	S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
Evans, 1934 An Iuligae An Iulig	275.	An. loyettae	1	1	1	í	1	1	1	1	1	+		-
An Iudiowae (Theobaid), 1903 An macarthuri Colless, 1956 An maculipalpis Glies, 1902 An malighis Bailly-Choumara and Adam, 1959 An marajoara (Galvao and Lane), 1942 An mascarensis De Meillon, 1947 An meraukensis Theobaid, 1903		Evans, 1934												
An. marajoara An. marajoara An. marajoara An. marajoara An. marajoara (Theobald, 1903 An. marajoara (Theobald, 1903 An. marajoara An. marajoara (Theobald, 1903 An. marajoara An. marajoara (Theobald, 1903 An. marajoara (Theobald, 1903 An. marajoara An. marajoara An. marajoara An. marajoara (Theobald, 1903 An. marajoara	.92	An. Iudiowae	1	1	1	1	ı	1	1	-		4		
An. Iungae Belkin and Schlosser, 1944 An. macarthuri Colless, 1956 An. maculatus Theobald, 1901 An. marajoana An. marajoana (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. mascarensis An. melas Theobald, 1903 An. mascarensis Venhuis, 1932 An. melas Theobald, 1903 An. melas Theobald, 1903 An. melas Theobald, 1903 An. melas Theobald, 1903 An. melas		(Theobald), 1903								1	ı	+	ı	1
Belkin and Schlosser, 1944 An. macarthuri Colless, 1956 An. maculipalpis Galve, 1902 An. margioara (Galve), 1906 An. margioara (Galve), 1903 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903	77.	An. lungae	1	ı	Ī	,	1	9						
An. macarthuri Colless, 1956 An. maculatus Theobald, 1901 An. malarsolara (Galvao and Lane), 1905 An. marakensis De Meillon, 1903 An. marakensis Volung and Meillon, 1903 An. marakensis An. marakensis De Meillon, 1903 An. marakensis Venhuis, 1903 An. marakensis Venhuis, 1903 An. marakensis Venhuis, 1903 An. marakensis Venhuis, 1903		Belkin and Schlosser, 1944						-	1	ı	ı	+	ı	1
Colless, 1956 An. machardyi Edwards, 1930 An. macaulatus Theobald, 1901 An. marajoara (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. meraukensis Vendus, 1932 An. maraukensis Vendus, 1932 An. macaukensis An. macaukensis An. macaukensis An. meraukensis Vendus, 1932 An. macaukensis Vendus, 1932 An. macaukensis Vendus, 1932	78.	An. macarthuri	1	ı	1	-)	4		100					
An macrandyi Edwards, 1930 An macralatus Theobald, 1901 An marayanus (Galvao and Lane), 1942 An marayanus An marayan		Colless, 1956					ı	ı	1	1	ı	+	1	1
Edwards, 1930 An. maculatus Theobald, 1901 An. maculands An. meraukensis De Meillon, 1932 An. meraukensis Theobald, 1903	.62	An. machardyi	1	1	I	ı	1	â	+					
An. maculatus Theobald, 1901 An. maculpalpis Giles, 1902 An. majidi Young and Majid, 1928 An. majidi Young and Majid, 1928 An. mailensis Bailly-Choumara and Adam, 1959 An. marajoara (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. mescarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis Venhuis, 1932		Edwards, 1930						1	+	ı	ı	ı	ı	1
An meraukensis Theobald, 1901 An maculipalpis Giles, 1902 An maculipalpis Giles, 1902 An maculipalpis Giles, 1902 An maculipalpis Giles, 1903 An maculipalpis Giles, 1903 An maculipalpis Theobald, 1903 An meraukensis Theobald, 1903	30.	An. maculatus	1	1	-	1	3					14		
An. maculpalpis An. meraukensis		Theobald, 1901						1	1	+	+	+	ı	1
Giles, 1902 An. majidi Young and Majid, 1928 An. maliensis Bailly-Choumara and Adam, 1959 An. manayanus (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. meraukensis Theobald, 1903 An. meraukensis An. meraukensis Venhuis, 1932	-	An. maculipalpis	1	1					-					
An. majidi Young and Majid, 1928 An. maliensis Bailly-Choumara and Adam, 1959 An. manayanus (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. marsarensis De Meillon, 1947 An. meraukensis Theobald, 1903 An. meraukensis		Giles, 1902				1	1	ı	+	ı	1	1	ı	1
Young and Majid, 1928 40. maliensis An. maliensis Bailly-Choumara and Adam, 1959 - + + - - + An. marajoara (Banks), 1906 -	2	An. majidi	1	J	1									
An. maliensis Bailly-Choumara and Adam, 1959 An. mangyanus (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. marsarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis An. meraukensis Theobald, 1903 An. meraukensis		Young and Majid, 1928						ľ	ı	+	+	ı	1	1
Bailly-Choumara and Adam, 1959 An. manayanus (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis An. meraukensis Theobald, 1903 An. meraukensis	3	An. maliensis	1	1	1		-							
An. mangyanus (Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis An. meraukensis Venhuis, 1932		Bailly-Choumara and Adam, 1959						ľ.		ı	ı	1	1	1
(Banks), 1906 An. marajoara (Galvao and Lane), 1942 An. marshallii (Theobald), 1903 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis An. meraukensis Venhuis, 1932	4	An. mangyanus	1	1	1	1	1	1	-	1	3	-		
An. marajoara (Galvao and Lane), 1942 An. marshallii (Theobald), 1903 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis Venhuis, 1932		(Banks), 1906									ı	+	1	1
(Galvao and Lane), 1942 An. marshallii (Theobald), 1903 An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis Venhuis, 1932	5.	An. marajoara	1	1	+	1	,	1	1	1				
An. marshallii		Lane),									1		ı	1
An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis Venhuis, 1932	9	An. marshallii	1	ı	1	1		-	+					
An. mascarensis De Meillon, 1947 An. melas Theobald, 1903 An. meraukensis Venhuis, 1932		(Theobald), 1903									ı	1	ı	1
An. melas Theobald, 1903 An. meraukensis Venhuis, 1932	7.	An. mascarensis	1	ı	,		-		4					
An. melas Theobald, 1903 An. meraukensis Venhuis, 1932		De Meillon, 1947						1	+	1	ı	1	ı	ĺ
Theobald, 1903 An. meraukensis Venhuis, 1932	8	An. melas	1	1	1	ì	1		+					
An. meraukensis – – – – – – – – – – – – – – – – – –		Theobald, 1903								1		1	ı	ı
	6	An. meraukensis	1	1	1	1	d	-	.1	- 1		-		130
		Venhuis, 1932						1		1	ı	+	ı	+

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An. merus	Doenitz, 1902 An. milloti	Grjebine and Lacan, 1953	An. minimus Theobald 1901	An. moghulensis	Christophers, 1924	An. mortiauxi	Edwards, 1938	An. moucheti	Evans, 1925	An. mousinhoi	De Meillon and Pereira, 1940	An. multicolor	Cambouliu, 1902	An. murphyi	Gillies and De Meillon, 1968	An. natalensis	(Hill and Haydon), 1907	An. nataliae	Belkin, 1945	An. negros	Colless, 1956	An. nemophilous	Peyton and Ramalingam, 1988	An. nili	(Theobald), 1904	An. nivipes	(Theobald), 1903	An. njombiensis	Peters, 1955
290.	291.		292.	293.		294.		295.		296.		297.		298.		299.		300		301		305.		303		304		305.	

Table 2: Contd.

306. An. notanandai Rattanrithikul and Green, An. notleyi Van Someran, 1949 308. An. novaguinensis Venhuis, 1933 309. An. pallidus Theobald, 1901 310. An. paltrinierii Shidrawi and Gillies, 1987 311. An. parangensis (Ludlow), 1914 313. An. parangensis (Ludlow), 1914 314. An. pattoni Christophers, 1926 315. An. pauliani Griebine, 1953 316. An. pharoensis Theobald, 1901 317. An. philippinensis Ludlow, 1902 318. An. pretoriensis (Theobald), 1903 319. An. pretoriensis (Theobald), 1903				-	2	rranean		-Idon	ra-	Chinese	SIGN	BSBU	asian
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र र र र र र र र र र र र र	1 200	1	1	1	1	1	1	1	1	1	+	1	1
 	l and Green, 1986												
र र र र र र र र र र र र		1	1	1	1	1	1	+	1	1	1	1	1
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 	nsis)	1	1	1	1	1	ı	1	1	ı	ı	+
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र र र र र र र र र र		1	1	1	1	1	1	ı	+	+	+	1	1
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य य य य य य य य	Buettiker and Beales, 1959												
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र र र र र र													
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र र र र र	, 1926												
र र र र		1	1	1	1	1	1	+	1	1	1	1	1
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4	903												
	esi	1	1	1	1	1	1	+	1	ı	1	ı	+
Strickland and	Strickland and Chowdhury, 1927						2						
320. An. pseudowillmori	nori	1	E	1	ī	1	1	1	+	+	+	4	1
(Theobald), 1910	910												

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Table 2: Contd.

Sawadwongpomi	S.No.	GENUS ANOPHELES	North Ameri- can	Central Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi- nese	Austral- asian
Ratanaanithikul and Green, 1986 An schuelharit Standa, 1986 An schuelharit Standa, 1986 An schuelharit An sch	336.	An. sawadwongporni	1	ı	1	1	1	1	1	1	+	1	1	1
Creen, 1986 An schweftzi Stanton, 1986 An schweftzi Evans, 1934 An schwetzi Evans, 1934 An schwetzi Evans, 1934 An schwetzi Evans, 1934 An schwetzi Evans, 1929 An smithi An sergentii An sergentii An sergentii An sergentii An sergentii An schweftzi A		Rattananarithikul and												
An schwelfrari An schwelfrari An schwelfrari An schwelfrari An schwelzi Batanon, 1915 An schwelzi Batanon, 1957 An seydeli Fewards, 1929 An solomonis Relatin, Kright and Rozeboan, 1957 An somalicus Rivola and Holstein, 1957 An stokesi Stokesi An subpictus Colless, 1955 An subpictus Koscoomawinangoen, 1954 An subpictus Koscoomawinangoen, 1954 An subpictus Koscoomawinangoen, 1954		Green, 1986												
An sergentii (Theobald), 1907 An sergentii (Theobald), 1907 An sergentii (Theobald), 1907 An sergentii (Theobald), 1907 An soldmain 1959 An somalicus Rivola and Holsien, 1957 An soldmain 1920 An stephensi Usion, 1901 An stephensi Colless, 1955 An subpictus Colless, 1955 An subpictus Colless, 1955 An subpictus Koaramainangoen, 1954 An subpictus Colless, 1955	337.	An. schueffneri	1	1	1	1	1	1	1	1		,		
An schwetzi Evans, 1934 An sergentii (Theobald, 1907 An sergentii (Theobald, 1907 An sergentii (Theobald, 1907 An sergentii (Theobald, 1905 An solomonis Belkin, Kright and Rozeboom, 1945 An solomonis Belkin, Kright and Rozeboom, 1945 An stephensi Liston, 1901 An stephensi Colless, 1955 An subpictus Grassi, 1955 An subwesi An subwesi		Stanton, 1915								i	ı		1	ı
Evans, 1934 An. sergentii (Theobald), 1907 An. sergentii An. s	338.	An. schwetzi	1	1	1	ı	ı	ı	+					
An sergentii An sergentii (Theobald), 1907 An sergentii (Theobald), 1907 An sergentii Theobald, 1905 An solomonis Belkin, Kriight and Rozeboom, 1945 An somalicus Rivola and Holstein, 1957 An splendidus Koidzumi, 1920 An stokesii Liston, 1901 An stokesii Colless, 1899 An sulpayesi Koesoemawinangoen, 1954 An sulpayesi Koesoemawinangoen, 1954 An sulpayesi Koesoemawinangoen, 1954		Evans, 1934								1	ı	ı	ı	1
An. seydeli + An. suldawesi 1929 An. seydeli + Edwards, 1929 + An. smithii	339	An. sergentii	1	1	ı	ı	+	+	+	1				
An. seydeli Edwards, 1929 An. smithii Theobald, 1905 An. subpictus An. subdwesi		(Theobald), 1907											ı	1
Edwards, 1929 An. smithii Theobald, 1905 An. solomonis Belkin, Knight and Rozeboom, 1945 An. solomonis Belkin, Knight and Rozeboom, 1957 An. solomonis Belkin, Knight and Rozeboom, 1957 An. solomonis An. stockesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Collesward An. sulawesi An. sulawesi Collesward Collesward	340	An. seydeli	1	1	1	ì	1	1			Ų			
An. smithii Theobald, 1905 An. solomonis Belkin, Knight and Rozeboom, 1945 An. somalicus Rivola and Holstein, 1957 An. solomoidus Koldzumi, 1920 An. stoakesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954		Edwards, 1929										ı	ı	ı
Theobald, 1905 An. solomonis Belkin, Knight and Rozeboom, 1945 An. somalicus Rivola and Holstein, 1957 An. spendiclus Koidzumi, 1920 An. squamosus An. stokesi Liston, 1901 An. stokesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954	_	An. smithii	ı	1			1	1	+					
An solomonis Belkin, Knight and Rozeboom, 1945 An somalicus Rivola and Holstein, 1957 An splendictus Koidzumi, 1920 An stephensi Liston, 1901 An stookesi Colless, 1955 An subpictus Grassi, 1899 An sulawesi Koesoemawinangoen, 1954		Theobald, 1905							-		1	ı	ı	F
Belkin, Knight and Rozeboom, 1945 An. somalicus Rivola and Holstein, 1957 An. splandicus Koidzumi, 1920 An. squamosus Theobald, 1901 An. stephensi Liston, 1901 An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954	٠.	An. solomonis	1	1	1	1	1	ì	,	1	20			
Rozeboom, 1945 An. somalicus Rivola and Holstein, 1957 + - An. splendidus + Koidzumi, 1920 + + - An. stephensi + + + Liston, 1901 + + + An. stookesi + + Colless, 1955 + + An. subpictus + + Grassi, 1899 + An. sulawesi + Koesoemawinangoen, 1954 +		Belkin, Knight and									ii.	i,	1	+
An. somalicus Rivola and Holstein, 1957 — — — — — — — — — — — — — — — — — — —		Rozeboom, 1945												
Rivola and Holstein, 1957 An. splendidus An. splendidus + - Koidzumi, 1920 An. squamosus Theobald, 1901 + + + An. stookesi + + Colless, 1955 + An. subpictus + Grassi, 1899 + An. sulawesi + Koesoemawinangoen, 1954 +		An. somalicus	1	1	1	1	1	1	+	1				
An. splendidus + - Koidzumi, 1920 + + An. squamosus + + Theobald, 1901 + + + An. stookesi + + Colless, 1955 + An. subpictus + Grassi, 1899 + An. sulawesi + Koesoemawinangoen, 1954 +											ı	ı	ı	1
Koidzumi, 1920 An. squamosus Theobald, 1901 An. stephensi Liston, 1901 An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi F. Theobald, 1954 An. sulawesi An. sulawesi An. sulawesi			i	,	1	+	1	1		,		-	1 44	
An. squamosus - - - - - - - - - - - - - - + + + + + + + + + + + + + - <		Koidzumi, 1920									٠	+	+	1
Theobald, 1901 An. stephensi Liston, 1901 An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Foesoemawinangoen, 1954		An. squamosus	1	1	1	ı	1	1	,		9	1		
An. stephensi Liston, 1901 An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954 - + + + +		Theobald, 1901											ı	1
Liston, 1901 An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954 — — — — — — — — — — — — — — — — — — —		An. stephensi	1	1	-1	+	+	+	1	+	Я	-		
An. stookesi Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954 - + + -		Liston, 1901									+	+	ı	ı
Colless, 1955 An. subpictus Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954 - + + -		An. stookesi	ı	1	1	1	-							
An. subpictus + + Grassi, 1899 An. sulawesi + + + - +		Colless, 1955								ı	1	+	ı.	ı
Grassi, 1899 An. sulawesi Koesoemawinangoen, 1954 - + + -		An. subpictus	ı	1	1	1	+		- 41		5	12.		
An. sulawesi + + Koesoemawinangoen, 1954		Grassi, 1899							1	٠	+	+	+	+
		An. sulawesi	1	1	1	+	1	1			1	+		
		Koesoemawinangoen, 1954											1	+

-									4	+		+	+
320.	An. sundaicus (Rodenwaldt) 1925	ı	1	1	ı		1						
351.	= 10	1	1	1	+	+	+	1	+		1	1	
352.	Grassi, 1899 An. swahilicus Gillies, 1964	T	1	1	1	1	1	+	1	1		1	1
353.	An. takasagoensis (Morishita) 1946	1	1	1	1	1	1	ï	1	1		1	+
354.	An tchekedii De Meillon and Leeson, 1940	1	1	1	ı	1	1	+	1	1		1	1
355.	An tessellatus Theobald, 1901	1	1	1	1	1	1	1	+	+		+	
356.	An. theileri Edwards, 1912	1	1	1	I.	1	1	+	T	1		1	
357.	An. theobaldi Giles, 1901	1	1	ı	T)	1	1	1	+	+		1	
358.	An. turkhudi Liston, 1901	1	ı	ı	ı	+	+	+	+	+		1	1
359.	An. upemba Lips, 1960	1	Ĭ	1	1	1	1	+	1	1		1	
360.	An. vagus Doenitz, 1902	1	i -	1	1	1	į.	1	+	+		+	
361.	An. vanhoofi Wanson and Lebied, 1945	1	1	1	1	1	1	+	1	ı		1	
362.	An. vaneedeni Gillies and Coetzee, 1987	1	1	1	I.	1	ſ	+	ı	ı		ſ	
363.	An. varuna Iyengar, 1924	1	1	1	1	1	1	1	+	t		ı	
364.	An. vernus Gillies and De Meillon, 1968	1	1	1	ı	1	1	+	1	ı		1	
365.	An. vinckei De Meillon, 1942	t.	î	ı	ı	1	1	+	1	1		1	1
366.	An. walravensi Edwards, 1930	r	ī	ı	1	1	1	+	1	1		1	

Table 2: Contd.

S.No.	GENUS ANOPHELES	North		South	North	Medite-	Desert	Afro-	-opul	-opul	Malay-	Chi-	
1000	The Bertant	Ameri- can	American	Ameri- can	Eura- sian	rranean		tropi- cal	Ira- nian	Chinese	sian	eseu	asian
367.	An. watsonii	1	1	1	1		,						
	(Leicester), 1908								1	1	+	ı	
368	¥	1	1	ı	1	1	1	4					
	Theobald, 1904								1	ı	ı	ı	
369	A	ı	1	1	1	- 1							
	(James), 1903						i.	1	+	+	L	ı	
370.	A	1	1										
	Evans, 1934		i		ı	1	ı	+	1	ı	ı	1	
	SUBGENUS KERTESZIA												
371.	An. bambusicolus	ı	ı	+	1								
	Komp, 1937						ı	1	ı	ı	ı	1	
372.	An. bellator	ı	1	+	1	1							
	Dyar and Knab, 1906						ij	1	ı	1	Î	1	
373.	A	1	+	+	1	-1	1	ı	1	,	_)	1	
	(Theobald), 1905										i	ı	
374		1	+	+	1	1		1					
	Dyar and Knab, 1908									ı	1	1	
375.	An. gonzalezrinconesi	1	1	+	ı		1						
	Cova Garcia, Pulido F. and						Į.	ı	1	1	1	ı	
	Escalante de Ugueto, 1977												
376.	An. homunculus	ı	+	+	-								
	Komp, 1937			-			ı	1	1	1	1	ì	
377.	An. laneanus	1	1	+	1		11						
	Correa and Cerqueira, 1944						ı	1	ı	1	ı	1	
378.	An. lepidotus	1	1	+	1								
	Zavortink, 1973						0	1	1	1	1	1	
379.	An. neivai	+	+	+	1								
	Howard Dvar and Knah 1912			70			j.	ſ	1	1	ı	ı	

Zavorunk, 1973											
The realization			4	1		-		1	1	1	
Cova Garcia, Pulido F. and	1	1	٠	ı		1					
Escalante de Ugueto, 1977											
SUBGENUS LOPHOPODOMYIA											
82. An. gilesi	1	1	+	1	1	1	1	1	1	1	
(Peryassu), 1908											
83. An. gomezdelatorrei	1	1	+	1	ı	1	1	1	1	1	
Levi-Castillo, 1955											
84. An. oiketorakras	1	1	+	1	1	1	1	1	1	1	
Osomo-Mesa, 1947											
85. An. pseudotibiamaculatus	1	1	+	£	1	1	1	1	1	1	
Galvao and Barretto, 1941											
86. An. squamifemur	1	+	+	£	ı	1	1	1	T.	1	
Antunes, 1937											
87. An. vargasi	1	1	+	1	ı	1	1	1	1	1	
Gabaldon, Cova Garcia,											
and Lopez, 1941											
SUBGENUS NYSSORHYNCHUS											
88. An. albimanus	+	1	+	1	1	1	1	1	1	1	
Wiedemann, 1820											
89. An. albitarsis	1	1	+	1	1	1	1	1	ı	1	
Lynch Arribalzaga, 1878											
90. An. allopha	1	1	+	1	1	ā	1	1	1	1	
Peryassu, 1921											
91. An. anomalophyllus	1	+	1	1	1	1	1	1	1	ı	
Komp, 1936											
92. An. antunesi	í	1	+	1	t	ŧ	1	1	ı	1	
Galvao and Amaral, 1940											
93. An. aquasalis	1	+	+	1	1	ı	1	1	1	1	
Curry, 1932											

Table 2: Contd.

	S.No. GENUS ANOPHELES	Ameri- can	Ameri- can	South Ameri- can	North Eura- sian	Medite- rranean	Desert	Afro- tropi- cal	Indo- Ira- nian	Indo- Chinese Hills	Malay- sian	Chi-	Austral- asian
394.	An. argyritarsis	+	+	+	1	1	1	1	1	1	1	1	1
	Robineau-Desvoidy, 1827			Ť									
395.	An. benarrochi	t	ı	+	1	1	ı	1	1	1	1	1	1
	Gabaldon, Cova Garcia, and Lopez, 1941												
396	An. braziliensis	1	+	+	1	ı	1	1	1	1	1	1	1
	(Chagas), 1907												
397.	An. darlingi Root, 1926	+	+	+	1	1	1	1	1	1	ı	1	ı
398	An. deaneorum	1	1	+	1	1	ı	1	1	1	1	i	1
	Rosa-Freitas, 1989												
399	An. evansae	1	1	+	1	1	1	1	1	1	1	1	1
	(Brethes), 1926												
400	An. galvaoi	1	1	+	1	1	1	1	1	1	1	1	1
	Causey, Deane, and												
	Deane, 1943												
401	An. ininii	1	1	+	1	1	ī	1	1	1	1	1	1
	Senevet and Abonnenc, 1938												
402.	An. lanei	1	1	+	1	1	1	1	1	1	1	1	1
	Glavao and Amaral, 1938												
403	An. lutzii	1	1	+	1	1	ı	1	1	1	1	1	1
	Cruz, 1901												
404	An. marajoara	1	+	+	1	1	1	1	1	ı	1	1	1
	Galvao and,												
	Damasceno, 1942												
405.	An. nigritarsis	1	1	+	1	1	1	1	1	ı	1	1	1
	(Chagas), 1907												
406.	An. noroestensis	1	1	+	1	1	ı	1	1	1	1	1	1
	Galvao and Lane, 1937												
407.	An. nuneztovari	1	1	+	1	1	1	1	1	1	1	1	1
	Gabaldon, 1940												

(Pervesell) 1922												
An. parvus	1	1	+	ı	1	ı	ı	1	ı	1	1	1
(Chagas), 1907 An. pictipennis	1	1	+	ı	1	1	1	ı	ī	1	1	1
(Philippi), 1865	1	1	+	1	,	1	1	ı	1	ı	1	'
Gabaldon, Cova Garcia, and												
Lopez, 1940												
(Neiva and Pinto), 1922	1	i	+		1	1	ı	ı	ı	ı	1	ly-
An. sanctielii	i	1	+	1	1	-1	ı	1	ı	1	1	1
Senevet and Abonnenc, 1938												
An. sawyeri	1	1	+	1	1	1	ţ	1	L	1	1	1
Causey, Denane, Denane and Sampaio, 1943												
An. strodei	+	+	+	1	1	Î	1	1	1	1	1	1
Root, 1926												
An. triannulatus	1	+	+	1	1	1	1	1	1	1	+	1
(Neiva and Pinto), 1922												
An. triannulatus davisi	1	I	+	1	1	1	1	1	1	1	1	1
An trinked												
Com 4070	ı	ı			1	1		1	1			
SUBGENUS STETHOMYIA												
An. acanthotorynus	1	1	+	1	1	1	1	1	1	ī	1	1
Komp, 1937												
An. canorii	1	1	+	1	í	1	1	ì	1	1	1	1
Floch and Abonnenc, 1945												
An. kompi	1	+	+	1	1	1	1	1	1	1	1	1
Edwards, 1930												
An. nimbus	1	+	+	1	1	1	1	1	1	1	1	1
(Theobald), 1902												
An. thomasi	1	1	+	1	1	1	1	ı	1	1	1	1
Shannon, 1933												

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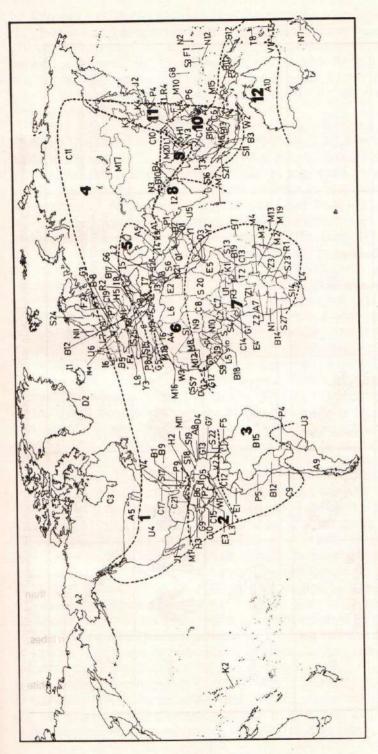
Table 3: COUNTRIES UNDER 12 EPIDEMIOLOGICAL ZONES OF MALARIA

NORTH	AMERICAN	CENTRA	L AMERICAN	SOUTH	AMERICAN
CODE	COUNTRIES	CODE	COUNTRIES	CODE	COUNTRIES
A2	Alaska	C12	Cayman Islands	A8	Argentina
A5	Amazonia	C15	Costa Rica	B12	Bolivia
A9	Antigua & Barbuda	C17	Cuba	B15	Brazil
B1	Bahamas	D5	Dominican Republic	C9	Chile
B8	Berlin	E3	El Salvador	C12	Colombia
B9	Bermuda	G9	Guatemala	E1	Ecuadior
C3	Canada	H2	Haiti	F5	French Guiar
D4	Dominica	НЗ	Hondrus	G10	Guina
G7	Grenada	J1	Jamaica	G13	Guyana
J1	Jamaica	N8	Nicaragua	P4	Paraguay
L3	Lesser (Antilles)	P2	Panama	P5	Peru
M11	Martinique	V2	Venezuela	P9	Puerto Rico
M14	Mexico	W1	West Indies	S22	Surinam
S3	Santa Cruz Islands	0.00		U3	Uruguay
S5	Sardinia			00	Oraguay
S8	Sicily				
S17	St. Kitts-Nivis				
S18	St. Lucia				
S19	St. Vincent				
U4	U.S.A.				
-	EURASIAN	MEDITE	RRANEAN	DESERT	
CODE	COUNTRIES	CODE	COUNTRIES	CODE	COUNTRIES
A1	Afghanistan	A3	Albania	A4	Algeria
A6	Andorra	A4	Algeria	B2	Bahrain
A11	Austria	B5	Belgium	E2	
C11	Common Wealth of	B17	Bulgaria	14	Egypt
~	Independent States	017	bulgana	14	Iraq
C19	Czechoslovakia	C18	Cyprus	J3	Jordan
D2	Denmark	G5	Gibraltar	L6	
F2	Finland	G6	Greece	M19	Libya Morocco
F4	France	17	Israel	SI	Sahara
G3	Germany	18	Italy	S6	
H5	Hungary	L2	Lebanon	S26	Saudi Arabia
1	Iceland	M9	Malta	T6	Syria
2	India	M16	Monago	W3	Tunisia
4	Iran	M18	A CONTRACT OF THE CONTRACT OF	11515	West Sahara
5	Iraq		Morocco	Y1	Yemen North
6	Ireland	P8 S2	Portugal		
7	Liechtenstein	S15	San Marino		
_8	Luxembourg	A STATE OF THE STA	Spain		
M17	Mongolia	S25	Switzerland		
VII/	Netherlands	T7	Turkey		
		Y3	Yugoslavia		
V11	Norway				
77	Poland				
R2	Romania				
524	Sweden				
J6	U.K.				
AFROTR	and the same of th	AFROTR		INDO-IRA	
CODE	COUNTRIES	CODE	COUNTRIES	CODE	COUNTRIES
47	Angola	M4	Malawi	C13	Comoros
36	Belize	M8	Mali	12	India
37	Benin			15	Iraq
314	Botswana				

	Contd.	AFROTE	ROPICAL	INDO-IR	
CODE	COUNTRIES	CODE	COUNTRIES	CODE	COUNTRIES
	Burkina Faso	M12	Mauritania	K5	Kuwait
B18	Burundi	M16	Monago	M7	Maldives
B19	Cameroon	M19	Mozambique	M21	Muscat
C2	Cape Verde	N1	Namibia	01	Oman
C4	Cape Verde Islands	N9	Niger	P1	Pakistan
C5	Central African Republic	N10	Nigeria	Q1	Qatar
C7	Chad	R1	Rhodesia	S6	Saudi Arabia
C8	Control of the Contro	R3	Rwanda	S9	Seychelles
C14	Congo Dahomey	S4	Sao Tome Principal	S16	Sri Lanka
D1 D3	Diibouti	S7	Senegal	U5	United Arab
E5	Ethiopia	S10	Sierra-Leone		Emirates
E4	Equatorial Guinea	S13	Somalia		
G1	Gabon	S14	South Africa		
G2	Gambia	S20	Sudan		
	Ghana	S23	Swazibad		
G4	Guinea	S27	South West African		
G11	Guinea	JL1	Republic Organization		
040	Guinea-Bissak	T2	Tanzania		
G12		T4	Togo		
19	Ivory Coast	U1	Uganda		
K1	Kenya	U2	Upper Volta		
L4	Lesotho	Y2	Yemen West		
L5	Liberia	Z1	Zaire Zaire		
M2	Madagascar	Z2	Zambia		
МЗ	Malagary Republic	Z3	Zimbabwe		
	A	MALA		CHINE	SE
The state of the s	CHINESE HILLS		COUNTRIES	CODE	COUNTRIES
CODE	COUNTRIES	CODE		Victoria de la composition della composition del	China
B4	Bangladesh	B3	Bangka Island	C10	C.I.S.
B10	Bhutan	B13	Borneo	C11	Japan
C1	Cambodia	B16	Brunei	J2	Korea North
C10	China	C6	Celebes	K3	Korea South
H1	Hainan Island	F3	Formosa	K4	
12	India	G8	Guam	R4	Ryokyo Islan Taiwan
L1	Laos	H4	Hong Kong	T1	laiwan
M1	Macau	13	Indonesia		
M20	Myanmar	M5	Malaya		
N3	Nepal	M6	Malaysia		
S11	Singapore	M15	Molucca		S SE THE S
S21	Sumatra	P6	Philippines		
T1	Taiwan				
ТЗ	Thailand				
V3	Vietnam				
The second second	RALASIAN	AUST	RALASIAN		
CODE		CODE			
11 (000) (000) (000)	7.7.7.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1.1	N7	New Zealand		
A10	Australia	P3	Papua New Guinea		
B11	Bismarck Archipelago		Solomon Islands		
F1	Fiji	S12	Tonga		
K2	Kiribati	T5	Tuvalu		
M10	Mariana Islands	T8			
M13	Mauritius	V1	Vanatua Viscia Jalanda		
N2	Nauru	V4	Virgin Islands		
N6	New Guinea	W2	West Irian		

Table 4: DIFFERENCES BETWEEN ANOPHELES AND CULEX MOSQUITOES

Anopheles	Culex
EGG	Project Company of the second of the
(1) Shape	
Boat like, provided with lateral floats.	Elongated, no lateral floats.
(2) Number per Oviposition: 80-140 Laid singly	90-170 Laid singly (Aedes) in rafts (Culex) or in cluster (Mansoniodes).
LARVA	
(1) Position Float parallel to the surface of water.	Hangs from surface of water.
(2) Siphon tubes	
Absent (3) Movement	Present
Characteristic swift movements.	Much slower and often snake-like movements
(4) Respiratory opening On 8th abdominal segment.	At the end of siphon tube.
(5) Palmate hairs Present on abdominal segments arranged in pairs (sometimes on metathorax).	Absent
PUPA	
(1) Siphon tubes Funnel shaped, short and broad.	Long and narrow.
ADULT	
(1) Resting posture Proboscis, head and abdomen are in a straight line with abdomen pointing away from resting surface.	Proboscis and body are not in a straight line, the abdomen being inclined towards resting surface.
(2) Wings Generally spotted with white and dark scales.	Generally unspotted with only dark scales.
(3) Palpi	
Female Palpi slender and equal to the proboscis.	Palpi stub-like, reduced.
Male	
Club-shaped at distal ends and nearly equal to proboscis.	Pointed, bent and usually longer than proboscis.
(4) Scutellum Half-moon-shaped with a uniform row of hairs along the margin.	Trilobed, with three bunches of hair on lobes.
(5) Abdomen Without scales or with a few scattered scales.	With uniform rows of overlapping flat, white and dark scales.



					Source: WHO/VBC/89,967
7. Afrotropical	8. Indo-Iranian	9. Indo-Chinese Hills	10. Malaysian	11. Chinese	12. Australasian
1. North American	2. Central American	3. South American	4. North Eurasian	5. Mediterranean	6. Desert
	2. (3. 6	4.	5.	6. [

Figure 1: The zones of malaria epidemiology.

Optimum temperature		22^{0} 28^{0} C for all stages			Z	Daily mean temperature 28°C to 30°C Daily mean relative humidity 60 to 80%
Development period in days	period in days		6-8	1.5-2	ΪŽ	Female approximately 30 days under ideal natural conditions; male 2 to 6 days for different species
Figure	NES				*	No.
	CULICINES	spelleng			一 。	
	ANOPHELINES ANOPHELES	opposition of			***	A STATE OF THE STA
Name		Eggs	Lava	Pupa	Head	Resting Position.

Figure 2: Differences between various developmental stages of Anopheles, Aedes and Culex mosquitoes.

James and Liston (1904), in a monograph on the anopheline mosquitoes of India, divided Indian anophelines into 10 groups: (1) An. barbirostris, An. nigerrimus; (2) An. leucosphyrus, An. punctulatus; (3) An. pulcherrimus, An. willmori; (4) An. karwari, An. fuliginosus, An. jamesii, An. maculipalpis, An. theobaldi, An. maculatus; (5) An. jeyporiensis, An. listoni, An. culicifacies; (6) An. rossi, An. stephensi; (7) An. turkhudi; (8) An. lindesayi, An. gigas; (9) An. aitkeni, An. immaculatus and (10) An. culiciformis.

In the revised edition published in 1911, they re-grouped the Indian anophelines into 13 genera: Neostethopheles, Patagiamyia, Myzomyia, Anopheles, Pyretophorus, Nyssorhynchus, Nyssomyzomia, Neomyzomia, Neocellia, Cellia, Myzorhynchus, Christophersia, and Aldrichia (40 spp. and 5 varieties). Christophers (1916) dealt with the synonyms and the correct names of the species of anophelines recorded. A total of 38 species was listed from India. Edwards in 1932 prepared a comprehensive catalogue for the identification of the family Culiciade and reduced the number of genera from 149 (Theobald, 1910) to 30. However, the number of species was increased from 1050 to 1400. Christophers (1933) published a monograph on the identification of anophelines of India, Ceylon (Sri Lanka) and Burma under the title The Fauna of British India. In a series of articles published in the Indian Journal of Medical Research, from 1923 to 1929, Barraud provided a revision of the culicine and toxorhynchine which eventually led to the publication of his monograph The Fauna of British India in 1934. Both Christophers (1933) and Barraud (1934) adopted the classification of Edwards (1932) in the arrangement of tribes, genera and subgenera.

Subsequently, Stone, Knight, and Starcke published a catalogue on the Culicidae of the world in 1959. This was followed by the publication of a catalogue on the systematics of mosquitoes of the world by Knight and Stone in 1977. A first supplement was written by Knight in 1978 and two additional supplements by Ward in 1984 and 1992. The authors raised the number of genera, and species from 31 to 37, and from 2,401 to 3,209 respectively.

The first key in the form of a wall chart to identify the Indian anophelines was published by the Central Malaria Bureau, Kasauli, in 1912. Subsequently several keys were published by Sinton and Covell (wall chart, 1916), Strickland and Choudhury (1927a), Christophers et al. (1927), Christophers (1933), Puri (1954, 1960), Wattal and Kalra (1961), and Das et al. (1990). Of the above identification keys, only 2 are commonly used, namely the keys of Puri (1960) and Wattal and Kalra (1961). The keys deal with the identification of 40 mosquito species as against the present fauna of 58, except the key by Das et al. (1990), which describes 54 species. Also, none of the keys gives a complete account of morphological variations except for some variations reported by Christophers (1933). These morphological variations are very important for the correct identification of species. There are several reports of morphological variations in 24 species recorded by Christophers (1933), Ramakrishna (1954), Subramanian and Nagendra (1955), Bhatnagar et al. (1958), Azeez and Beig (1959), Rajagopal and Chakraborty (1960), Wattal et al. (1960), Rahman et al. (1960), Sen (1962), Nagpal and Sharma (1983 a,b), Gunasekaran et al. (1990), and Nagpal (1990). The morphological variations are mostly found in the palpi, wings, and legs.

In 1930, Covell compiled the information available on the distribution of 35 species of Indian anophelines, which was later modified to include 40 species by Puri (1955). On the basis of physiography, climate, and hydrological zones, Rao (1984) compiled information on 51 species, which was updated by Nagpal (1986) to include survey information from 1900 to 1984. The records of gut and gland infections of these anophelines were first tabulated in Vectors of Malaria in India, published by the National Malaria Society in 1957. Horsfall (1955), updated in 1972, gives the gut and gland infection in regard to the anophelines of the world in his book Mosquitoes - Their Bionomics and Relation to Diseases, Later, Rao (1984) gives an exhaustive account of the biology of the Indian anophelines. In the present monograph an attempt is made to update the information available on adult anophelines, their differences from other genera at every stage, brief distinguishing features of identification of 58 anopheline species, their biology (typeform available, species complex if any, sitting posture, resting habits, breeding habits. biting time, feeding habits, flight range), susceptibility to insecticides, relation to disease, records of vector incriminations for human plasmodia, distribution in India and the world, and morphological variations reported. Also included are a bibliography and a glossary.

MORPHOLOGICAL DESCRIPTION

The life cycle of an anopheline mosquito has four distinct stages: adult, egg, larva and pupa. The adult stage is aerial and the other three stages are aquatic. The terminology used here is taken from Harbach and Knight (1980).

ADULT

The body of an anopheline mosquito comprises the head, the thorax, and the abdomen (Fig. 3). The main parts of the body and their appendages are briefly described here.

Head

The head is the smallest part of the mosquito body, is almost spherical, and is connected to the thorax through a narrow, membranous neck. The head consists of a pair of antennae, a proboscis, a pair of maxillary palpi and a pair of compound eyes (Fig. 4). The eyes occupy the bulk of the surface area of the head. The head is characterized by the presence of white and black scales: the back (occiput) and side portions of the head are covered with black scales while the vertex is covered with white or pale and black scales which are truncated and notched apically, giving a forked appearance. Between the two eyes on the narrow anterior part of the vertex a group of pale or white scales and setae arise which form a tuft, known as frontal or vertical tuft a characteristic of most anophelines.

Maxillary palpi (palpi): A pair of palpi arises from the lower front margin of the head near the proboscis. The palpus has five palpomeres. The small basal segment is designated as the first palpomere, which is fused with the second and is vestigial. The distal segment, which is away from the head, is known as the fifth palpomere (see Fig. 4). The palpi in the female are slender while they are club shaped towards the apical end in males. The palpi are also covered with scales except along their inner surface. The thickness of palpi depends on the type of scales, i.e., whether they are erect or flat. Erect scales give thickness to the palpi (e.g. An. barbirostris complex and An. hyrcanus complex), while flat scales make them thin (e.g., An. aitkenii complex). The pale scales on the palpi form the bands at the joints 2–3 and 3–4, and at the apex. The absence or presence of the pale bands, their number, their width, and the distance between the two bands are important characters for species identification.

The terminology used for the bands present on the palpi is the same as used by Christophers (1933). The pale or the white band which is away from the head, i.e., towards the distal end, is called the apical pale band; the second pale band is known as the subapical pale band; and the third band, which is towards the

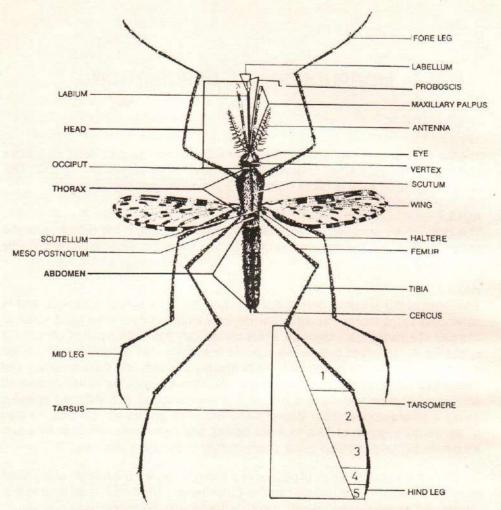


Figure 3: Generalized diagram of a female adult anopheline.

basal side, is known as the basal or proximal pale band. The dark band between the apical and subapical pale bands is known as the pre-apical dark band or intervening dark band. Palpi generally contain three pale bands because of the shorter apical segment. When the apical segment increases in size it frequently contains one more pale band and the palpi become four-banded.

In Indian anophelines the presence of pale bands is more dominant in subgenus Cellia but in subgenus Anopheles most of the species contain completely dark palpi except for An. hyrcanus group species, An. lindesayi, An. nilgirius, An. baileyi, and An. gigas. In subgenus Cellia, out of the 34 species recorded from India only six contain four-banded palpi: An. balabacensis, An. dirus, An. elegans, An. pulcherrimus, An. karwari, and An. tessellatus. In a few three-banded species, viz., An. annularis, An. pallidus, An. subpictus, An. vagus, An. sundaicus,

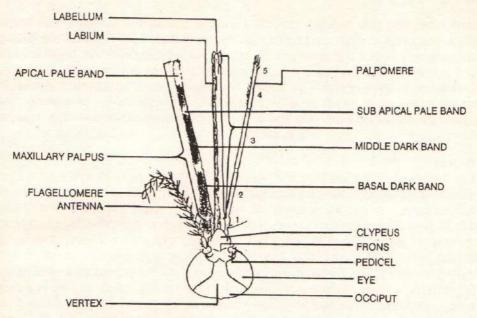


Figure 4: Parts of the head of a female anopheline mosquito.

and An. fluviatilis, an extra dark band appears on the apical pale band and gives the appearance of four-banded palpi. This variation creates a problem in the identification, but it is found mostly in winter collections. Another variation which gives the appearance of four-banded palpi and may mislead in the identification is the presence of speckling between the basal and subapical pale bands (middle dark band) of the palpi. Although there are only two species—An. stephensi and An. splendidus—in India in which speckling is present on the palpi, sometimes speckling is also recorded in An. maculatus. The other variation which is common in An. aconitus is the absence of the pre-apical dark band which increases the length of the apical pale band and creates confusion in the identification of the species.

Antennae: Antennae are sense organs which arise on the front of the head between the eyes and consist of a flagellum of 13 flagellomeres in females and 14 flagellomeres in males (see Fig. 4). The basal flagellomeres is an inconspicuous ring and is known as torus or pedicel. In the male mosquito the antennae are plumose owing to the presence of whorls of long hairs, and the size of the pedicel is comparatively large. In females the hairs are smaller, fewer, and less conspicuous, and the pedicel and the first few flagellar segments often bear a few scales.

Proboscis: The proboscis projects downwards and forwards from the lower front margin of the head and generally is continuously dark in colour, except the labellum, which is light yellow (Fig. 4). In An. aconitus, An. kochi, and An. tessellatus, the apical half of the proboscis is flavescent, i.e., turns yellow in colour, and

sometimes has pale patches towards the distal end as in *An. vagus*. The length of the proboscis is measured by comparing it with that of the femur of the foreleg and is a very useful character to differentiate *An. elegans* from *An. balabacensis* and *An. dirus*. The proboscis also consists of a sheath-like structure, known as labium, enclosing a group of six styles. The labium serves as a protective sheath for the styles but does not enter into the wound when the mosquito is biting. The styles enter into the host skin and form a duct through which saliva is injected into the wound and liquid food is sucked in.

Thorax

The thorax is the middle part of the body. It bears two pairs of wings (one pair, haltere, modified for balancing during flight) and three pairs of legs and has three distinct parts — pronotum, scutum, and metanotum — each part consisting of one pair of legs. The pronotum is reduced to form a pair of antepronotal lobes lying dorsally on either side of the neck with a postpronotary area. The scutum is the largest part of the thorax. It is posteriorly attached with a small scutellum and mesopostnotum. The metanotum is also reduced as the pronotum and consists of balancing organs (halteres) and one pair of hind legs (Figs. 5 and 6). In Indian anophelines the thorax has very little importance in the identification of species. The colour of scutum helps in the identification of An. aitkenii group, An. lindesayi, An. kochi, and An. maculatus. The presence or absence of hair and setae is useful in the identification of An. stephensi, An. umbrosus group, An. turkhudi and An. multicolor.

Wings: A pair of wings arises from the scutum and bears a number of important characters which help to identify the Indian subgenus and species. The wing membrane consists of a number of longitudinal veins, which are connected with each other through small cross veins. The thickened anterior border of the wing is known as costa and parallel to it is the subcosta, which fuses with the costa in the

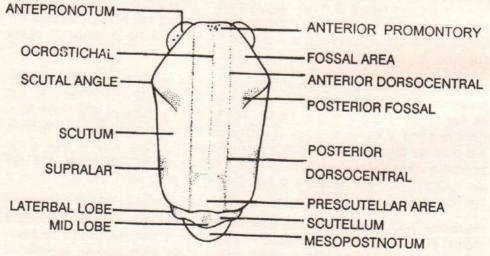


Figure 5: Parts of the thorax of an anopheline mosquito (dorsal view).

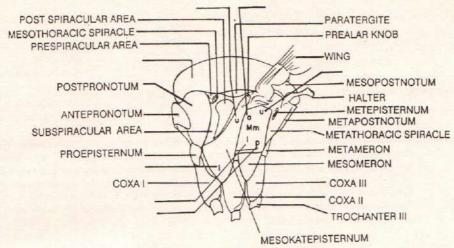


Figure 6: Lateral view of an anopheline thorax.

middle of the anterior border of the wing. Besides these two important veins, there are six longitudinal veins known as veins 1(R1), 2(RS), 3(R4 + 5), 4(M), 5(Cu), and 6(Anal); as given in Fig. 7. Veins 1(R1), 3(R4 + 5), and 6(Anal) are undivided and veins 2(RS), 4(M), and 5(Cu) are bifurcated and their numbers designated 2(RS), 2.1(R2), 2.2(R3), 4(M), 4.1(M1), 4.2(M2), 5(Cu), 5.1(Cu1), 5.2(Cu2). Veins 2(RS) and 4(M) and subcosta are concave and prominent on the under surface of the wing and veins 1(R1), 3(R4 + 5), 5(Cu), and 6(Anal) are convex and prominent on the upper surface of the wing. The cross veins are known as veins 2-3 (RS)-(R4 + 5), 3-4, (R4 + 5)-(M) and 4-5 (M)-(Cu) as they connect the stems of veins 2(RS) and 3(R4 + 5), 3(R4 + 5) and 4(M) and 4(M) and 5(Cu) respectively (Fig. 7). The part of vein 2(RS) between its bifurcation and the origin of the 2-3 cross vein is known as petiole, an important character in the identification. The areas of the wing enclosed by the branches of veins 2(RS) and 4(M) are known as anterior and posterior forked cells respectively, these being the most important characters to ident y the species An. moghulensis. This species resembles An. jeyporiensis but can be easily identified by measuring the distance of the anterior forked cell from the base of the wing, which is more in this species than in An. jeyporiensis. The costa, subcosta, and veins 1(R1) to 6(Anal) are covered with scales on both the upper and under surfaces. Both the type of scales and their size vary from place to place on different veins. The scales are known as squame, plume, fringe, and border scales (Christophers 1933).

The ornamentation of the wing or scaling on the wing is of great importance in identification. Generally in all Indian anophelines, dark and white areas alternate with each other, but in An. barbirostris and An. nigerrimus white and black scales on some veins are mixed together and give a speckled appearance. There are seven species in Indian fauna in which the wing is completely dark, giving a Culex like appearance. These species are An. aitkenii, An. culiciformis, An. bengalensis, An. pinjaurensis, An. insulaeflorum, An. sintoni, and An. barianensis of the subgenus Anopheles. In India only two subgenera are recorded, viz. Cellia

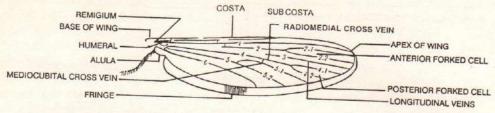


Figure 7: Wing of an anopheline mosquito showing costa, subcosta and 6 longitudinal veins; 1-first longitudinal vein (Radius, R1); 2-second longitudinal (Radial sector, Rs); 2.1-upper or anterior branch of 2nd longitudinal (R2); 2.2-lower or posterior branch of 2nd longitudinal (R3); 3-third longitudinal (R4+5); 4-fourth longitudinal (Media, M); 4.1-upper branch of 4th longitudinal (M1); 4.2-lower branch of 4th longitudinal (M2); 5-fifth longitudinal (Cubitus, Cu); 5.1-upper branch of 5th longitudinal (Cu1); 5.2-lower branch of 5th longitudinal (Cu2); 6-sixth longitudinal (Anal).



Figure 8: Wing showing position of pale and dark spots and their nomenclature.

and Anopheles. In subgenus Cellia the dark and pale areas are very clear on the wings, especially on costa, subcosta and vein 1(R1). These pale and dark areas, also known as spots, have a characteristic position and are of great importance in the identification of subgenus and species. The subgenus Cellia has four or more pale areas while the subgenus Anopheles has three or fewer pale areas (also, the wing is completely dark) on costa and subcosta, including vein 1(R1). These pale and dark areas are recognized by their names (Fig. 8). The medium pale area, i.e. subcostal pale spot, is present a little beyond the middle of the costa where the subcosta join the costa. The dark area which starts after the subcostal pale spot towards the distal side (i.e. nearer the apex of the wing) is known as the pre-apical dark mark followed by the pre-apical pale spot, apical dark mark, and apical pale spot. The pale spot, which is present just before the subcostal dark spot (i.e. towards the base of the wing), is known as the middle dark spot followed by the accessory sector pale spot (present mostly on vein 1(R1)), sector pale spot, pre-sector dark spot, pre sector pale spot, and a few other pale and dark spots, which vary in number and position and are important in the identification of the species. The terminology given above for the dark and pale areas is taken from Harbach and Knight 1980; but different authors have used different terminologies for the dark and pale areas. The names used by different authors are given in Table 5 (taken from Wilkerson and Peyton, 1990).

Towards the apex and hind margin of the wing, fringe scales or spots are present; usually the fringe scales are pale in colour and are present opposite to the end of the veins. Their number and presence or absence is very useful in the identification of some Indian anophelines.

Legs: There are three pairs of legs originating from the lower side of the thorax: the forelegs, midlegs and hindlegs. Each leg consists of conical coxa, a small hinge-like trochanter, a long femur, a slender tibia, and a five-segmented tarsus (Fig. 9). The fifth tarsomere, which is towards the end of the leg, is smallest in size while the first tarsomere, which is attached with the tibia, is longest in size (nearly equal to the tibia) in female mosquitoes. But in male mosquitoes the fifth tarsomere is often longer than the fourth tarsomere of the foreleg. The fifth tarsomere of each leg consists of claws in females but the foreleg of the male consists of very few spurred claws with some stout hairs.

Ornamentation of the leg is of great importance in identification. The ornamentation is dependent on the presence or absence of the pale and dark bands on the different parts of the leg. The legs are covered with small oppressed scales except the coxae and sometimes the trochanter. Both the femur and tibia are sometimes pale towards their tips, and this condition is referred to as tibia or femur with knee spots. The presence of a broad pale area at the junction of tibia and tarsal of the hind leg is of great importance in the identification of *An. balabacensis, An. elegans* and *An. dirus* complex. In some species the femur of forelegs is swollen in their basal half and sometimes the femur and tibia both pale towards the posterior side.

In the case of An. lindesayi the hind femur has basal and subapical pale bands or sometimes the femur has a tuft of pale scales in place of subapical pale band which can be seen through the naked eye as in the case of An. annandalei and An. interruptus (An. asiaticus group). Out of 34 Indian species under the subgenus Cellia the legs of 20 species are referred to as speckled legs, due to the presence of scattered and irregular patches of pale spot on the femur, tibia and some tarsal segments. There is no speckled leg in the Indian mosquito under the subgenus Anopheles. The most important features of the leg are the tarsal segments. These segments may be completely dark (An. culicifacies, An. fluviatilis, An. minimus, An. varuna in subgenus Cellia and An. culiciformis, An. aitkenii complex in subgenus Anopheles) or have pale bands at the joints. There are two types of bands: (1) very narrow bands confined to the apices, i.e., towards the tips only and leaving the bases dark (e.g., An. jeyporiensis, An. stephensi), and (2) broad bands spreading across the joints on the bases of the next segment (e.g., An. subpictus, An. vagus). These type of bands are more prominent on the foreleg than on the mid- and hind-leg.

In eight Indian species of subgenus Cellia—An. annularis, An. pallidus, An. philippinensis, An. nivipes, An. pseudojamesi, An. jamesii, An. splendidus, and An. pulcherrimus, the pale band is very broad on hind-leg tarsomeres because of the fusion of distal tarsomeres 5,4,3, and apex of 2, which are completely white. It is important that the segments be counted from the tip (segment 5) of the leg to the first dark band.

Table 5: COMPARISON OF SELECTED SYSTEMS FOR NAMING PALE

Proposed nomenclature	Christophers 1913	Root 1926	King 1932	Christophers 1933
Basal pale	Not shown	B1	Not shown	Not shown
Basal dark	Basal accessory spot (in part)	Not named	Inner accessory dark spot (in part)	Inner prehu- meral dark accessory spot
Prehumeral pale	Not shown	Not shown	Not shown	Prehumeral pale interr.
Prehumeral dark	Basal accessory spot (in part)	Not named	Inner accessory dark spot (in part)	Outer prehu- meral dark accessory spot
Humeral pale	Humeral nodal point	B2	White interr. basal 1/3 of costa	Humeral pale interr.
Humeral dark	Basal accessory pigment area	Not named	Outer accessory dark spot	Humeral dark accessory spot
Presector pale	Inner pale spot	В3	Presector white spot	Presector pale area
Presector dark	Inner main costal spot center	Not named	Presector dark spot	Presector dark spot
Sector pale	Sector nodal point	M1	Sector spot	Sector pale area
Sector dark	Middle main cos- tal spot center	Not named	Midcostal spot	Middle dark spot
Accessory sector pale	Accessory sector nodal point	M2 and M3	Accessory sector spot	Accessory sector pale area
Subcostal pale	Subcostal nodal point	Sc	Subcostal white spot	Subcostal pale area
Preapical dark	reapical dark Subapical main costal spot center		Subapical dark spot	Pre-apical dark spot
Preapical pale	C.1	Apical pale (in part)	Subapical white spot	Pre-apical pale area
Apical dark	Apical main cos- tal spot center	Not named	Apical dark spot	Apical dark spot
Apical pale	Apical nodal point	Apical pale (in part)	Apical white spot	Apical pale area

[&]quot;Undivided, prehumeral dark accessory spot.

b"C" is pale interruption of third dark area of first vein.

Also contains accessory R1 pale on R1.

When absent, forms the subbasal dark.

AND DARK COSTAL WING SPOTS IN GENUS ANOPHELES

Gater 1935	Russell et al. 1943	Reid 1968	Gillies & de Meillon 1968	Zavortink 1973
Not named	Not named	Not named	Not shown	Basal pale
Inner prehu- meral dark accessory spot ^a	Not shown	Prehumeral dark mark (in part)	Not named	Basal dark (in part)
Prehumeral pale interr.	H1	Prehumeral pale spot	Not shown	Not shown
Outer prehu- meral dark accessory spot ^e	Not named	Prehumeral dark mark (in part)	Not named	Basal dark (in part)
Humeral pale	H2	Humeral pale spot	"a," humeral pale marking	Humeral pale
Humeral dark accessory spot	Not named	Humeral dark mark	Not named	Humeral dark (in part)
Presector pale area	НЗ	Presector pale spot	"a," presector pale	Not shown
Presector dark spot	Basal dark spot	Presector dark mark	"A"	Humeral dark (in part)
Sector pale area	Sectoral spot	Sector pale spot	"b," sector spot	Presectoral pale
Middle dark spot	Median dark spot	Middle dark mark	"B"	Presect, dark (in part) sec- toral dark (in part)
Accessory sector pale area	Accessory sector spot	Accessory sector pale spot	"b," accessory sector pale spot	Sectoral pale
Subcostal pale area	Subcostal spot	Subcostal pale spot	"c," subcostal	Subcostal pale
Pre-apical dark spot	Pre-apical dark spot	Pre-apical dark mark	"C"b	Pre-apical dark
Pre-apical pale area	Pre-apical spot	Pre-apical pale spot	"d," subapical	Pre-apical pale
Apical dark spot	Apical dark spot	Apical dark mark	"D"	Not named
Apical pale Irea	Apical spot	Apical pale spot	"e," apical spot	Not shown

(Contd.)

Table 5: Contd.

Proposed nomenclature	Harrison & Scanlon 1975	Faran 1980	Harrison 1980	Harbach & Knight 1980
Basal pale	Not shown	Basal pale	Basal pale	Basal pale
Basal dark	Inner prehu- meral access- ory dark	Not shown	Basal dark	Basal dark
Prehumeral pale	Prehumeral pale	Basal pale	Prehumeral pale	Prehumeral pale
Prehumeral dark	Not named	Basal dark	Prehumeral dark	Prehumeral dark
Humeral pale	Humeral pale	Humeral pale	Humeral pale	Humeral pale
Humeral dark	Humeral dark	Subbasal dark	Humeral dark	Humeral dark
Presector pale	Presector pale	Subbasal pale	Presector pale	Presector pale ^d
Presector dark	Presector dark	Presectoral dark	Presector dark	Presector dark
Sector pale	Sector pale	Presectoral pale	Sector pale	Sector pale
Sector dark	Middle dark	Sectoral dark (in part) sub- costal dark (in part)	Sector dark	Median dark
Accessory sector pale	Accessory sector pale	Sectoral pale	Accessory sector pale	Accessory sector pale
Subcostal pale	Subcostal pale	Subcostal pale	Subcostal pale	Subcostal pale
Pre-apical dark	Pre-apical dark	Pre-apical dark	Pre-apical darke	Pre-apical dark
Pre-apical pale	Pre-apical pale	Pre-apical pale	Pre-apical pale	Pre-apical pale
Apical dark	Apical dark	Apical dark	Apical dark	Apical dark
Apical pale	Not named	Not shown	Apical pale	Apical pale

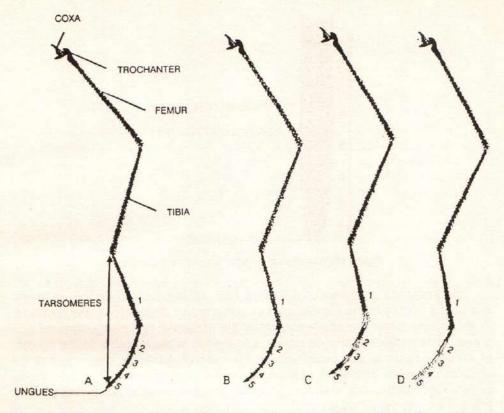


Figure 9: Leg of an anopheline mosquito showing different parts (A); speckling on femur, tibia and tarsomeres (B); foreleg tarsomeres with broad pale bands (C); hind tarsomeres 5, 4 and 3 portion of 2nd completely white (D).

In the case of *An. maculatus, An. theobaldi, An. willmori, An. pseudowillmori* only the tarsomeres 5 and 4 of the hind leg are completely white. Sometimes in a few cases the tarsomere 5 of *An. kochi, An. balabacensis, An. elegans, An. dirus* complex and *An. tessellatus* is not completely white and therefore old keys refer to their identification at two places; but because of the presence of four-banded palpi, however, the identification of these species in this book is given at only one place.

Abdomen

The abdomen of anophelines is composed of 10 segments but externally only eight segments can be seen, the last two being modified into genital organs (Fig. 10). The dorsal part of the abdomen is called dorsum and the segments are known as tergites; the ventral side is known as venter and segments are called sternites. The tergites and sternites join laterally through a membrane known as pleural membrane anteriorly and as intersegmental membranes posteriorly. In the case of male anophelines the dorsal surface of the eighth segment is made up of sternite instead of tergite as after emergence this segment rotates with genitalia at an angle of 180°.

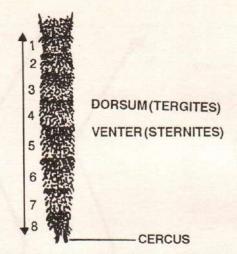


Figure 10: Abdomen of an adult anopheline mosquito.

The presence or absence of scaling, hair, coxites, or ceri on the abdomen is a useful feature in the identification of some Indian anophelines. For example, in *An. pulcherrimus* the entire dorsum of the abdomen is densely covered with broad, white battledore-shaped scales, and the tuft of black scales on the corners of middle tergites is also prominent. In the case of *An. kochi* the venter of the abdomen consists of the tuft of black scales on segments 2 to 7 at their joints, while in *An. barbirostris* and *An. hyrcanus* group of mosquitoes a single tuft is present on segment 7 only.

Male genitalia: The male genitalia is also known as external genitalia, terminalia, or hypopygium—a copulatory apparatus. The detailed structure of the genitalia is given in Fig. 11. The most important parts of the genitalia on which the identification of species is based are parabasal and other setae on the inner sides of the ganolorites, the claspette lobes with their-setae or clubs, and the single median aedeagus with its paired terminal leaflets. In Indian anophelines, however, genitalia are used in the identification of only An. pinjaurensis (aitkenii complex), in which the aedeagus is very long and large, expanded at opening with a thickened rim and entirely devoid of leaflets or processes.

Female genitalia: Female genitalia have no special characters which can help in the species identification, but they may be of use in the separation of species belonging to subgenus *Anopheles* from subgenus *Cellia*. In subgenus *Anopheles* transparent dots are present all over the spermatheca and a horseshoe-shaped structure is present near the spermatheca, while in subgenus *Cellia* the dots are restricted to certain areas on spermatheca and the horseshoe-shaped structure is absent (Hara, 1959).

EGGS

There are three types of mosquito eggs: (1) those laid singly on the surface of

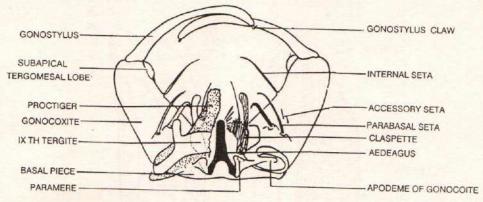


Figure 11: Male genitalia of an anopheline mosquito, true dorsal view (actual ventral after rotation), right half of proctiger and 9th tergite removed.

the water, e.g. *Anopheles*; (2) those glued together to form rafts that float on the water surface, e.g. *Culex, Culiseta, Mansonia*, and *Uranotaenia*; and (3) those laid singly out of water on wet surface, e.g. *Orthopodomyia* and some species of genus *Aedes*. Generally, eggs which are laid on water surface hatch within two to three days. Eggs of some species belonging to genera *Aedes* and *Psorophora* survive up to three to four years on wet grounds till the surface becomes flooded. Eggs of anophelines are white in colour when first deposited on the surface of water but darken after a few hours. The eggs are laid singly on the water surface and lie lengthwise on the water. These eggs are elongated, oval and pointed at one end (boat-shaped), and provided with a pair of lateral floats. The average length of an egg is about 1 mm. The number of eggs laid by a single female anopheline mosquito varies from 80 to 150 depending on the species. The inner and outer walls of the egg are known as endochorion and the exochorion respectively. The general appearance of an anopheline egg is shown in Fig. 12.

The shape, size, inside view of upper surface (concave, convex, or flat), number of floats or ribs, and the shape and width of the deck of the eggs vary from species to species. For example, the width of the deck is very useful to differentiate An. hyrcanus complex from An. barbirostris. The number of the floats on the egg surface also differentiates An. stephensi type form from An. stephensi var. mysorensis and An. stephensi intermediate.

LARVAE

Larvae hatch from the eggs and live in water. Mosquitoes have a strong preference for particular breeding sites. Some larvae develop in permanent waters like canals, ponds, wells, and riverbed pools, and some in temporary waters like pools, rainwater collections, tree holes, or leaf axils. For example, *An. sundaicus* larvae always develop in brackish waters, and *An. minimus* larvae in streams. The larvae get their food from the water and breathe by coming to the surface of water, except *Mansonia* larvae which attach themselves to aquatic plants and breathe from the submerged portion of plants. All mosquito larvae undergo three

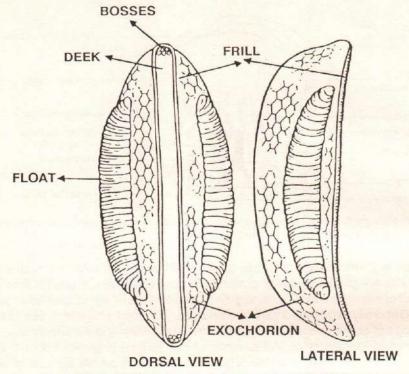


Figure 12: Egg of an anopheline mosquito (Reid, 1968).

successive moultings (ecdysis) during their development (7-10 days, depending upon the environmental factors) when they shed their chitinous skin. The successive moults are known as I, II, III, and IV instars or stages. The IV instar is a mature stage and with the fourth moult the pupa appears.

Generally, there are two types of larval movements: (1) by jerks of the body and by propulsion with mouth brushes, e.g., *Anopheles* and (2) by crawling, e.g., all other culicine larvae. The position of anopheline larvae is parallel to the water surface while culicine larvae hang head down with only the tip of the air tube penetrating the surface film (see Fig. 2).

The anopheline larvae are worm-like and the body of the "wrigglers" can be easily distinguished in three parts: head, thorax, and abdomen (Fig. 13). All these parts contain some important hairs or plates which are used in the identification of the species. The number and position of hairs are important characteristics. The head, which is cylindrical in shape and is covered with a chitinous sheet, consists of antennae in front and eyes lateral to it. The head contains a number of important hairs on the dorsal side (see Fig. 14).

The thorax is formed of pro-, meso-, and metathorax fused together and is thicker and broader than the head or abdomen and somewhat flattened. The thorax also has a number of hairs which are used in identification but there are no other special structures (Fig. 15).

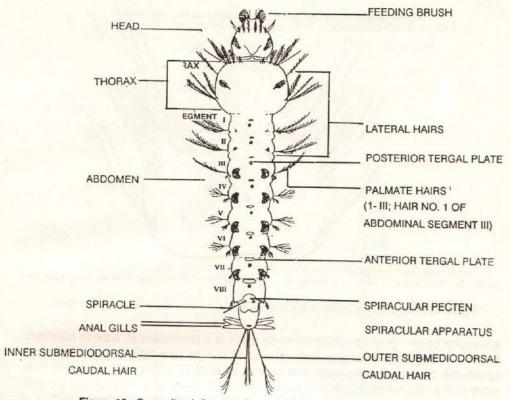


Figure 13: Generalized diagram of an anopheline larva (subgenus Cellia).

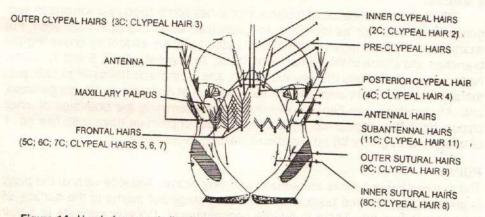


Figure 14: Head of an anopheline larva showing important hair in dorsal view (Puri, 1960).

The abdomen is long and sub-cylindrical and is composed of 10 segments. The structures of the first seven segments are similar but the eighth and ninth segments are considerably modified and form a complicated structure known as spiracular or respiratory apparatus. The respiratory apparatus consists of paired

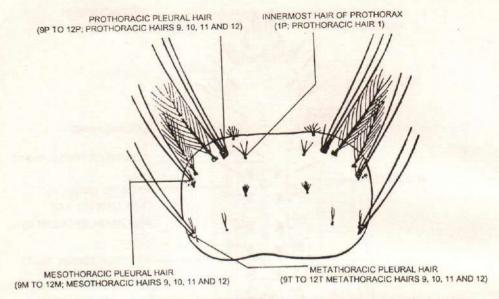


Figure 15: Hair of thorax in dorsal and ventral view of an anopheline larvan (Reid, 1968).

spiracular openings in anophelines while there is a prominent air tube in culicines. The tenth segment is simple and consist of two to four tapering membranous structures towards its posterior end known as anal gills (Fig. 13). The gills help more in the regulation of osmotic pressure than in respiration. The abdomen also consists of very important hairs and tergal plates which are the characteristics of a species.

During the identification of Indian anophelines some species belonging to subgenus Anopheles can be identified only after their larval characters are seen. For example, An. culiciformis can be differentiated from An. sintoni by observing the branches and shape of the frontal hairs (5c, 6c, 7c; clypeal hairs 5, 6 and 7), lateral hairs, subantennal hairs (IIc, clypeal hair II), and prothoracic hairs (9P to 12P; prothoracic hairs 9, 10, 11 and 12). The identification of An. aitkenii, An. bengalensis, and An. insulaeflorum can be confirmed after examining the branches of inner clypeal hairs (2c; clypeal hair 2) and the number of palmate hairs (I-III; hair no. 1 of abdominal segment III) on the abdominal segments 1 to 7.

PUPAE

The pupae of anophelines are aquatic and very active. The side view of the pupa is given in Fig. 16. The pupa is a nonfeeding stage but swims to the surface of the water to breathe. There is a large cephalothorax. Dorsally consists of a pair of trumpets and ventrally the developing mouth parts, eyes, legs, and wings of the adults. The abdomen consists of eight freely movable segments with a pair of paddles at the tip. The pupae move with considerable speed because of the vigorous movement of the abdomen.

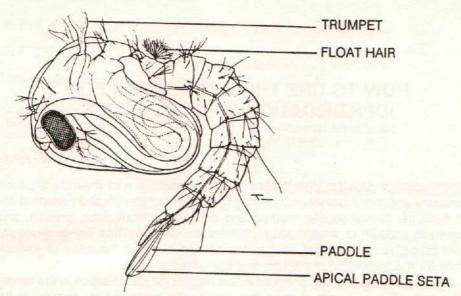


Figure 16: Pupa of an anopheline mosquito in side view (Reid, 1968).

HOW TO USE THE MONOGRAPH FOR IDENTIFICATION OF MOSQUITOES

This monograph provides descriptions of 58 anophelines with their identification characters and morphological variations. The pattern of identification adopted differs from the formal couplet method and provides a more clear, precise, and systematic account to enable easy identification. In using this key to mosquito identification, readers should refer to the section numbers indicated at various places for morphological characters and drawings.

The first character to be checked in a mosquito for identification is the length of the palpi. The palpi are of the same size as the proboscis in *Anopheles* (in females the palpi are rod-shaped, while in males they are club-shaped). The next step is the examination of the presence of the total number of pale areas on the costa, subcosta, and vein 1(R1) on the wings. If the number of pale areas is three or less and the wing is completely dark, the specimen belongs to the subgenus *Anopheles*; if the number of pale areas is four or more it belongs to subgenus *Cellia*. For further identification up to the species level, flow charts (Tables 6 and 7) and diagrams are given for the subgenus *Cellia* is divided into four groups on the basis of palpi ornamentation, and using the morphological features, these groups are further divided.

Twenty-four species of the subgenus *Anopheles* are divided into two main groups on the basis of the presence or absence of pale areas on the wings. In one group the wing is completely dark and in the other it has pale areas. The group with a completely dark wing is divided into two groups which are further grouped into subgroups and finally to the species. The group comprising spotted wings is divided into three groups on the basis of palpi ornamentation and further divided into smaller subgroups leading to species identification. For confirmation of a particular species additional morphological features are given.

For each species the monograph provides a full diagram with main identification characters, name, its derivative, type form available, brief biology, status of insecticide resistance, reported distribution in India and in the world, vector incrimination, and the morphological variations. It contains 209 diagrams of the head, wing, leg, thorax and abdomen, 58 full diagrams, one for each of the species, and 189 illustrations of morphological variations (belonging to 24 species).

Mosquito identification is generally based on female morphological features, but in the case of subgroups, viz. An. bengalensis, An. culiciformis, An. aitkenii, An. insulaeflorum, of the subgenus Anopheles, identification is based on larval characters. The identification of An. pinjaurensis is based on the male genitalia.

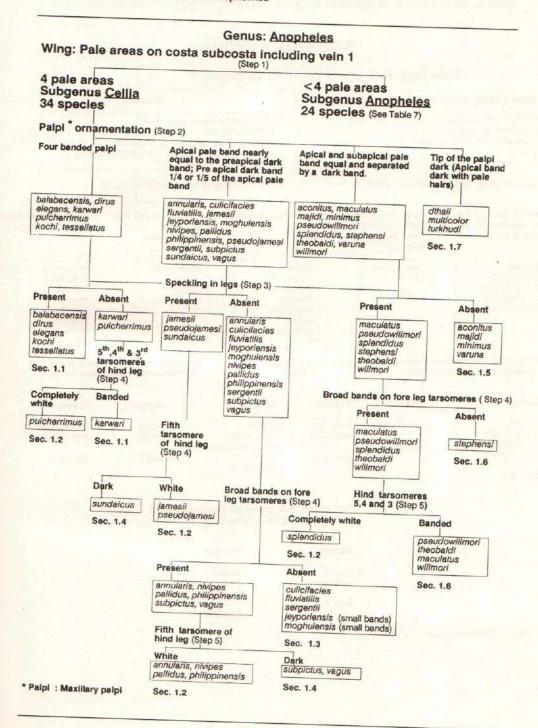
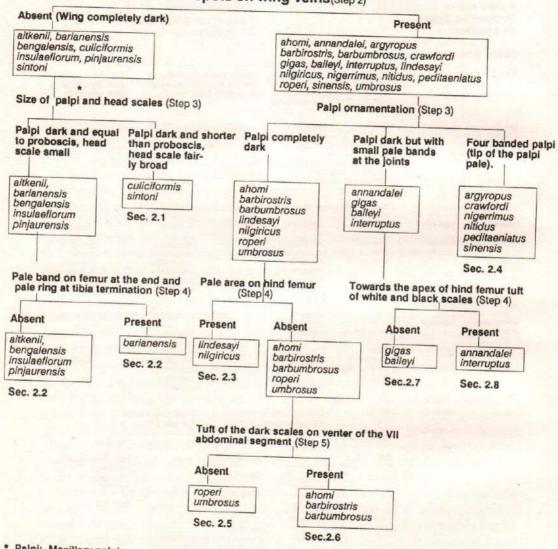


Table 7: Identification of 24 species of subgenus Anopheles

Sub-genus: Anopheles

Pale and dark spots on wing veins(Step 2)

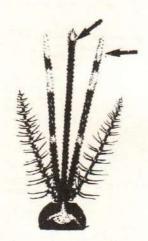


^{*} Palpi: Maxillary palpi

Full drawings of 35 Indian anophelines have been prepared with the help of a brush and Indian ink after measuring their size under a dissecting microscope with an occulometer and stage meter. The 35 species are available in the museum of the Malaria Research Centre. The other 23 anopheline drawings have been prepared by modifying their closely related species with the help of Christophers (1933), Reid (1968) and Harrison and Scanlon (1975).

GENUS ANOPHELES

PALPI LONG, SLENDER (IN FEMALES) OR CLUB-SHAPED (IN MALES) AND EQUAL TO THE PROBOSCIS



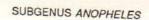


Genus Anopheles comprises subgenera Anopheles, Cellia, Kerteszia, Lophopodomyia, Nyssorhynchus, and Stethomyia. In India only subgenera Anopheles and Cellia are recorded.

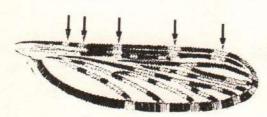
SUBGENUS IDENTIFICATION IS BASED ON WING CHARACTERS. EXAMINE EACH SPECIMEN IN THE FOLLOWING ORDER

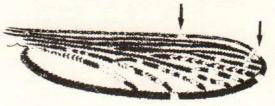
SUBGENUS CELLIA

Costa, subcosta including vein 1(R1) with 4 or more pale areas



Costa, subcosta including vein 1(R1) with 3 or less pale areas





Species in Subgenus Cellia

An. aconitus, An. annularis, An. balabacensis, An. culicifacies, An. dirus, An. dthali, An. elegans, An. fluviatilis, An. jamesii, An. jeyporiensis, An. karwari, An. kochi, An. maculatus, An. majidi, An. minimus, An. moghulensis, An. multicolor, An. nivipes, An. pallidus, An. pseudojamesi, An. pseudowillmori, An. pulcherrimus An. philippinensis, An. sergentii, An. splendidus, An. stephensi, An. subpictus, An. sundaicus, An. tessellatus, An. theobaldi, An. turkhudi, An. vagus, An. varuna, An. willmori

Species in subgenus Anopheles

An. ahomi, An. aitkenii, An. annandalei, An. argyropus, An. baileyi, An. barianensis, An. barbirostris, An. barbumbrosus, An. bengalensis, An. crawfordi, An. culiciformis, An. gigas, An. insulaeflorum, An. interruptus, An. lindesayi, An. nigerrimus, An. nilgiricus, An. nitidus, An. peditaeniatus, An. pinjaurensis, An. roperi, An. sinensis, An. sintoni, An. umbrosus

1. Subgenus Cellia

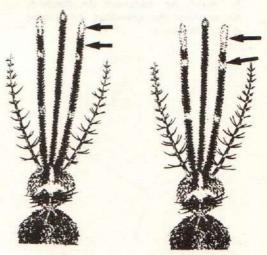
IDENTIFICATION IS BASED ON PALPI ORNAMENTATION

Four-banded palpi



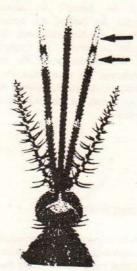
An. kochi, An. karwari, An. tessellatus, An. balabacensis, An. dirus, An. elegans, An. pulcherrimus

Apical pale band nearly equal or equal to the pre-apical dark band (intervening dark band); pre-apical dark band 1/4 or 1/5 of the apical pale band.



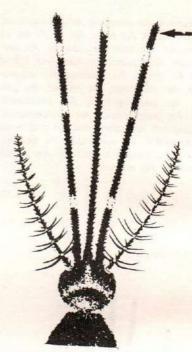
An. culicifacies, An. fluviatilis, An. sergentii, An. moghulensis, An. jeyporiensis, An. subpictus, An. vagus, An.sundaicus, An. annularis, An. pallidus, An. philippinensis, An. nivipes, An. jamesii, An. pseudojamesi

Apical and subapical pale band equal and separated by a small or same-sized dark band



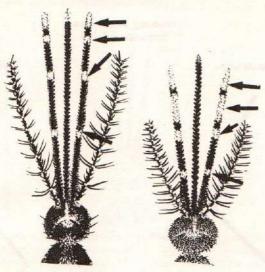
An. minimus, An. varuna, An. aconitus, An. majidi, An. maculatus, An. theobaldi, An. stephensi, An. splendidus, An. willmori, An. pseudowillmori

Tip of palpi dark



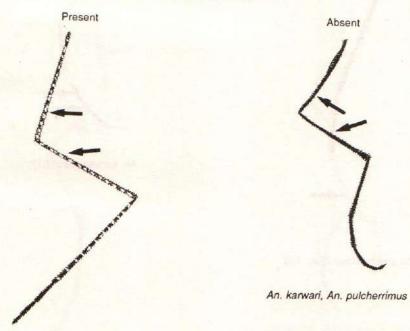
An. turkhudi, An. multicolor, An. dthali (Sec. 1.7)

Four-banded palpi



An. kochi, An. karwari, An. tessellatus, An. balabacensis, An. dirus, An. elegans An. pulcherrimus

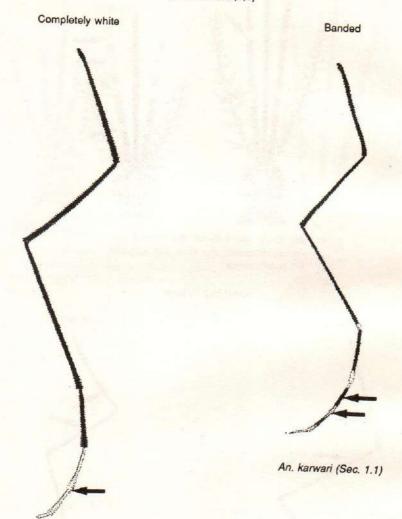
Speckling in legs



An. kochi, An. elegans, An. tessellatus An. dirus, An. balabacensis (Sec. 1.1)

An. pulcherrimus, An. karwari

Hind-leg tarsomeres (tarsomeres 5,4,3)



An. pulcherrimus (Sec. 1.2)

Apical pale band nearly equal or equal to the pre-apical dark band (intervening dark band); pre-apical dark band 1/4 or 1/5 of the apical pale band



An. culicifacies, An. fluviatilis, An. sergentii, An. moghulensis, An. jeyporiensis, An. subpictus, An. vagus, An. sundaicus, An. nivipes, An. annularis, An. pallidus, An. philippinensis, An. jamesii, An. pseudojamesi

Speckling in legs

1

Present

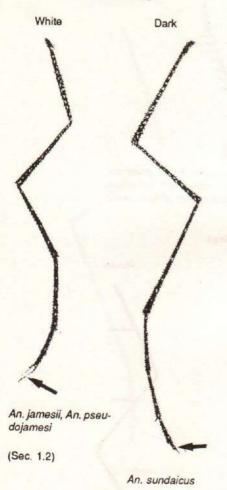
An. sundaicus, An. jamesii, An. pseudojamesi



An. culicifacies, An. fluviatilis, An. sergentii An. moghulensis, An. jeyporiensis, An. subpictus, An. vagus, An. annularis, An. pallidus, An. philippinensis, An. nivipes

An. sundaicus, An. jamesii, An. pseudojamesi

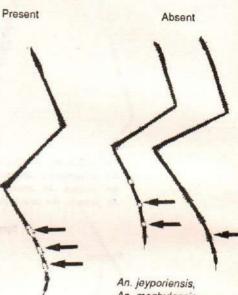
Fifth tarsomere of hind-leg



(Sec. 1.4.)

An. culicifacies, An. fluviatilis, An. sergentii, An. moghulensis, An. jeyporiensis, An. subpictus, An. vagus, An. annularis, An. pallidus, An. philippinensis, An. nivipes

Bands on foreleg tarsomeres



An. subpictus,
An. vagus,
An. annularis,
An. pallidus,
An. nivipes,
An. philippinensis

An. moghulensis (Bands small in size), An. culicifacies, An. fluviatilis, An. sergentii

(Sec. 1.3)

An. subpictus, An. vagus, An. annularis, An. pallidus, An. philippinensis, An. nivipes

Fifth tarsomeres of hind leg

White (tarsomeres 5,4,3) completely white



An. annularis, An. pallidus, An. philippinensis, An. nivipes

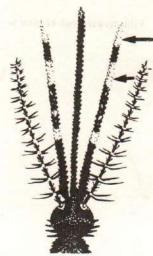
(Sec. 1.2)



An. subpictus, An. vagus

(Sec. 1.4)

Apical and subapical pale band equal and separated by a small or same-sized dark band

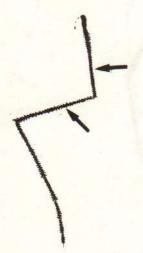


An. minimus, An. varuna, An. aconitus, An. majidi, An. maculatus, An. theobaldi, An. stephensi, An. splendidus, An. willmori, An. pseudowillmori

Speckling in legs

Absent

Present



An. minimus, An. varuna, An. aconitus, An. majidi



An. maculatus, An. theobaldi, An. willmori, An. pseudowillmori, An. splendidus, An. stephensi An. minimus, An. varuna, An. aconitus An. majidi

Bands on foreleg tarsomeres

Absent Present

An. majidi

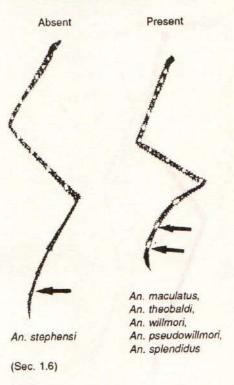
(Sec. 1.5)

An. minimus, An. varuna, An. aconitus

(Sec. 1.5)

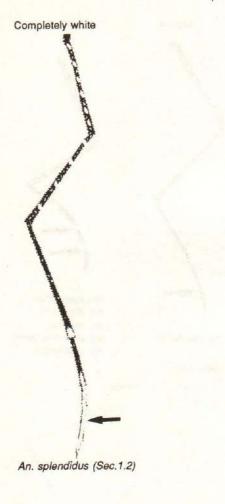
An. maculatus, An. theobaldi, An. willmori, An. pseudowillmori, An. splendidus, An. stephensi

Bands on foreleg tarsomeres



An. maculatus, An. theobaldi, An. willmori An. pseudowillmori, An. splendidus

Hind-leg tarsomeres (tarsomeres 5,4,3)





An. maculatus, An. theobaldi, An. willmori, An. pseudowillmori

(Sec. 1.6)

1.1 An. kochi, An. balabacensis, An. dirus, An. elegans, An. karwari, An. tessellatus, An. pulcherrimus

Wing with 4 or more pale areas on the costa, subcosta including vein 1 (R-1) Four-banded palpi

A big white spot on the junction of tibiotarsal joint of hind leg



An. balabacensis, An. dirus, An. elegans

Other characters

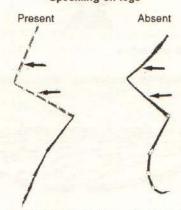
Legs with speckling. Foreleg tarsomeres with broad pale bands



An. tessellatus, An. kochi, An. karwari, An. pulcherrimus*

Speckling on legs

Absent



An. tessellatus, An. kochi

An. karwari

^{*} Further identification is given in Sec. 1.2

An. balabacensis, An. dirus, An. elegans Other characters

Wing vein 6(Anal) with more than 6 dark spots and similar spots on veins 5(Cu) and 5.1(Cu1)

Fringe spot absent on veins 6(Anal) and 5(Cu) but present between these two veins.



An. balabacensis, An. dirus, An. elegans

Length of the proboscis

Proboscis quite long, longer than fore-femur



Other characters

Pre-sector dark spot of vein 1(R1) completely dark

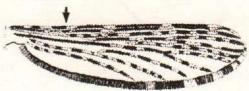


Proboscis normal in size (nearly equal to forefemur)

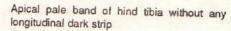


Other characters

Pre-sector dark spot of vein 1(R1) with pale interruption

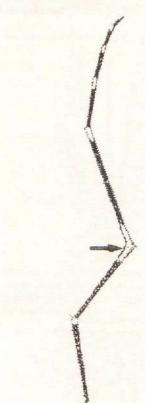


Apical pale band of hind tibia with longitudinal dark strip on ventral side





An. elegans



An. balabacensis, An. dirus

Wing vein 1(R1)

Pre-sector dark mark of vein 1(R1) extended basally beyond the end of the corresponding marking on the costa, but not reaching up to the distal end of humeral dark spot



An. balabacensis

Pre-sector dark mark of vein 1(R1) extending basally beyond the distal end of the humeral dark mark on the costa



An. dirus

An. tessellatus, An. kochi

Banding on hind-leg tarsomeres

Narrowly banded

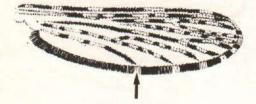


Other characters

Apical half of proboscis yellow



Wing as in An. dirus but fringe spot present on vein 6(Anal)



An. tessellatus

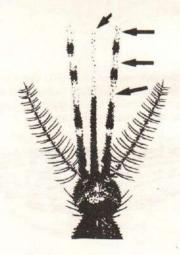
Narrowly banded but remaining joints with broad pale bands



An. kochi

Other characters

Apical 3 broad pale bands separated by 2 small dark bands; apical half of proboscis yellow



Other characters

An. kochi

Vein 6(Anal) with three small dark areas and inner costa interrupted



Venter of the abdomen with prominent tuft of black scales on each segment e.g., from 2 to 7



An. karwari

Banding pattern of palpi as given in figure

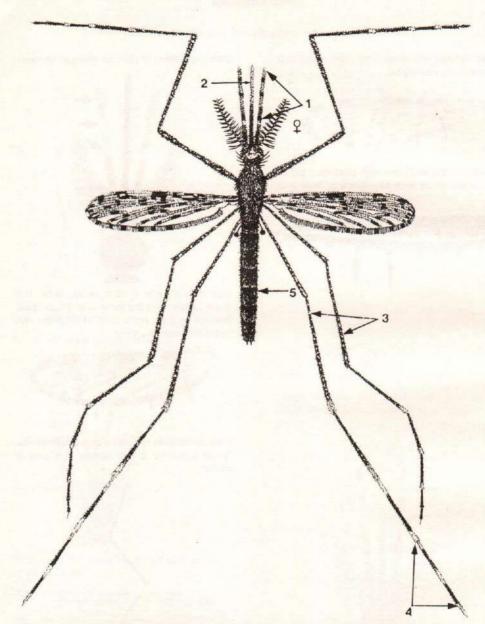


Vein 6(Anal) with 2 dark areas; outer half much longer than the inner one; fringe spots present on wing veins 5.1(Cu1), 6(Anal) and between these two veins



Hind tarsomeres with three broad pale bands. Tarsal segments 5 and portion of 4 pale in colour





An. kochi Doenitz 1901, main identification characters

- 1. Four banded palpi.
- 2. Half of the proboscis yellow in colour.
- 3. Legs with speckling.
- 4. Hind leg tarsomeres banded as shown in figure.
- 5. Venter of the abdomen with tuft of black scales.

	Anopheles kochi					
Name	:	An. kochi Doenitz, 1901. Insektenborse 18: 36-38.				
Derivative		Doenitz gave the name at the Berlin meeting of the entomological club after his Professor Heinrich Hermann Robert Koch (1843–1910), the famous bacteriologist.				
Type form	:	Available at the Zoologisches Museum, Humboldt University, Berlin.				
Sitting posture	:	Body and mouth parts at an angle to resting surface.				
Resting habit	:	Adults rest mainly outdoors but sometimes in houses and cattlesheds also.				
Breeding ecology	:	Breeds in shallow muddy water collections, ground pools with or without grass, hoof marks, fallow ricefields, also in streams of jungles.				
Biting time	•	Before midnight, preferably in the first quarter.				
Feeding preference	Feeding preference: Prefers feeding on cattle to feeding on man. Also f on monkeys.					
Flight range		About 1 km.				
Susceptibility to insecticides	:	NA				
Relation to disease	:	Not an important vector of human malaria, although some gut- and gland-infected specimens were found in India.				
Reported distribution	:	Widely found in oriental region from Bangladesh, South China, India, Indonesia, Malaysia, Myanmar, Nepal, and the Philippines. In India, recorded from Arunachal Pradesh, Andaman Islands, Assam, Manipur, Meghalaya, Mizoram, Tripura, Uttar Pradesh, and West Bengal.				
Vector incrimination	:	Results of studies made so far are summarized in the table.				

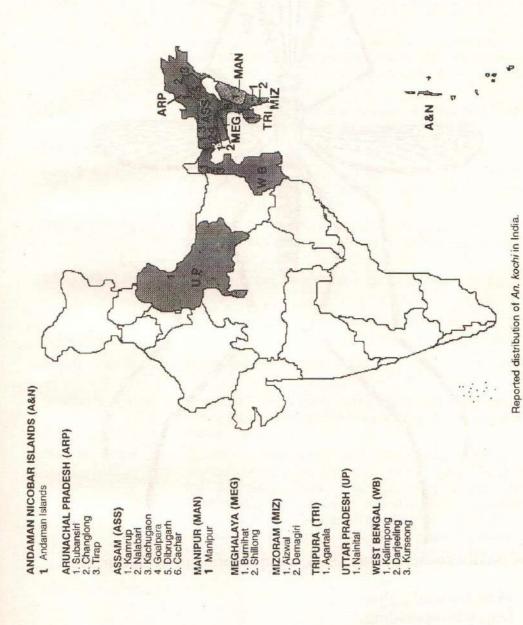
Table: Anopheles kochi: Results of vector incrimination studies

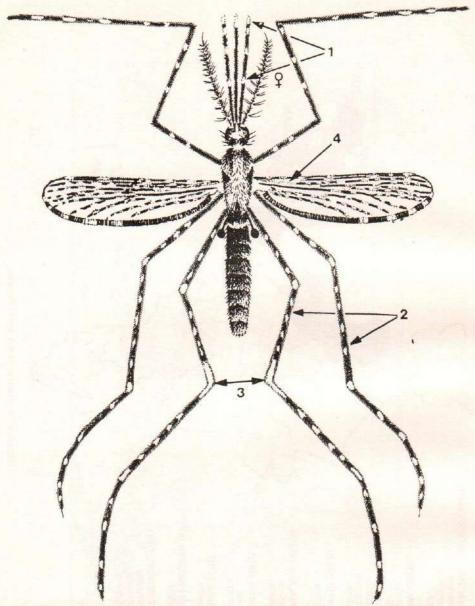
SI. No.	Year	Author	Locality	Number dissected	Number Positive			
					Gut	Gland	Total	
1.	1929	Strickland	Cachar, Assam	535	1	0	1	
2.	1930	Ramsay	Assam	2094	2	0	2	
3.	1933	Manson & Ramsay	Jorhat, Assam	NM	1	0	1	

NM-Not mentioned.



Reported distribution of An. kochi in the World.



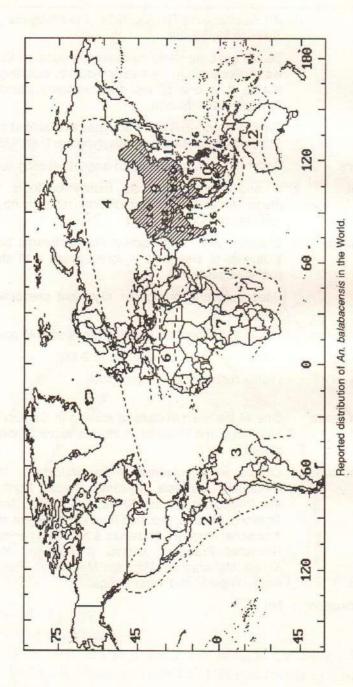


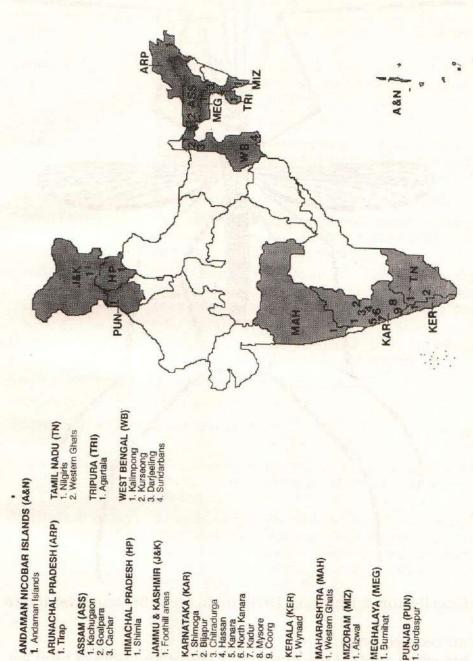
An. balabacensis Baisas 1936, main identification characters

- 1. Four banded palpi.
- 2. Legs with speckling.
- 3. A big white spot at the junction of femur and tibia of hind leg.
- 4. Pre-sector dark mark on vein 1 (R1) not extending basally up to humeral dark mark of costa.

Anopheles balabacensis				
Name	:	An. balabacensis Baisas, 1936. The Philippine Journal of Science 59: 65-84.		
Derivative	:	Baisas gave the name balabacensis because the species was collected from the western islands including Balabac Island. It is one of the most westerly islands in the Philippine archipelago.		
Type form	:	Type form non-existent. Paratypes available at the United States National Museum, Washington, D.C., USA.		
Sitting posture	:	Body and mouth parts at an angle to resting surface.		
Resting habit	:	A wild species, it rests mostly outdoors in forest areas (exophilic) but infrequently rests in houses and cattlesheds during the day.		
Breeding ecology	:	Breeds in pools by the side of mainly streams, borrowpits, footprints of elephants in forest areas with shade and rotting vegetations.		
Biting time	•	Bites outdoors throughout night but predominantly at midnight.		
Feeding preference	:	Highly anthropophilic but also feeds on wild animals.		
Flight range	:	About 1 km and dispersal up to 2 km.		
Susceptibility to insecticides	:	Highly susceptible to insecticides.		
Relation to disease		One of the main vectors of malaria in Cambodia, India, Indonesia, and Thailand. In India, a vector in northeastern states.		
Reported distribution		Occurs in Bangladesh, India, Indochina, Indonesia, Cambodia, Malaysia, Myanmar, Nepal, North Borneo, the Philippines, South China, Sri Lanka, Taiwan and Thailand. In India, reported from northeastern states and Arunachal Pradesh, Andaman & Nicobar Islands, Assam, Himachal Pradesh, Jammu & Kashmir, Karnataka, Kerala, Maharashtra, Mizoram, Meghalaya, Punjab, Tamil Nadu, Tripura, and West Bengal.		

Vector incrimination: NA





MIZORAM (MIZ)

1. Aizwal

PUNJAB (PUN) 1. Gurdaspur

KERALA (KER)
1. Wynaad

1. Shimoga 2. Bijapur 3. Chitradurga 4. Hassan

5. Kanara 6. North Kanara

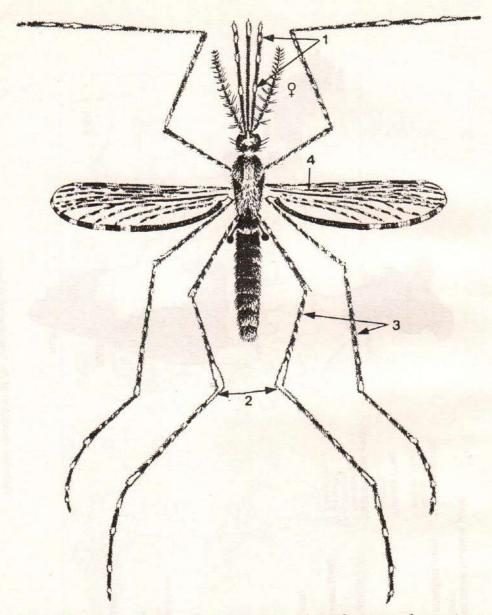
8. Mysore 9. Coorg

7. Kadur

ASSAM (ASS)
1. Kachugaon
2. Goalpara
3. Cachar

1. Tirap

Reported distribution of An. balabacensis in India.



An. dirus Peyton & Harrison 1979, main identification characters

- 1. Four banded palpi.
- 2. A big white spot at the junction of femur and tibia of hind leg.
- 3. Legs with speckling.
- 4. Pre-sector dark mark on vein 1 (RI) basally extended up to humeral dark mark of costa.

		Anopheles dirus*
Name	:	An. dirus Peyton and Harrison, 1979. Mosquito Systematics 11: 40-49.
Derivative	:	Latin, dirus, fearful, horrible, dire. Peyton and Harrison chose the name dirus to reflect high vector efficacy in the transmission of malaria.
Type form	:	Location not known.
Species complex	;	Seven sibling species (known as A, B, C, D, E, F and takasagoensis); species E is only recorded from western India, all other species are recorded from Thailand.
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit		Wild species, infrequently present inside houses and cattlesheds during day. Adults can be collected at night on human and cattle bait collections.
Breeding ecology	•	A pool breeder in dense jungles. Also breeds in pools by the side of rocky streams, disused wells, rain-water collections, borrowpits, drains in densely-shaded foliage and stagnant waters in jungles.
Biting time	:	Exophagic, bites from 19.00 to 4.00 hrs but the peak biting activity is between 22.00 and 2.00 hrs.
Feeding preference		Highly anthropophilic but also bites cattle.
Flight range		1 to 1.5 km.
Susceptibility to insecticides	:	Susceptible to DDT.
Relation to disease	:	Efficient vector of human malaria in Bangladesh, East Borneo, India, Cambodia, Myanmar, Sumatra and Thailand.
Reported distribution	:	Occurs in Bangladesh, India, Indonesia (Java), Malaysia, North Myanmar, Myanmar, and Thailand. In India, recorded from Arunachal Pradesh, Andaman Islands, Assam, Karnataka, Kerala, Meghalaya, Mizoram, Tamil Nadu, Tripura, and West Bengal.
Vector incrimination	:	Results of studies made so far are summarized in the

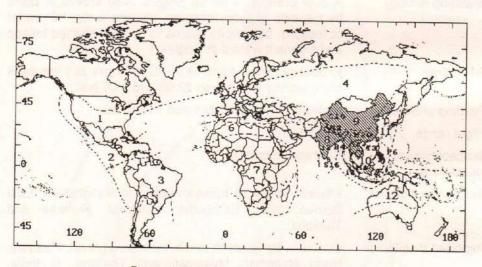
table.

^{*}In India previously it was misidentified as An. balabacensis but recent studies show that the species is An. dirus.

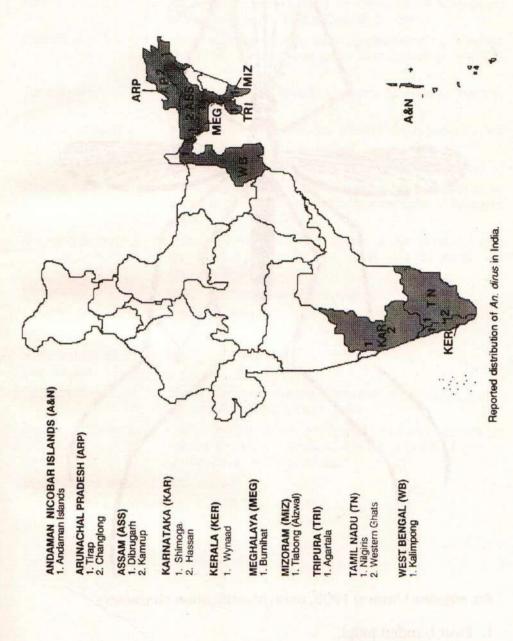
Table: Anopheles balabacensis*: Results of vector incrimination studies

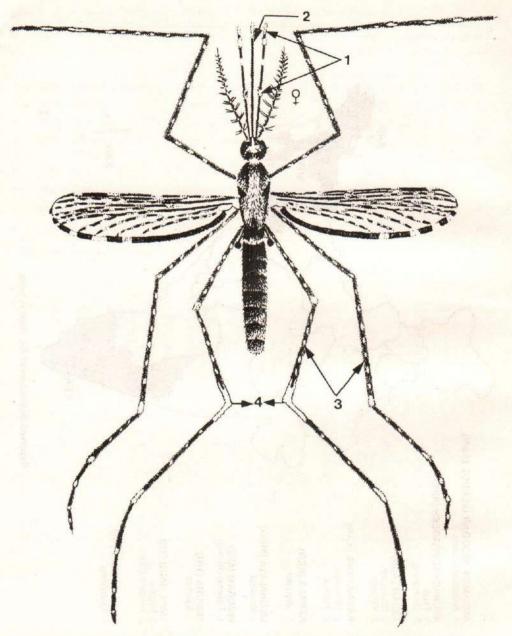
SI. No.	1505/	Author	Locality	Number	Number Positive			
					Gut	Gland	Total	
1.	1941	Clark & Choudhury	Digboi area, Assam	859	8	21	29	
2.	1948	Macan	Assam	92	_	_	1	
3.	1973	Sen et al.	Tirap, Arunachal Pradesh	1811	0	3	3	
4.	1984	Annual Report, NMEP	Boko, Assam	199	0	1	1	
5.	1985	Das and Baruah	Mizoram	134	0	1	1	
6.	1989	Dutta et al.	Dibrugarh, Assam	74	0	1	1	
7.	1989	Dutta et al.	Changlang, Arunachal Pradesh	609	0	7	7	
8.	1990	Das et al.	Tlabong, South Mizoram	178	0	2	2	

^{*} Now identified as An. dirus.



Reported distribution of An. dirus in the World.



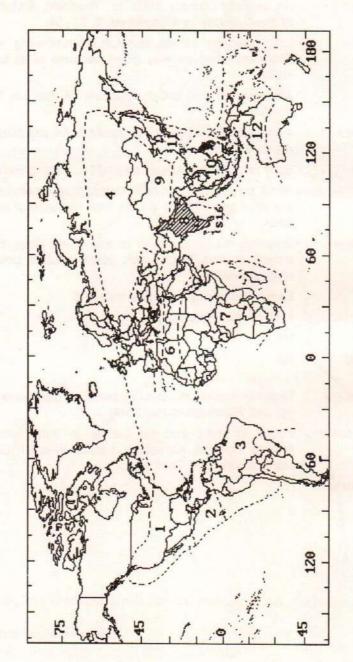


An. elegans (James) 1903, main identification characters

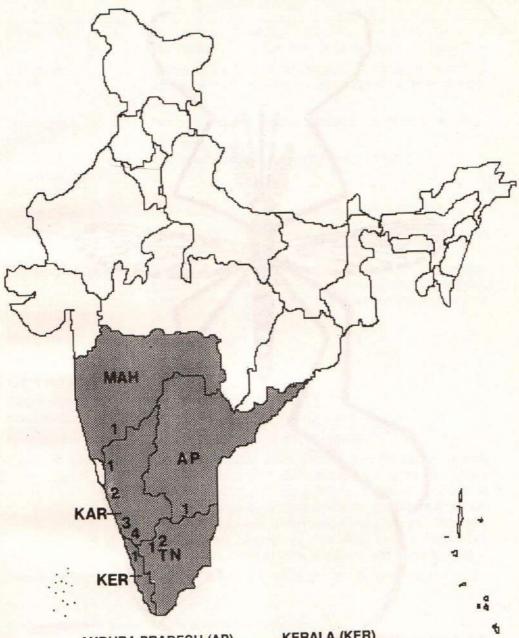
- 1. Four banded palpi.
- 2. Proboscis longer than the fore femur.
- 3. Legs with speckling.
- 4. A big white spot at the junction of femur and tibia of hind leg.

		Anopheles elegans
Name	:	An. elegans (James), 1903. In: Theobald, A Monograph of the Culicidae or Mosquitoes, 3: 51-54.
Derivative	:	Latin, <i>elegans</i> , choice, tasteful, discriminating, elegant. The name <i>elegans</i> was given because of its beautiful looks.
Type form	:	Available at the British Museum of Natural History, London.
Species complex	:	4 species (An. elegans, An. hackeri, An. pujutensis, and An. sulawesi).
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Rests only outdoors in dark, damp, shady places such as arecanut gardens and shrubs near the base of arecanut trees.
Breeding ecology		Breeding habitat is similar to that of <i>An. dirus</i> , but also breeds in tree holes, rock pools, ground pools and stagnant waters.
Biting time	:	Exophagic and bites before midnight.
Feeding preference	:	Prefers monkey blood.
Flight range	:	NA
Susceptibility to insecticides		NA
Relation to disease	•	Vector of monkey malaria (<i>Plasmodium</i> disease <i>cynomologi</i> and <i>Plasmodium inui</i>) only.
Reported distribution	:	Found in India and Sri Lanka. In India, occurs in Andhra Pradesh, Karnataka, Kerala, Western Ghats in Maharashtra, and Tamil Nadu.

Vector incrimination : NA



Reported distribution of An. elegans in the World.



ANDHRA PRADESH (AP)

1. Chittoor

KARNATAKA (KAR) 1. North Kanara

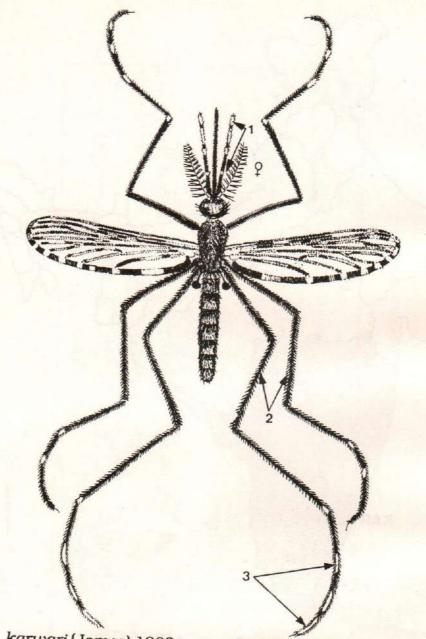
- 2. Shimoga 3. Hassan 4. Mysore

KERALA (KER) 1. Wynaad

MAHARASHTRA (MAH) 1. Western Ghats

TAMIL NADU (TN) 1. Nilgiris 2. Western Ghats

Reported distribution of An. elegans in India...



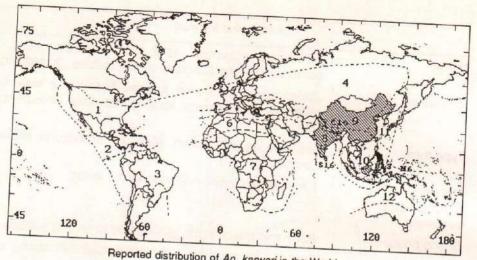
An. karwari (James) 1903, main identification characters

- Four banded palpi.
 Legs without speckling.
- 3. Hind leg tarsomeres banded as shown in figure.

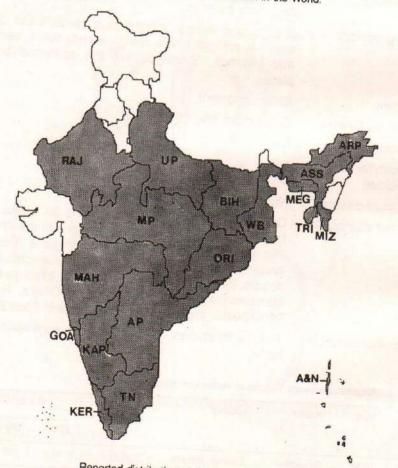
		Anopheles karwari
Name	:	An. karwari (James), 1902. In: Theobald, 1903. A Monograph of the Culicidae or Mosquitoes, 3: 102-104.
Derivative	:	James gave the name <i>karwari</i> because it was first collected from Karwar locality (Bombay Presidency), Goa.
Type form	:	Available at the British Museum of Natural History, London.
Reported morphological variations	:	1 in maxillary palpi, 1 in wing and 1 in leg
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Rests in human dwellings and cattlesheds during day. Also rests outdoors.
Breeding ecology	:	Breeds in a variety of habitats such as clear- shaded streams, springs, pools, drains, weedy tanks, seepages, and swamps. Seepages are the preferred breeding places.
Biting time	:	Before midnight.
Feeding preference	:	Cattle more often than man.
Flight range	:	NA
Susceptibility to insecticides	:	NA
Relation to disease	:	Not regarded as a vector of malaria.
Reported distribution	1:	Occurs in Borneo, New Guinea, and Sri Lanka extends from India excluding northwest regions to the Philippines and north China to Hong Kong. In India it occurs in Andaman Islands, Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Goa, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Meghalaya, Mizoram, Orissa, Rajasthan, Tamil Nadu, Tripura, Uttar Pradesh, and West Bengal.
Vector incrimination	:	Results of studies made so far are summarized in the table.

Table: Anopheles karwari: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive			
					Gut	Gland	Total	
1	1929	Strickland	Cachar, Assam	1697	1	. 0	1	



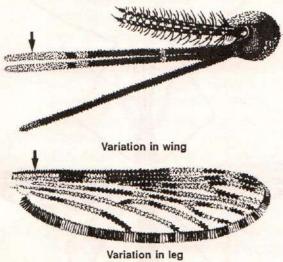
Reported distribution of An. karwari in the World.



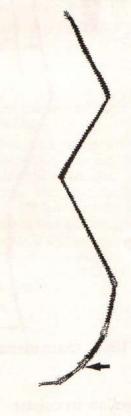
Reported distribution of An. karwari in India.

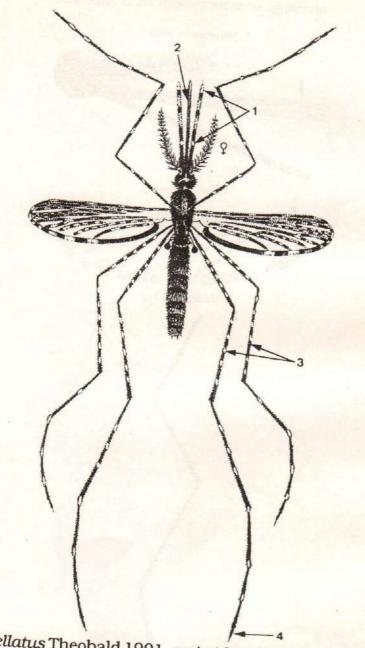
REPORTED IMPORTANT VARIATIONS OF An. karwari

Variation in palpi









An. tessellatus Theobald 1901, main identification characters

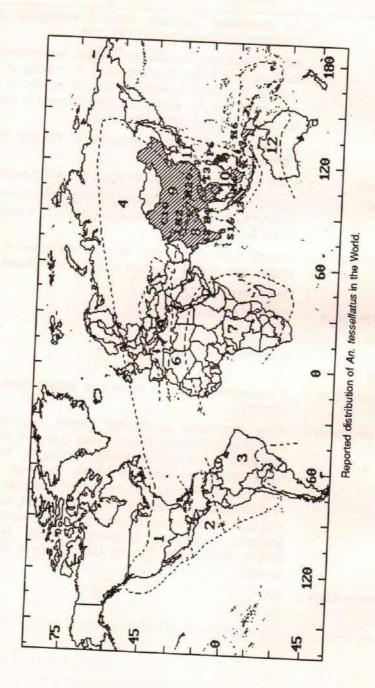
- 1. Four banded palpi
- 2. Half of the proboscis yellow in colour
- 3. Legs with speckling.
- 4. Hind leg tarsomeres 5 completely dark but sometimes white.

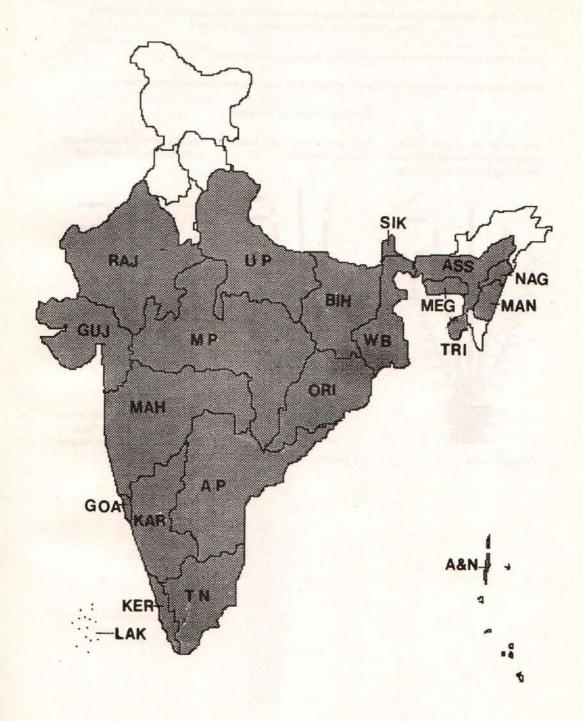
		Anopheles tessellatus
Name	:	An. tessellatus Theobald, 1901. A Monograph of the Culicidae or Mosquitoes, 1: 175–176.
Derivative	:	Latin, tessellatus, mosaic, checkered, tessellated; (tessella, a small cube of stone used in making of pavements). Theobald likened the thoracic pattern to a mosaic, much like An. kochi., but differs from it in the more tessellated appearance of the thorax.
Type form	:	Available at the British Museum of Natural History, London.
Sitting posture		Body and mouth parts at an angle to resting surface.
Resting habit	:	Adults rest in houses, mixed dwellings, cattlesheds and outdoors. The species rests particularly on the lower parts of the walls of houses.
Breeding ecology	:	Breeds in wells, ricefields, or channels either in sun or shade; also in dirty stagnant waters under shade.
Biting time	:	Before midnight.
Feeding preference		Cattle in preference to man.
Flight range	:	NA
Susceptibility to insecticides	:	Susceptible to DDT.
Relation to disease	:	A suspected vector of malaria in India (Lakshadweep Islands).
Reported distribution	n :	Wide occurrence in oriental region; recorded from Bangladesh, Borneo, South China, Hong Kong, India, Indonesia, Malaysia, Maldives, Moluccas, Myanmar, Nepal, the Philippines, Sri Lanka, Sulawesi, Taiwan, and Thailand. In India, recorded from Andaman Islands, Andhra Pradesh, Assam, Bihar, Goa, Gujarat, Karnataka, Kerala, Lakshadweep, Madhya Pradesh, Maharashtra,

Manipur, Meghalaya, Nagaland, Orissa, Rajasthan, Sikkim, Tamil Nadu, Tripura, Uttar Pradesh, and West

Vector incrimination : NA

Bengal.





Reported distribution of An. tessellatus in India.

1.2 An. splendidus, An. pulcherrimus, An. jamesii, An. pseudojamesi, An. annularis, An. pallidus, An. philippinensis, An. nivipes

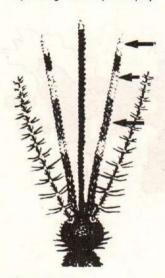
Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1)

Hind tarsus with a continued white area embracing at least the two terminal tarsomeres

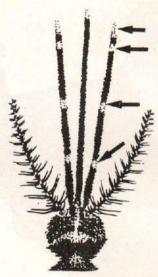
Banding pattern of the palpi

Two broad apical pale bands separated with a small dark band. Speckling in lower part of palpi Palpi with 4 small pale bands (four-banded palpi)

Apical pale band nearly equal to the pre-apical dark band



An. splendidus



An. pulcherrimus



An. jamesii, An. pseudojamesi, An. annularis, An. pallidus, An. philippinensis, An. nivipes

An. splendidus

Other characters

Tibia and femur with speckling

Tarsomeres of hind legs 5, 4, 3, and 20% of 2 white in colour

An. pulcherrimus

Other characters

Tibia and femur without speckling but with more white areas

Tarsomeres of hind legs 5, 4, 3, and 50% of 2 white in colour

An. jamesii, An. pseudojamesi, An. annularis, An. pallidus, An. philippinensis, An. nivipes

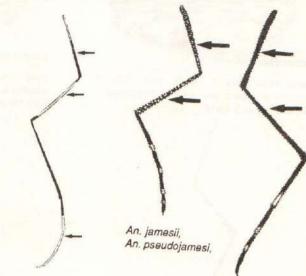
Speckling in legs

Present

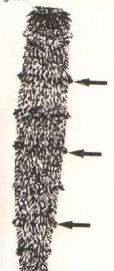
Absent



General appearance dark and moderate size



Abdomen densely clothed with white-scales sometimes forming tuft



An. annularis, An. pallidus,

An. philippinensis,

An. nivipes,

An. pseudojamesi, An. jamesii

At the bifurcation of wing vein 5(Cu) and inner costa

At the bifurcation of wing vein 5(Cu) the area is pale; base of the costa mainly dark



Other characters

Hind tarsomeres 5, 4, 3, and 20% of tarsomere 2 pale but apex of tarsomere 1 without any pale band



Abdomen clothed with golden hairs



Golden and medium-size mosquito An. jamesii

At the bifurcation of the 5(Cu) vein the area is dark; base of the costa interrupted



Other characters

Hind tarsomeres 5, 4, 3, and 20% of tarsomere 2 pale but apex of tarsomere 1 with pale band



Abdomen without golden hairs



Dark and small-size mosquito

An. pseudojamesi

An. annularis, An. pallidus, An. philippinensis, An. nivipes

At the bifurcation of wing vein 5(Cu)

The area is dark



Other characters

Hind tarsomeres 5, 4, 3, and 30% of 2 pale in colour. At the apex of 1st tarsomere of hind leg a well-marked pale band is present

The area is pale



An. pallidus, An. philippinensis, An. nivipes



Medium size and dark in colour

An. annularis

An. pallidus, An. philippinensis, An. nivipes

Hind tarsomeres

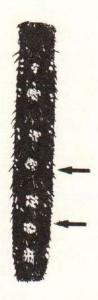
Hind tarsomeres 5, 4, 3, and 70% of tarsomere 2 pale. At the apex of 1st tarsomere of hind leg pale band absent



Hind tarsomeres 5, 4, 3, and 50% of tarsomere 2 pale. At the apex of 1st tarsomere of hind leg a small pale band present



Most of the venter with scattered pale scales



Medium size and light yellow in colour

An. pallidus

Pale scales restricted to last few apical segments of the venter



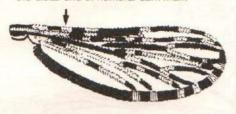
Medium size and dark in colour

(An. philippinensis, An. nivipes)

An. philippinensis, An. nivipes

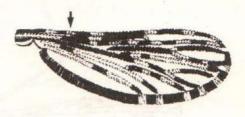
Pre-sector dark mark of vein 1(R1)

Pre-sector dark mark of vein 1(R1) extended basally beyond the end of the corresponding marking on the costa, but not reaching up to the distal end of humeral dark mark

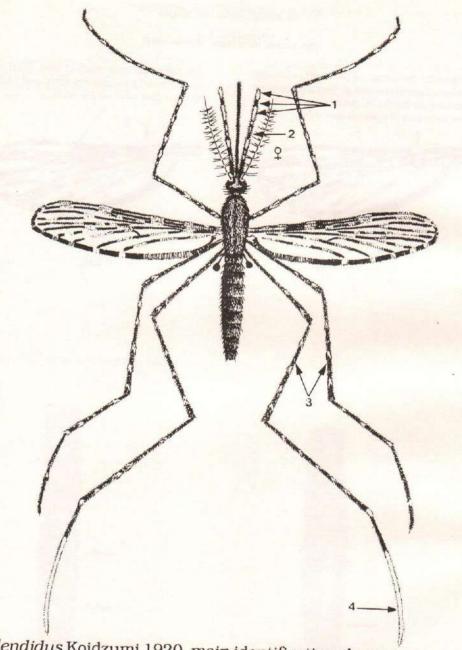


An. philippinensis

The pre-sector dark mark of vein 1(R1) extending basally beyond the distal end of the humeral dark mark on the costa



An. nivipes



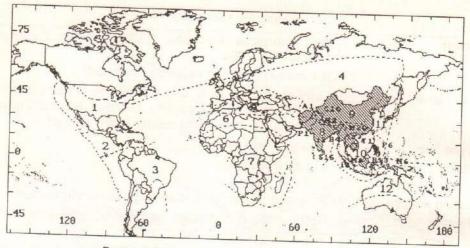
An. splendidus Koidzumi 1920, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band.
- 2. Palpi with speckling.
- 3. Legs with speckling.
- 4. Hind tarsomeres 5, 4 and 3 atleast completely white.

		Anopheles splendidus				
Name	:	An. splendidus Koldzumi, 1920. Daiwan Kenkyujo Hokoku, 8: 1–158.				
Old name	:	An. maculipalpis var. indiensis, Theobald.				
Derivative		Latin, splendidus, clear, bright, spotlessly or transparently brilliant, gleaming, showy, fine, beautiful. Koidzumi gave the name because of the beauty of this species.				
Type form	:	Location not known.				
Reported morphological variations	:	4 in maxillary palpi				
Sitting posture		Body and mouth parts at an angle to resting surface.				
Resting habit	:	Rests indoors in houses and cattlesheds; also rests outdoors in forest areas.				
Breeding ecology		Breeds in pools with algal vegetation in riverbeds, clear pools in riverbeds, jungle streams, ponds with aquatic vegetation, tanks with weeds, marshy lake margins, and tree holes.				
Biting time		Before midnight.				
Feeding preference	:	Primarily a cattle feeder but may occasionally bite man.				
Flight range	:	1 to 1.5 km.				
Susceptibility to insecticides	:	Resistant to dieldrin.				
Relation to disease	;	No role in malaria transmission.				
Reported distribution		Occurs in Afghanistan, Bangladesh, China, India, Indochina, Myanmar, Nepal, Pakistan, Taiwan, and Thailand. In India, found in Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Delhi, Goa, Gujarat, Haryana, Himachal Pradesh, Karnataka, Kerala, Jammu & Kashmir, Madhya Pradesh, Maharashtra, Manipur, Meghalaya, Orissa, Punjab, Rajasthan, Sikkim, Tamil Nadu, Uttar Pradesh, and West Bengal.				
Vector incrimination	:	Results of studies made so far are summarized in the table.				

Table: Anopheles splendidus: Results of vector incrimination studies

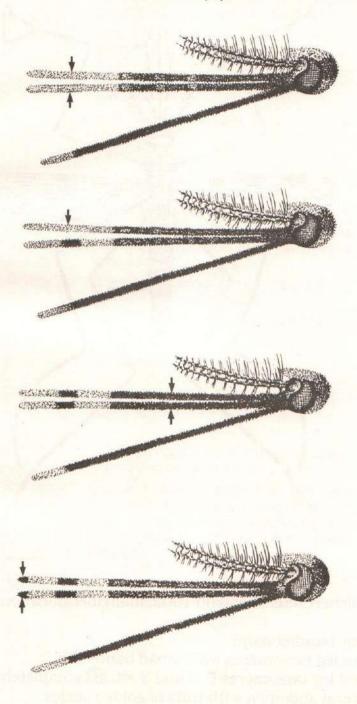
SI. Yea	Year	Author	Locality	Number dissected	Number Positive			
					Gut	Gland	Total	
1.	1931	Macdonald & Majid	Karnal, Haryana	26	1	0	1	
2.	1935	Senevet	India	289	1	0	1	
3.	1944	Covell	India	26	_	-	1	
4.	1948	Subramanian & Dixit	Khandwa, Madhya Pradesh	9	0	9	9	

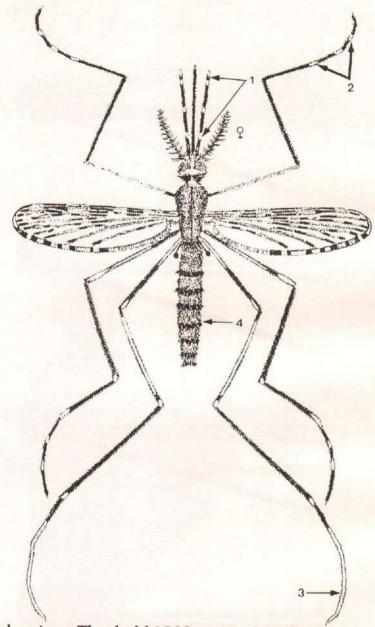


Reported distribution of An. splendidus in the World.



REPORTED IMPORTANT VARIATIONS OF An. splendidus

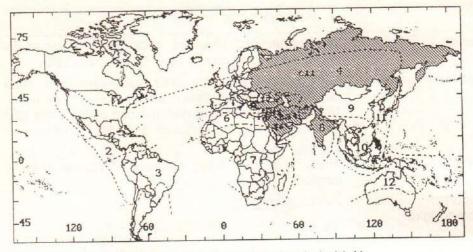




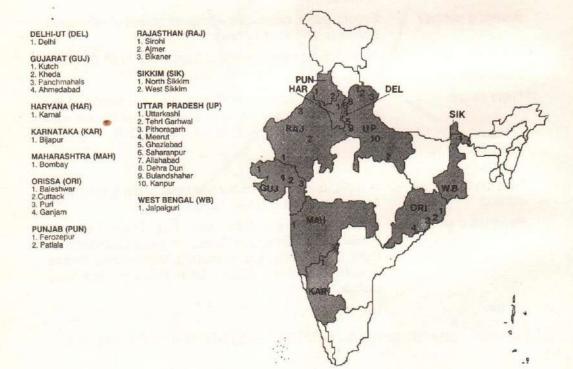
An. pulcherrimus Theobald 1902, main identification characters

- 1. Four banded palpi
- 2. Fore leg tarsomeres with broad bands
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white
- 4. Lateral abdomen with tufts of golden scales

		Anopheles pulcherrimus
Name	:	An. pulcherrimus Theobald, 1902. Proceedings of the Royal Society of London, Series B, 69: 367-394.
Derivative	:	Latin, pulcher, beautiful; -imus, superlative suffix, most beautiful. A beautiful species, closely related to An. kochi, Doenitz, but quite easily distinguished by the white hind tarsomers, wing ornamentation, and absence of thoracic ocelli.
Type form	:	Available at the British Museum of Natural History, London.
Reported morphological variations	:	2 in maxillary palpi
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit		Prefers human dwellings to cattlesheds, also rests outdoors. Adults prefer to rest in partly shaded places in houses.
Breeding ecology		Ground pools, borrowpits with good growth of vegetation, ricefields, and shallow lakes.
Biting time	*	Peak activity before midnight from 18.00 to 21.00 hrs.
Feeding preference	:	Cattle to man.
Flight range	:	A hardy species and strong flier, e.g., the swarm of this species was observed over a ship about 25 km from mainland in Shat-al-Arab in the Persian Gulf.
Susceptibility to insecticides	:	NA
Relation to disease		In India, is not a vector but transmits malaria in northern Afghanistan and southern parts of USSR.
Reported distribution	:	Occurs in Bahrain. India, Iran, Iraq, Israel, Lebanon, Saudi Arabia, Syria, and Turkey. In India, recorded from Delhi, Gujarat, Haryana, Karnataka, Maharashtra, Orissa, Punjab, Rajasthan, Sikkim, Uttar Pradesh, and West Bengal.
Vector incrimination	•	NA

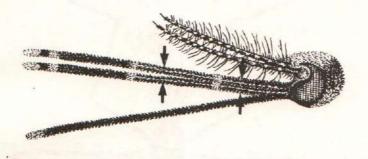


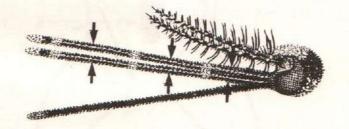
Reported distribution of An. pulcharrimus in the V/orld.

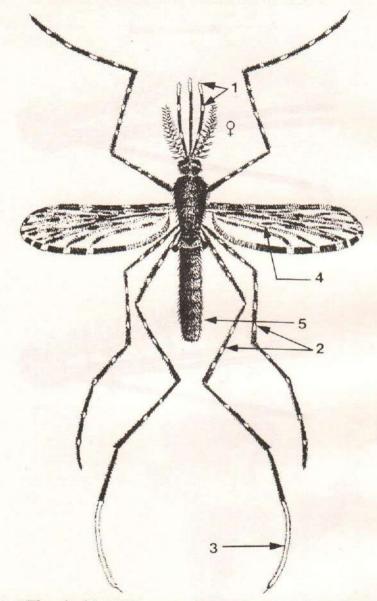


Reported distribution of An. pulcherrimus in India.

REPORTED IMPORTANT VARIATIONS OF An. pulcherrimus







An. jamesii Theobald 1901, main identification characters

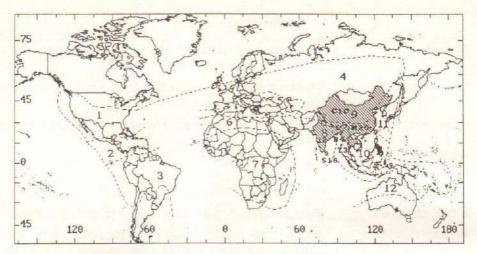
- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. Legs with speckling.
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white.
- 4. At the bifurcation of vein 5 (CU) the area is pale.
- 5. Abdomen with golden hairs.

		Anopheles jamesii
Name	:	An. jamesii Theobald, 1901. A Monograph of the Culicidae or Mosquitoes, 1: 134–135.
Derivative	:	Theobald named this species after Captain Sydney Price James, a wellknown malariologist.
Type form		Available at the British Museum of Natural History, London.
Reported morphological variations		2 in maxillary palpi
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	***	Adults rest inside houses, cattlesheds, and mixed dwellings during the day. The species also rests outdoors in forest areas.
Breeding ecology		Prefers to breed in tanks, fallow and growing ricefields with green vegetation, rainwater pools, ponds, riverbed pools, springs and surface wells along with green vegetation.
Biting time		After dusk.
Feeding preference	2	Cattle but sometimes man.
Flight range	:	NA
Susceptibility to insecticides		NA
Relation to disease	:	NA
Reported distribution		Distributed in Bangladesh, China, India, Indo-China,

Malaya, Myanmar, Nepal, Sri Lanka, and Thailand. In India, recorded from Andhra Pradesh, Arunachal Pradesh, Assam, Delhi, Goa, Gujarat, Haryana, Karnataka, Bihar, Kerala, Madhya Pradesh, Maharashtra, Meghalaya, Mizoram, Orissa, Punjab, Rajasthan, Tamil Nadu, Tripura,

Uttar Pradesh, and West Bengal.

Vector incrimination: NA

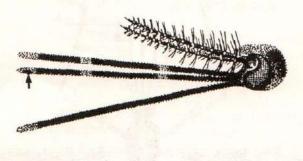


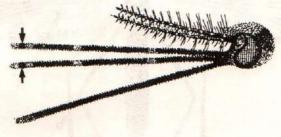
Reported distribution of An. jamesii in the World.



Reported distribution of An. jamesii in India.

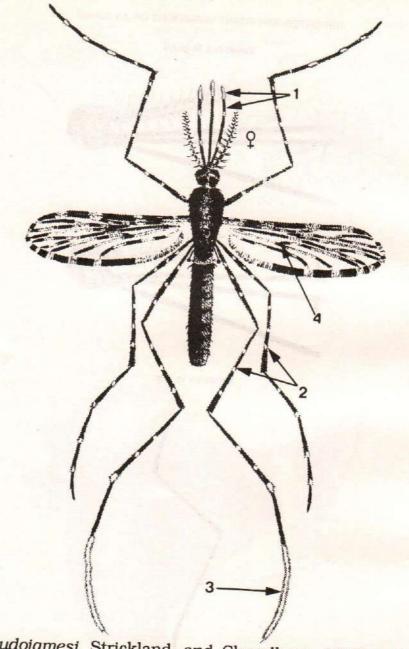
REPORTED IMPORTANT VARIATIONS OF An. jamesii





Variation in leg





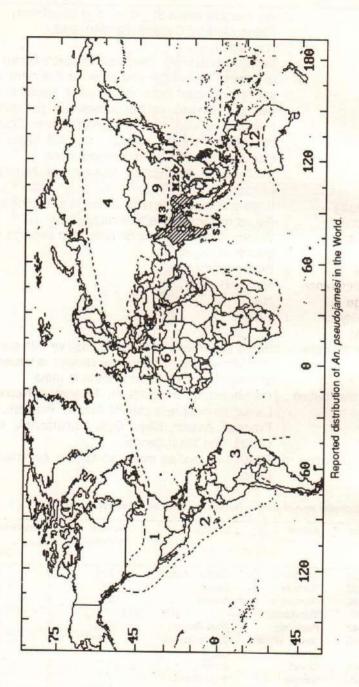
An. pseudojamesi Strickland and Chowdhury 1927, main identification charecters

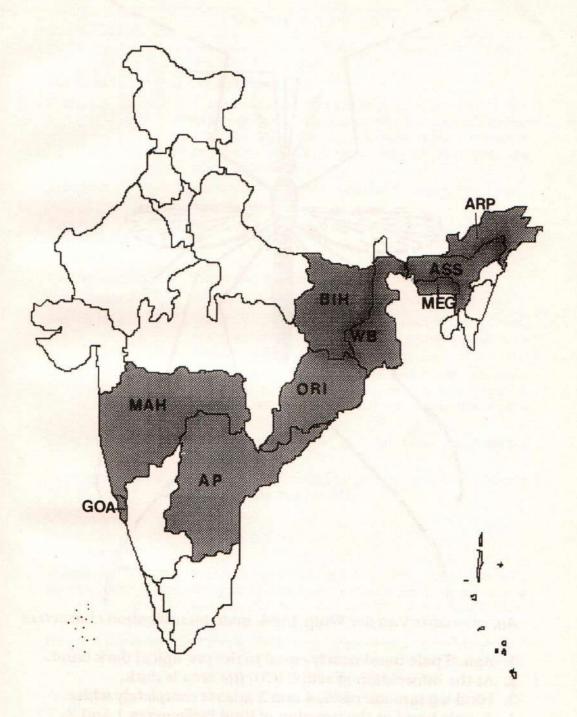
- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. Legs with speckling.
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white.
- 4. At the bifurcation of wing vein 5 (CU) the area is dark.

		Anopheles pseudojamesi
Name	:	An. pseudojamesi Strickland and Chowdhury, 1927. The Indian Medical Gazette. 62: 240-243.
Old Name	:	An. ramsayi
Derivative		Larvae which were identified as pulcherrimus gave rise to jamesi-like adults, and when a sufficient number of specimens had been collected it became clear that this species was neither jamesi nor pulcherrimus and therefore it was named pseudojamesi. Covell (1927) described it as An. ramsayi. In 1985 Nurul Huda and
		Harrison renamed it An. pseudojamesi.
Type form	:	Available at the British Museum of Natural History, London.
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Adults rest in houses and cattlesheds.
Breeding ecology	:	Breeds in rainwater pools, tanks and swamps with heavy growth of vegetation.
Biting time	:	
Feeding preference	:	Man and cattle.
Flight range	:	NA
Susceptibility to insecticides	;	NA
Relation to disease	:	Some specimens were found positive with gut and gland infection in Assam; hence the species is known as a poor or insignificant vector of malaria in India.
Reported distribution	:.	Distributed in Bangladesh, Myanmar, Nepal, and Sri Lanka; in India restricted to Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Goa, Maharashtra, Meghalaya, Orissa, and West Bengal.
Vector incrimination	:	Results of studies made so far are summarized in the table.

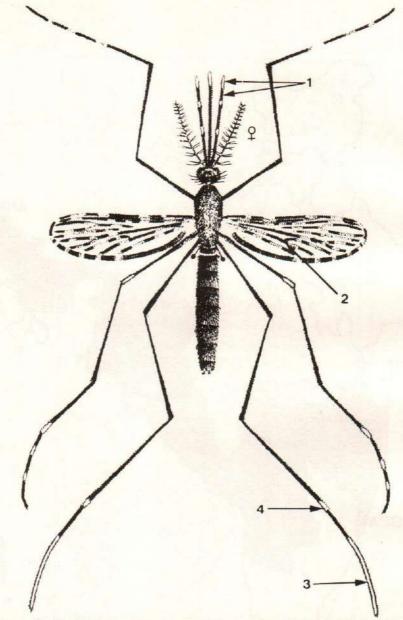
Table: Anopheles pseudojamesi: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive			
					Gut	Gland	Total	
1.	1929	Strickland	Cachar, Assam	256	0	1	1	
2.	1930	Ramsay	Assam	287	1	0	1	
3.	1936	Ramsay & Macdonald	Assam	2217	4	0	4	
4.	1942	Panigrahi	West Bengal	1658	1	0	1	
5.	1943	Senior White et al.	Orissa coastal	556	i	o	1	
6.	1944	Covell	Assam	2217	0	1		
7.	1940	lyengar	West Bengal	557	1	0	1	
8.	1944	lyengar	Deltic West Bengal	742	i	0	1	





Reported distribution of An. pseudojamesi in India.



An. annularis Van der Wulp 1884, main identification characters

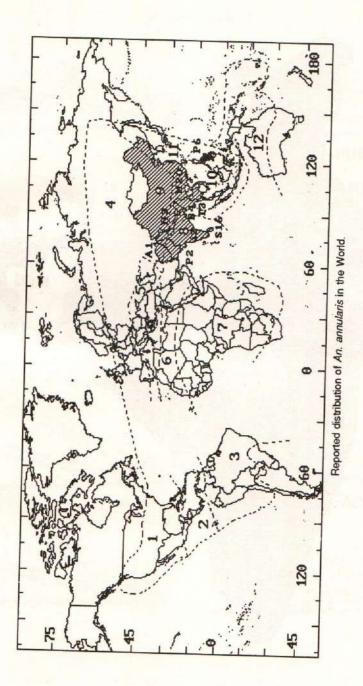
- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. At the bifurcation of vein 5 (CU) the area is dark.
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white.
- 4. A pale band at the junction of hind tarsomeres 1 and 2.

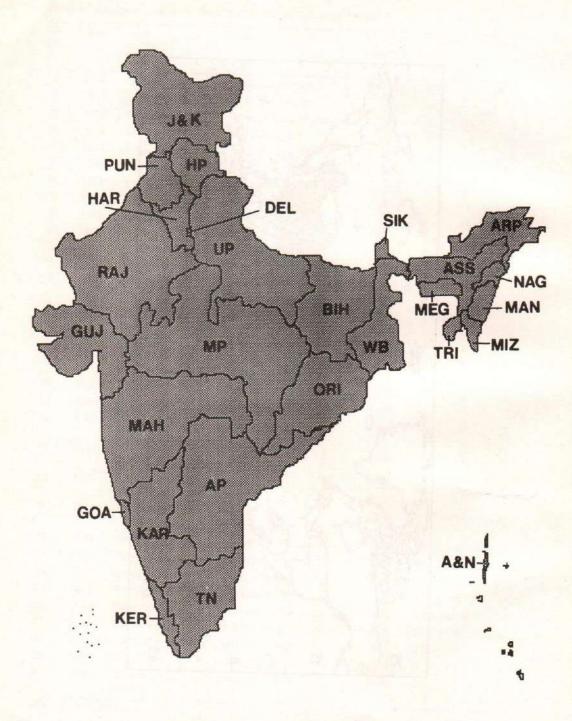
		Anopheles annularis
Name	:	An. annularis Van der Wulp, 1884. Notes from the Leyden Museum. 6: 248-256.
Old name	:	An. fuliginosus Giles
Derivative		Latin, annularis, of a ring; from annulus, a late Latin form of annulus, diminutive of anus, ring. Van der Wulp named it An. annularis because it is a ringed species, with white palpal rings as well as white rings on the anterior and posterior tarsi.
Type form	:	Available at the State Museum of Natural History, Liyden, Netherlands.
Reported morphological variations	:	10 in maxillary palpi, 6 in leg and 6 in wing
Species complex	:	Two sibling species.
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Adults rest in good numbers in houses, cattlesheds, and mixed dwellings during day. The species also rests outdoors in small numbers.
Breeding Ecology		Predominantly breeds in still water with abundant vegetation in tanks, ponds, borrowpits. Also breeds in ricefields, wells, margins of rivers, lakes, stream with green vegetation.
Biting time	:	Bites throughout night, but peak biting occurs before midnight.
Feeding preference	:	Predominantly a cattle biter (zoophilic) but in the absence of cattle bites man readily.
Flight range		Up to 1 km.
Susceptibility to insecticides		Resistant to DDT
Relation to disease	1	Secondary vector of malaria in China, India, Indonesia, Malaysia, Myanmar and Nepal. In India, known to be a vector of malaria in several localities of coastal Orissa, Bihar, and West Bengal.
Reported distribution	n	recorded from all zones including Kashmir.
Vector incrimination	1	: Results of studies made so far are summarized in the table.

Table: Anopheles annularis: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive			
_					Gut	Gland	Total	
1.	1905	Adie	Ferozepur, Punjab	142	0	1	1	
2.	1911	Christophers	Amritsar, Punjab	100	0	-	-	
3.	1913	Graham	Meerut, Uttar Pradesh	251	3	0	3	
4.	1914	Bentley	West Bengal	685	0	3	3	
5.	1914	Fry	West Bengal	1245	0	1	1	
6.	1924	Watts	Singbhum, Orissa	NM	_	_	_	
7.	1927	lyengar	West Bengal	156	0	2	2	
8.	1927	Covell	West Bengal	2511	3	4	7	
9.	1928	lyengar	Hooghly, West Bengal	125	0	1	1	
10.	1929	Sur & Sur	Krishna Nagar, West Bengal	3944	0	6	6	
11.	1929	Sur & Sur	Birnagar, West Bengal	787	0	1	1	
12.	1929	King & Krishnan	Udayagiri, Madras Tamil Nadu	8	1	0	1	
13.	1931	Macdonald & Majid	Karnal, Haryana	NM	0.5%	-	-	
14.	1935	Timbers	Birbhum, West Bengal	49698	0	8	8	
15.	1939	Senior White & Adhikari	Chilka lake area, Orissa	1048	3	0	3	
16.	1940	lyengar	West Bengal	5155	1	1	2	
17.	1940	lyengar	West Bengal	NM	_	-	0.49	
18.	1940	Senior White & Adhikari	West Bengal	1627	2	0	2	
19.	1941	Viswanathan et al.	Assam	7481	4	7	11	
20.	1941	Anderson & Viswanathan	Assam	16760	3	6	9	
21.	1941	Rao	Bengal Prov. West Bengal	884	5	2	7	
22.	1942	Covell & Singh	Chilka lake, Orissa	20844	1	0	1	
23.	1942	Panigrahi	Puri, Orissa	9096	5	7	12	
24.	1943	Senior White et al.	Orissa Plain	9183	14	7	21	
25.	1943	Senior White	Hazaribagh, Bihar	1030	1	0	1	
26.	1944	Rao	Dhanbad, Bihar	NM	0.5%		-	
27.	1946c	Senior White	E. Cen. India	215	0.5%	1	1	
28.	1982	Dash et al.	Keonjhar, Orissa	174	0	1	1	
29.	1985	Ghosh et al.	West Bengal	5428	0	1	1	
30.	1989	Gunasekaran et al.	Koraput, Orissa	3608	2	1	3	

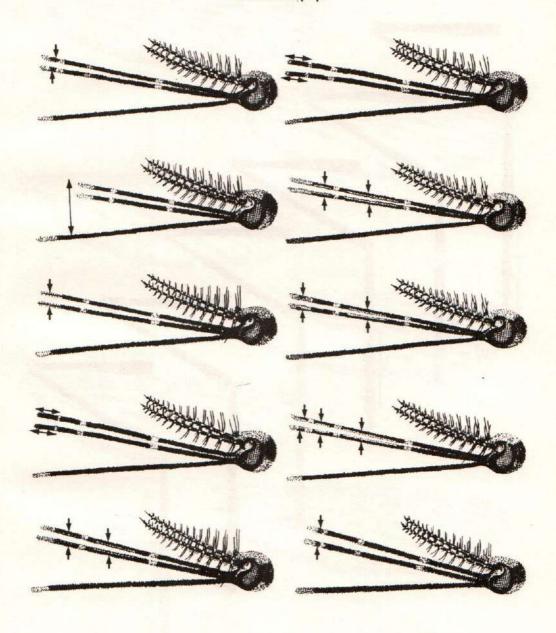
NM - Not Mentioned



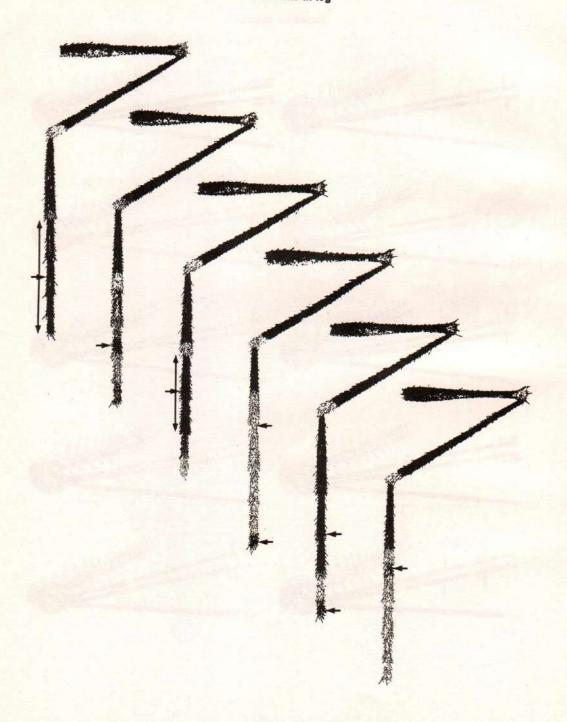


Reported distribution of An. annularis in India.

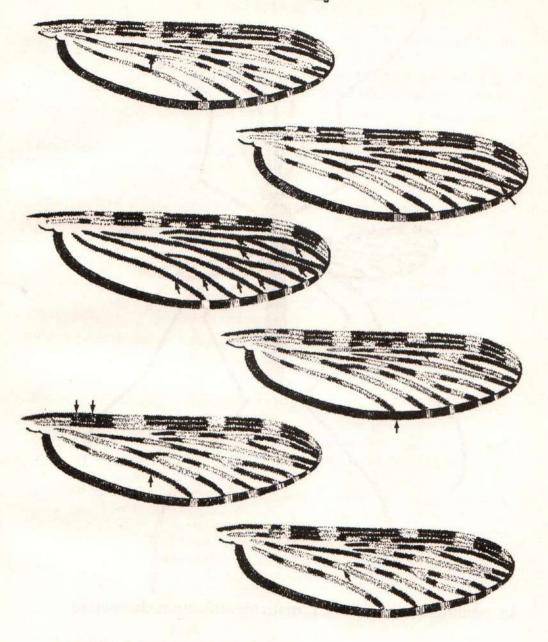
REPORTED IMPORTANT VARIATIONS OF An. annularis

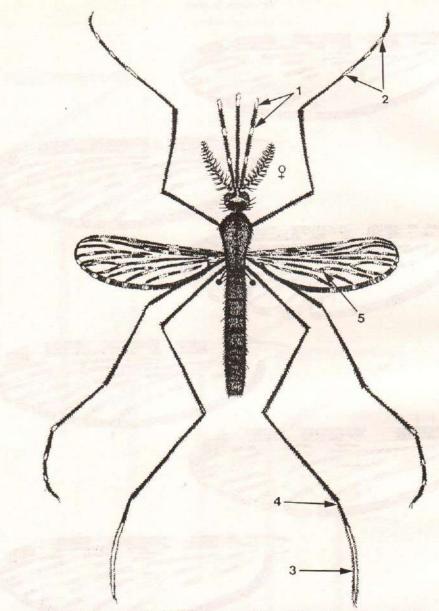


Variations in leg



Variations in wing





An. pallidus Theobald 1901, main identification characters

- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. Fore leg tarsomeres with broad bands.
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white
- 4. Pale band at the junction of hind tarsomeres 1 and 2 absent
- 5. At the bifurcation of vein 5 (CU) the area is pale

Ano	nhei	las	nal	lidus
MILL	PITCI	62	pai	iluus

Name : An. pallidus Theobald, 1901. A Monograph of the Culicidae or Mosquitoes, 1: 132-134

 Latin, pallidus, pale, ashen. Theobald described it as a variety but it is now accorded the full status of a species.

Type form : Available at the British Museum of Natural History,

London.

Reported morphological variations

Derivative

: 3 in maxillary palpi and 2 in leg.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests in houses and cattlesheds, and sometimes

outdoors.

Breeding ecology : Breeds in tanks, ponds, ditches with vegetation, shallow

pools in beds of streams, seepages, borrowpits, irrigation channels, wells, and fallow and growing ricefields.

Biting time : Before midnight.

Feeding preference : A zoophilic species but in the absence of cattle rapidly

bites man.

Flight range : NA

Susceptibility to insecticides

Susceptible to DDT.

Relation to disease : Not a vector of malaria of any significance. Some gut- and

gland-infected Plasmodium specimens were detected in

India.

Reported distribution: Occurs in Bangladesh, Myanmar, India, Indonesia,

Malaysia, Nepal, Sri Lanka, and Thailand. In India, recorded from Assam, Arunachal Pradesh, Andhra Pradesh, Bihar, Delhi, Gujarat, Goa, Haryana, Karnataka, Kerala, Manipur, Maharashtra, Madhya Pradesh, Orissa, Punjab, Rajasthan, Tamil Nadu, Tripura, Uttar Pradesh,

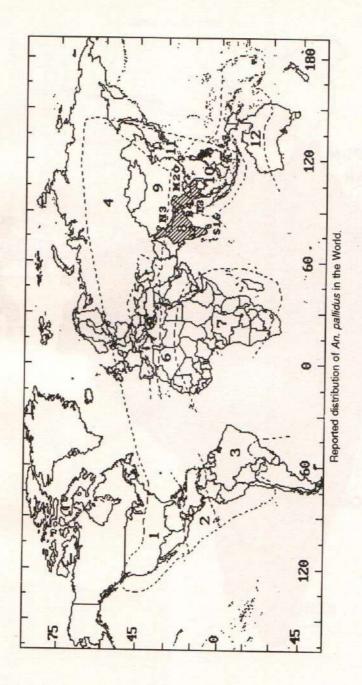
and West Bengal.

Vector incrimination : Results of studies made so far are summarized in the

table.

Table: Anopheles pallidus: Results of vector incrimination studies

SI. No.	iodi	Author	Author Locality		Number Positive			
1.	1929	Sur & Sur			Gut	Gland	Total	
2.			Krishnagar, West Bengal	1232	0	3	3	
	1931	Bose	West Bengal	1000				
3.	1935	Timbers	Birbhum, West Bengal	1236	3	0	3	
4.	1935	Timbers	Birbhum, West	35429	_	_	0.39	
-			Bengal Bengal	27238	0	9	9	
5.	1939	lyengar	Birbhum, West Bengal	254	2	0	2	
6.	1939	lyengar	West Bengal					
7.	1940	lyengar	West Bengal	508	0	20	20	
8.	1940	Senior White		408	2	0	2	
nan tier		& Adhikari	Satpura, Madhya Pradesh	3157	4	0	4	
9.	1942	loy & Biswas	Udaipur, Rajasthan				7	
10. 3	1942	Roy & Biswas	Orissa	854	0	6	6	
1.	1943	Senior White	Orissa	7513	60	0	60	
		et al.	Olissa	1409	1	0	1	
2.	1944	Covell	West Bengal			HOTEL S	Marie II	
3.	1944	Rao	Dhanbad, Bihar	403	1	0	1	
4.	1950	Viswanathan		NM	0.8%	0.1%	_	
	lot mentio		Panchamahal, Gujarat	187	1		la la	

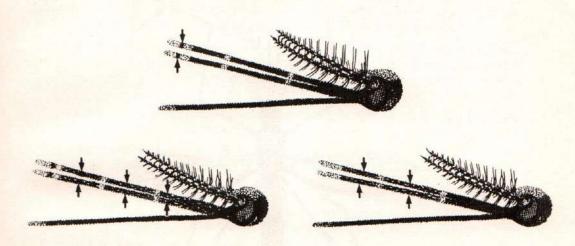




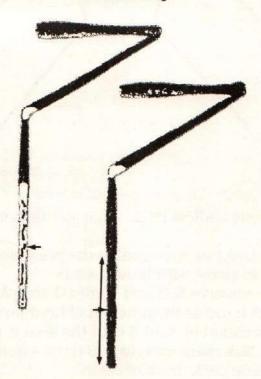
Reported distribution of An. pallidus in India.

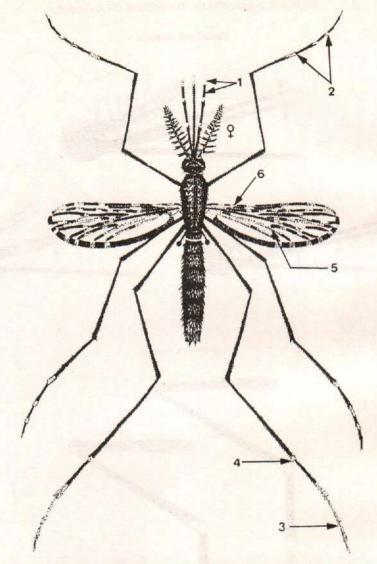
REPORTED IMPORTANT VARIATIONS OF An. pallidus

Variations in palpi



Variations in leg





An. philippinensis Ludlow 1902, main identification characters

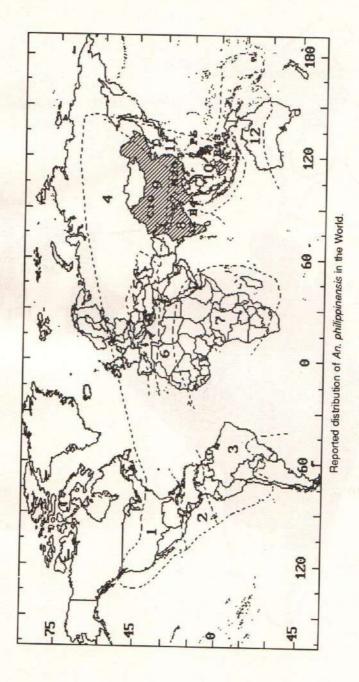
- 1. Apical pale band nearly equal to the pre-apical dark band
- 2. Fore leg tarsomeres with broad bands
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white.
- 4. A small pale band at the junction of hind tarsomeres 1 and 2
- 5. At the bifurcation of vein 5 (CU) the area is pale
- 6. Pre-sector dark mark on vein 1 (R1) not extending basally up to the humeral dark mark of costa

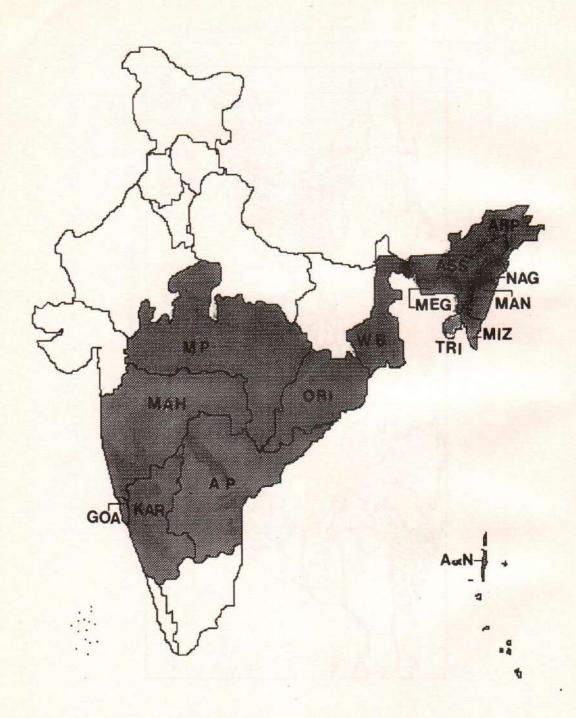
	Anopheles philippinensis*
Name	: An. philippinensis Ludlow, 1902. The Journal of the American Medical Association. 39: 426-427.
Derivative	: The species was collected during the course of Ludlow's Classification and Study of the Geographical Distribution of the Mosquitoes of the Philippine Islands and therefore the author named this species after the Philippine Islands.
Type form	: Available at the United States National Museum, Washington, D.C.
Reported morphological variations	: 1 in maxillary palpi and 1 in leg
Sitting posture	: Body and mouth parts at an analy
Resting habit	 Body and mouth parts at an angle to resting surface. Rests in human dwellings, cattlesheds and stables. Also rests outdoors on vegetation in forests.
Breeding ecology	Breeds in a variety of places particularly swamps, tanks, ponds, ditches, ricefields etc., generally with a good growth of vegetation in stagnant water. Also breeds in shaded parts of lakes, inundated nullahs and pools, borrowpits with vegetation, tree holes, leaf axils of plants and grassy edges of very slow running streams.
Biting time	Bites outdoors and indoors throughout night, with peaks between 20.00 and 22.00 hrs and 2.00 and 4.00 hrs.
Feeding preference	: Predominantly zoophilic, also bites man.
Flight range	: 0.4-0.8 km.
Susceptibility to insecticides	: Susceptible to DDT and HCH.
Relation to disease	: A secondary vector of malaria in Bangladesh, India and Myanmar.
Reported distribution	: Found in Bangladesh, Borneo, China, France, Hainan-Island, India, Indochina, Java, Malaya, Myanmar, the Philippines, and Sumatra. In India, recorded from Andaman Islands, Andhra Pradesh, Arunachal Pradesh, Assam, Goa, Karnataka, Madhya Pradesh, Maharashtra, Manipur, Meghalaya, Mizoram, Nagaland, Orissa, Tripura, and West Bengal
ector incrimination	Results of studies made so far are summarized in the table.

^{*}The presence of this species yet to be confirmed in India. Studies so far have revealed that true An. philippinensis is not found in India and all specimens examined from Eastern India were An. nivipes.

Table: Anopheles philippinensis: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Nu	ımber Posi	tive
					Gut	Gland	Tota
1.	1929	Sur	West Bengal	223	3	5	8
2.	1929	Sur & Sur	Krishnagar, West Bengal	770	0	15	15
3.	1929	Sur & Sur	Birnagar, West Bengal	458	0	7	7
4.	1932	Bose	Birnagar, West Bengal	499	_	_	11
5.	1935	Timbers	Birbhum, West Bengal	12594			131
6.	1937	Niogi & Khan	West Bengal	101	0	2	2
7.	1939	lyengar	West Bengal	1918	81	76	157
8.	1940	lyengar	West Bengal (1 locality)	1830	68	65	133
9.	1940	lyengar	West Bengal (10 localities)	3758	149	141	290
10.	1940	Krishnan	Burdwan, West Bengal	1053	_	-	89
11.	1941	Viswanathan et al.	Assam	4239	2	2	4
12.	1940	lyengar	West Bengal	3165	149	126	275
13.	1944	lyengar	Deltaic West Bengal	865	-		64
14.	1947	Ganguli	West Bengal	297	5	7	12
15.	1948a	Sen	West Bengal	1053	_	_	89
16.	1948a	Sen	Madhagram, Calcutta, West Bengal	180	-	_	12
17.	1976	Rajagopal	Burnihat, Meghalaya	174	0	- 1	1

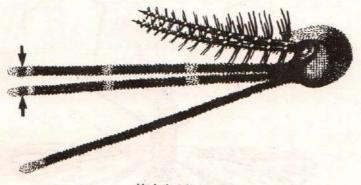




Reported distribution of An. philippinensis in India.

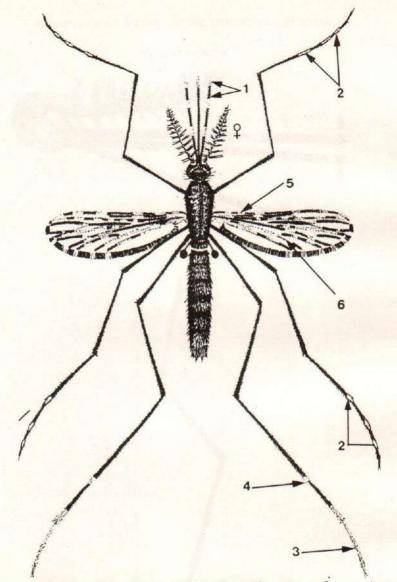
REPORTED IMPORTANT VARIATIONS OF An. philippinensis

Variation in palpi



Variation in leg





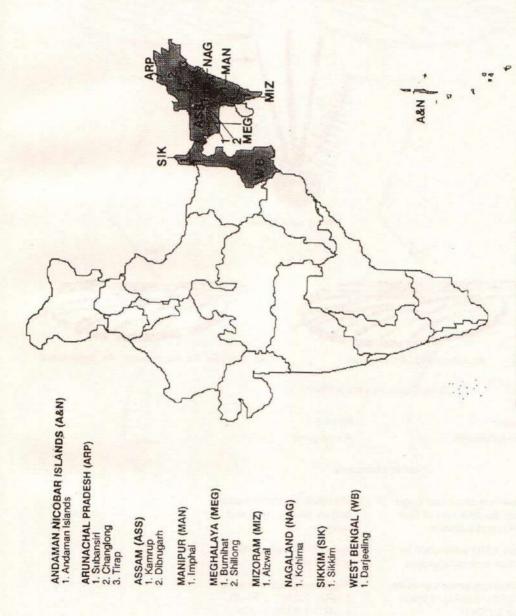
An. nivipes (Theobald) 1903, main identification characters

- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. Fore leg tarsomeres with broad pale bands.
- 3. Hind leg tarsomeres 5, 4 and 3 atleast completely white.
- 4. A small pale band at the junction of hind tarsomeres 1 and 2.
- Pre-sector dark mark on vein 1 (R1) basally extended up to humeral dark mark of costa.
- 6. At the bifurcation of vein 5 (CU) the area is pale.

		Anopheles nivipes
Name	:	An. nivipes (Theobald), 1903. The Entomologist. 36: 256-259.
Derivative	:	Latin, <i>niveus</i> , of snow, snowy; <i>pes</i> , foot. Theobald actually uses the term "snowy white" in reference to the scales on the thorax, but it is clear that the name refers to the hind-leg tarsi.
Type form	:	Available at the British Museum of Natural History, London.
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Rests indoors in cattlesheds and human dwellings and outdoors on vegetation in forests.
Breeding ecology	:	Breeds in a variety of places particularly swamps, tanks, ponds, ditches, ricefields, etc., generally with a good growth of vegetation in stagnant waters. Also breeds in shaded parts of lakes, inundated nullahs and pools, borrowpits with vegetation, tree holes, leaf axils of plants and grassy edges of very slowrunning streams.
Biting time	;	Bites outdoors as well as indoors throughout night, with peaks between 20.00 and 22.00 hrs and 2.00 and 4.00 hrs.
Feeding preference		Predominantly zoophilic, also bites man.
Flight range		0.4-0.8 km.
Susceptibility to insecticides	:	NA
Relation to disease	:	Known as a secondary vector of malaria in Malaya.
Reported distribution	n :	Occurs in India and Malaya. Recent studies show that the species which is prevalent in northeastern states of India is An. nivipes and not An. philippinensis.
Vector incrimination	:	NA



Reported distribution of An. nivipes in the World.

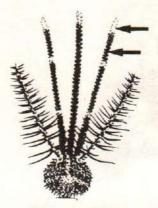


Reported Distribution of An. nivipes in India.

1.3 An. culicifacies, An. fluviatilis, An. jeyporiensis, An. sergentii, An. moghulensis

Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1)

Palpi with three pale bands, apical pale band nearly equal to the pre-apical dark band



Wing vein 3(R4+5)

Dark



An. culicitacies, An. sergentii

Pale



An. fluviatilis, An. moghulensis, An. jeyporiensis

Fringe spot on vein 3(R4+5)

Absent An. culicifacies Present

An. sergentii

Other characters

Main dark costal spot longer than the dark spot of vein 1(R1) and subcosta

Vein 1(R1) interrupted by a dark spot in basal area

Fringe spot present on veins 2.1(R2), 2.2(R3), 4.2(M2) and 5.1(Cu1)

Main dark costal spot equally long as on vein 1(R1) and subcosta

Vein 1(R1) completely pale at basal area

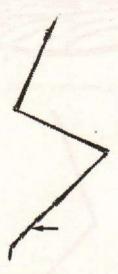
Fringe spot present on veins 2.1(R2), 2.2(R3), 3(R4+5), 4.1(M1), 4.2(M2), 5.1(Cu1) and 5.2(Cu2)

An. fluviatilis, An. jeyporiensis, An. moghulensis

Banding on foreleg tarsomeres

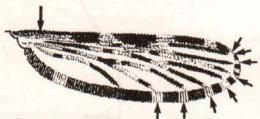
Absent

Present (small bands)

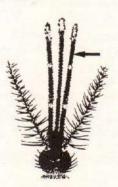


Other characters

Inner costa completely dark
Fringe spot present on all the veins except on
vein 6(Anal)



Palpi thin as compared to An. jeyporiensis and An. moghulensis



An. fluviatilis



An. jeyporiensis, An. moghulensis

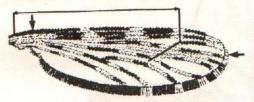
An. jeyporiensis, An. moghulensis

The distance of the anterior forked cell from the base of the costa compared to that of the posterior forked cell from the base of the costa.

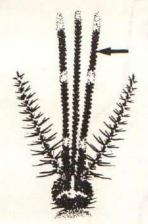
More An. moghulensis Less An. jeyporiensis

Other characters

Inner costa completely dark Apex of wing usually extremely pale Fringe spot absent on wing vein 6(Anal)



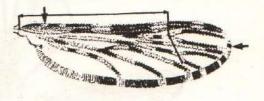
Palpi thick and long as compared to An. fluviatilis



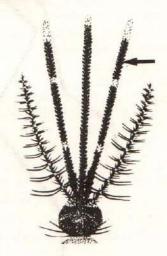
Mesonotum with broad scales and with a line of broad overlapping scales at the sides in front of wing root

An. moghulensis

Inner costa interrupted
Apex of wing not extremely pale
Fringe spot present on wing vein 6(Anal)

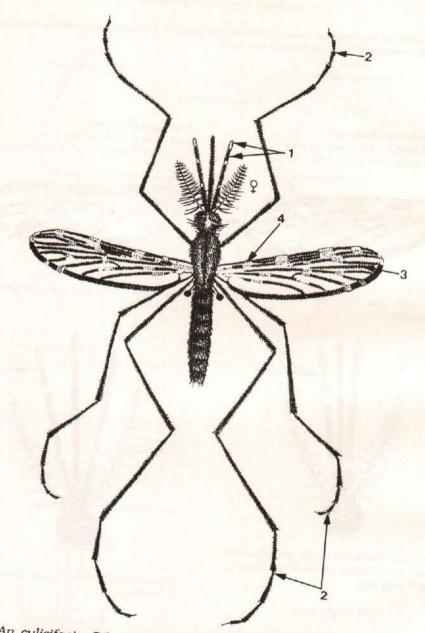


Palpi thick and long as compared to An. fluviatilis



Mesonotum with narrow scales forming a tuft in median area

An. jeyporiensis



An. culicifacies Giles 1901, main identification characters

- Apical pale band nearly equal to the pre-apical dark band.
 Tarsomeres without bands
- 3. Vein 3 (R4+5) mainly dark
- 4. Inner costa interrupted

		Anopheles culicifacies
Name	:	An. culicifacies Giles, 1901. Entomologists' Monthly Magazine, 37: 196–198.
Derivative	•	Latin, culex (genitive, culicis), a gnat, midge; facies, external form, look, appearance, aspect. It mimics Culex fatigans (-Culex quinquefasciatus) when resting, especially the females rest/sit 'humped up' in exactly the same way as those of the genus Culex and therefore named An. culicifacies.
Type form	:	Available at the British Museum of Natural History, London.
Reported morphological variations	:	8 in maxillary palpi and 11 in wing
Species complex	:	Four sibling species (known as A, B, C and D).
Sitting posture	:	Body parallel to resting surface.
Resting habit		Prefers to rest in cattlesheds and houses during the day. The species was also collected from straw, mudcakes, etc. near stables and from dense vegetation, under the bushes and in tree holes, an indication of outdoor resting.
Breeding ecology	•	Prefers to breed in river margins, river-bed pools, canal systems, seepage waters from canals and dams, rainwater collections in borrowpits, or in any lowlying grounds and shoreline pockets in dams and lakes. Also breeds in hoof marks and wheel ruts, ricefields, wells, ponds, pools of water supply systems, and brackishwater pools.
Biting time	:	Throughout night but peak biting occurs between 19.00 and 4.00 hours with variations from place to place.
Feeding preference	:	The species is predominantly zoophilic, a wide range of variation found in anthropophily.
Flight range	:	1–3 km.
Susceptibility to insecticides	:	Resistant to DDT, and HCH in many parts of the country. Also resistant to malathion in Gujarat and Maharashtra and a few other areas.
Relation to disease	:	Efficient vector of human malaria in India and neighbouring countries, e.g., Afghanistan, Iran, Muscat, Myanmar, Nepal, Pakistan, and Sri Lanka. Recently W. bancrofti larvae were detected in this species from Shahjahanpur (U.P.), India.
Reported distribution	:	Occurs very widely, i.e. from Afghanistan, Bahrain, Bangladesh, India, Indochina, Iran, Muscat, Myanmar, Nepal, Pakistan, Southern China, Sri Lanka, Trucial Oman and Thailand. In India, recorded throughout the country except Andaman & Nicobar Islands.
Vector incrimination		Results of studies made so far are summarized in the

table.

Table: Anopheles culicifacies: Results of vector incrimination studies

2. 3. 4. 5. 6. 7. 8. 9. 10. 11. 11. 11. 11. 11. 11. 11. 11. 11	Year	Author	Locality	Number dissected	N	umber Pos	sitive
					Gut	Gland	Tota
	1902	Stephens & Christophers	Ennore, Tamil Nadu	69	0	6	6
	1902	Stephens & Christophers	Punjab	252	0	11	11
	1902	Cornwall	Ennore, Tamil Nadu	25	0	4	4
	1910	Graham	Kosi, Uttar Pradesh	134	2	2	4
	1910	Graham	Kairana, Haryana	128	1	ō	1
	1912	Ross	Ennore, Tamil Nadu	85	0	1	1
	1913	Ross	Cuddapah, Tamil Nadu	200	2	ò	2
	1913	Graham	Meerut, Uttar Pradesh	211	1	0	1
	1913	James & Gunasekara	Talimannar	36	0	1	1
C-CATACO	1914	Kenrick	Cent. Prov., (Madhya Pradesh)	675	0	2	2
	1914	Horne	Penukonda	16	0	1	4
	1914a	Hodgson	Delhi, Union Territory	111	3	o	3
13.	1915	Mhaskar	N. Kanara, Karnataka	837	6	2	
14.	1915	Rao	Hospet	116	0	2	8
15.	1915	Rao	Ramnad, Tamil Nadu	44	2	0	2
16.	1925	Krishnan	Vizagapatnam, Andhra Pradesh	31	1	0	2
17.	1925	Sinton	Mani Majra	6	1		
18.	1925	Ayer	Sappal Hill	98	0	0	1
19.	1927	lyer	Bimlipatam	37	4	1	1
20.	1927	Covell	Ennore, Tamil Nadu	81	0	0	4
21.	1927	Covell	Hospet	128	U	1	1
22.	1927	Covell	Kanara, Karnataka	837	_	_	2
23.	1927	Covell	Ramnad, Tamil Nadu		6	. 2	8
24.	1927	Covell	Penukonda, Andhra	16	0	1	2
25	1007	0 "	Pradesh		•	1	
	1927	Covell	Vizagapatam, Andhra Pradesh	31	1	0	1
	1928	Mayne	Saharanpur, Uttar Pradesh	2021	2	3	5
	1928	Gill	Punjab	57	0	2	2
	1929	King & Krishnan	Udayagiri, Tamil Nadu	42	4	1	5
	1929	Venkataraman	Vizagapatnam, Andhra Pradesh	186	3	o	3
30.	1929	Rao	Nandyal, Andhra Pradesh	68	1	1	2
11.	1929	King & Iyer	Mopad, Madras, Tamil Nadu	297	18	4	22
2.	1929	lyer	Udayagiri, Tamil Nadu	52			
3.	1931	Sweet & Rao	Kolar, Karnataka		1	1	2
4.	1931	Macdonald & Majid	Karnal, Punjab	NM	9.8%	2	2
5.	1931	Sweet & Rao	Mysore, Karnataka	E740			
6.	1934	Nursing et al.	Nagenhalli	5748	1	7	8
		Jan San Car.	ragernan	271	4	0	4

(Contd.)

Table: Anopheles culicifacies: Results of vector incrimination studies (Contd.)

SI. No.	Year	Author	Locality	Number dissected	Nu	mber Pos	sitive
				XIII XIII XIII XIII XIII XIII XIII XII	Gut	Gland	Tota
37.	1934	Nursing et al.	Mysore, Karnataka	1964	46	14	60
38.	1934	Nursing et al.	Hariyur, Karnataka	447	4	2	6
39.	1934	Nursing et al.	Mandya, Karnataka	1151	26	12	38
40.	1934	lyengar	Travancore, Kerala	984	0	1	1
41.	1934	Ahuja	Ajmer, Rajasthan	53	4	2	6
42.	1936	Viswanathan	Madakasira	70	2	2	4
43.	1936	Lamprell	Lumding, Assam	516	0	4	4
44.	1937c	Senior White	Chatikona, Orissa	1168	2	0	2
45.	1937c	Senior White	Jeypore hills, Orissa	4744	3	0	3
46.	1937	Hicks & Majid	Karnal, Haryana	8850	20	32	52
47.	1937	Sweet	T. Narsipur (Mysore, Karnataka)	1191	45	8	53
48.	1937	Niogi & Khan	Sylee (Dooar),	124	0	1	1
			North Bengal				
49.	1937c	Senior White	Lanjigarh Road	1067	1	0	1
			(Jeypore hills) Orissa				
50.	1937	Ramsay	Dharbanga, Bihar	2988	2	2	4
51.	1938	Covell et al.	Delhi	2507	1	2	3
52.	1938	Senior White	Jeypore hills, Orissa	2446	2	0	2
53.	1938	Russell et al.	Madras, Tamil Nadu	6484	4	5	9
54.	1938	Barber & Rice	Poona, Maharashtra	1016	0	3	3
55.	1938	Senior White & Das	Singhbhum hills, Bihar	1611	5	ō	5
56.	1938	Ramsay	Peermade	401	14	8	22
57.	1939	Russell & Jacob	Ennore, Madras Tamil Nadu	984	7	1	8
58.	1939	Sundaresan	Baruva, Orissa	122	1	0	1
59.	1939	Mathew	Travancore, Kerala	1131	6	3	9
60.	1940	Russell & Rao	Madras, Tamil Nadu	13156	8	8	16
61.	1940	Senior White	Chota Nagpur, Bihar	7482	15	0	15
62.	1940	Senior White	Kesinga, Orissa	129	1	0	1
63.	1940	Senior White	Khodri, Satpura, Madhya Pradesh	700	1	0	1
64.	1940	Senior White	Khongsara, Satpura, Madhya Pradesh	1317	2	0	2
65.	1940	Senior White	Balghara	541	1	0	1
66.	1940	Senior White	Bhadrak, Orissa	119	1	0	1
67.	1940	Senior White	Bhanwar Tank	426	1	1	2
68.	1940	Senior White	Paniyajobi	425	1	o	1
69.	1940	Senior White &	Singhbhum hills,	236	1	o	1
		Narayana	Bihar	200		•	-
70.	1940	Senior White & Adhikari	Satpura ranges, Madhya Pradesh	909	0	2	2
71.	1941	Anderson & Viswanathan	Lumding, Assam	1232	2	5	. 7
72.	1942	Roy & Biswas	Dharamjoi Garh	969	0	5	5
73.	1943	Singh & Jacob	Ahmedabad, Gujarat	234	0	1	1
74.	1943	Covell &	Delhi, Union	9628	19	19	38
		Jaswant Singh	Territory				

(Contd.)

Table: Anopheles pallidus: Results of vector incrimination studies (Contd.)

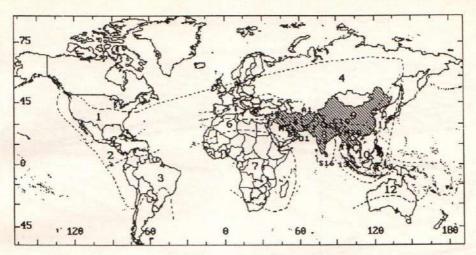
SI.		Author	Locality	Number dissected	110	mber Pos	sitive
95		- July		-10000100	Gut	Gland	Tota
75		COLUCT THINKS	Hazaribagh, Bihar	9571			
76	. 1943	Senior White	Orissa coastal		16	2	18
_	1000000	et al.	plain	735	1	0	-
77	1943		Thane, Maharashtra	171			
		Organization	Thanks don't a	174	-	_	1
78.	1943	Senior White &	Vizagapatnam,	0000			
Burn		Venkata Rao	Andhra Pradesh	6608	3	1	4
79.	1943	Senior White &	Vizagapantam,	470			
		Rao	Andhra Pradesh	179	1	1	2
80.	77,000 (100,000)	Rao	Dhanbad, Bihar	4500			
81.		Soman	Bombay, Maharashtra	1509	9	2	11
82.	1945	Senior White	Delhi, Union	629	1	0	1
		et al.	Territory	906	2	0	2
83.	1945	Pal	Delhi, Union				
			Territory	665	0	10	10
84.	1945	Subbarao &	Vizagapatnam (Urban),	14-14			
		Appa Rao	Andhra Pradesh	844	1	2	3
85.	1945	Subbarao &	Vizagapatnam (Rural),				
		Appa Rao	Andhra Pradesh	1377	1	1	2
86.	1946	Senior White	West Bengal	7			
		&Ghosh	vvest bengal	3041	1	0	1
87.	1946	De Burca	Central Provinces,				
		Name and Associated to the Party of the Part	(Modbus Deades)	424	6 .	6	12
88.	1946c	Senior White	(Madhya Pradesh) Dongargarh				
				4816	1	1	2
89.	1946	De Burca	(Satpura)				
			Jabalpur, Madhya Pradesh	166	4	4	8
90.	1946	De Burca					
91.	1946	Rao et al.	Mhow, Madhya Pradesh	258	1	1	2
92.	1948	Godbole et al.	Madras, Tamil Nadu	8357	9	4	13
		associa at al.	Bombay, Maharashtra	3453	_		9
93.	1948	Subramanian &	Cen. Prov.,				
	1000	Dixit	Cen. Prov.,	7337	0	16	16
94.	1949	Raghavan &	(Madhya Pradesh)				
	(Action)	Krishnan	Madras, Tamil Nadu	54	1	0	1
95.	1949	Bhatt	N3- Di				
		Dilati	Nasik Distt.,	3675	0	4	4
96.	1950	Viswanathan	Maharashtra				7
97.	1950	Viswanathan	Ahmedabad, Gujarat	737	4	2	6
98.	1950	Subramanian &	Bijapur, Karnataka	3463	10	12	22
1.5		Gupta	Dharamjaigarh,	2277	0	2	2
99.	1950	Jacob	Madhya Pradesh		Nijo.		-
00.	1950	Viswanathan	Jammu & Kashmir	1021	3	3	6
01.	1950	Viswanathan	Nasik, Maharashtra	10057	1	1	2
02.	1950	Viswanathan	Poona, Maharashtra	400	1	1	2
03.	1950	Viswanathan	Dharwar, Maharashtra	119	_		4
04.	1950	Viswanathan	Kaira, Maharashtra	1343	1	2	
٠.	1950	Viswanathan	East Khandesh,	2786	_	-	3 7
05.	1052	land, .	Maharashtra		DATE !		1
	1953	Issaris et al.	Terai, Uttar	5742	0	5	E
06.	1057		Pradesh			3	5
٠٠.	1957		Udaipur,	255	0	1	
			Rajasthan				1

Table: Anopheles culicifacies: Results of vector incrimination studies (Contd.)

119. 120. 121. 122. 123.	Year	Author	Locality	Number dissected	Number Positive			
				Salasasinasi	Gut	Gland	Total	
	1958a	Bhatia et al.	Khurja, Bulandshahar Uttar Pradesh	634	0	1	1	
	1974	Vaid et al.	Ratlam, Madhya Pradesh	209	0	1	1	
109.	1977	Pattanayak	Delhi, Union	1058	0	2	2	
		et al.	Territory					
110.	1978	Sitaraman	Rameswaram	91	0	1	1	
	4070	et al.	Island, Tamil Nadu					
111.	1979	Rahman et al.	Alwar,	798	0	3	3	
110	4000		Rajasthan					
112.	1980	Annual Report,	Jagdalpur	19571	1	3	4	
112	1980	NICD	0111 0 111	ATTEMPT OF				
113.	1900	Subba Rao et al.	Okhla, Delhi	17	0	1	1	
114	1980	Cubbs Des et et	Union Territory					
Control of the same	1982	Subba Rao et al. Annual	Mandora, Haryana	120	0	1	1	
110.	1302	Report, MRC	Kharkhoda PHC,	1254	0	1	1	
116	1982	Ansari et al.	Haryana Basastaur Harrana		200			
	1982a	Choudhury &	Basantpur, Haryana Kharkhoda PHC,	38	1	0	1	
	10024	Ghosh	Haryana	1862	0	19	19	
118.	1983	Choudhury	Delhi, Union	812	0	0		
		on out on one	Territory	012	U	8	8	
119.	1983	Choudhury	Ghaziabad,	814	0	10	10	
			Uttar Pradesh	014	U	10	10	
120.	1983	Choudhury	Kharkhoda PHC,	8447	0	83	83	
			Haryana	0447	0	03	03	
121.	1983	Choudhury et al.	Gadarpur,	1598	0	21	21	
			Uttar Pradesh	1550	0	21	21	
122.	1983-84	Annual Report,	Arthala, Uttar	333	0	8	8	
		MRC	Pradesh	-	-			
123.	1983-84	Annual Report,	Basantpur,	841	0	- 1	1	
		MRC	Haryana					
124.	1983-84	Annual Report,	Delhi, Union	633	0	3	3	
		MRC	Territory			11 (73)		
126.	1983-84	Annual Report,	Bhanera, Ghaziabad	1021	0	5	5	
250	Name of the last	MRC	Uttar Pradesh					
127.	1983-84	Annual Report,	Ghaziabad,	760	0	14	14	
		MRC	Uttar Pradesh					
128.	1983-84	Annual Report,	Raipur,	227	0	1	1	
100	1000 01	MRC	Uttar Pradesh					
129.	1983-84	Annual Report,	Khanpur,	134	0	1	1	
120	1000 04	MRC	Uttar Pradesh					
130.	1983-84	Annual Report,	Maseet	54	0	1	1	
131.	1000	MRC	Uttar Pradesh					
131.	1983	Choudhury	Ghaziabad,	1089	0	14	14	
132.	1983	Choudhury	Uttar Pradesh	4500		FIRE		
UL.	1903	Choudhury	Delhi,	1528	0	3	3	
33.	1983-1984	Annual	Union Territory Gadarpur,	4700	•			
	.000-1004	Report, MRC	Uttar Pradesh	1730	0	3	3	
34.	1984	Ansari et al.	Bareilly,	040	0			
	1004	rinoan et al.	Uttar Pradesh	243	0	2	2	
			Uttar Pradesh					

Table: Anopheies pallidus: Results of vector incrimination studies (Contd.)

SI. No.	Year	Author	Locality	Number	Nu	mber Pos	sitive
201					Gut	Gland	Total
135.	1984	Mani et al.	Tamil Nadu	3267	7	13	20
136.	1984	Sabesan et al.	Rameswaram Island, Tamil Nadu	4371	3	21	24
137.	1984-85	Annual Report, MRC	Bulandshahar, Uttar Pradesh	386	0	2	2
138.	1984-85	Annual Report, MRC	Bhanera Loni Ghaziabad, Uttar Pradesh	4147	0	9	9
139.	1984-85	Annual Report, MRC	Ghaziabad, Uttar Pradesh	3356	0	14	14
140.	1984-85	Annual Report, MRC	Mandora, Haryana	9124	0	83	83
141.	1984-85	Annual Report, MRC	Kichha, Uttar Pradesh	1050	0	2	2
142.	1985	Sharma et al.	Shahjahanpur, Uttar Pradesh	232	0	1	1
143.	1985-86	Annual Report, MRC	Delhi, Union Territory	163	0	1	1
144.	1985-86	Annual Report, MRC	Ghaziabad, Uttar Pradesh	3112	0	9	9
145.	1985-86	Annual Report, MRC	Bulandshahar, Uttar Pradesh	2483	0	16	16
146.	1985-86	Annual Report, MRC	Gadarpur, Uttar Pradesh	400	0	2	2
147.	1986	Nagpal & Sharma	Ganjam, Orissa	100	2	0	2
148.	1986	Nagpal & Sharma	Balangir, Orissa	72	0	1	1
149.	1986	Nagpal & Sharma	Cuttack, Orissa	309	1	2	3
150.	1986	Nagpal & Sharma	Phulbani, Orissa	69	0	1	1
151.	1986	Nagpal & Sharma	Sambalpur, Orissa	127	0	1	1
152.	1986	Ansari et al.	Jangethi, Meerut, Uttar Pradesh	816	0	4	4
153.	1987	Subba Rao et al.	Arthala, Uttar Pradesh	460	0	8	8
154.	1987	Subba Rao et al.	Basantpur, Haryana	243	0	1	1
155.	1988	Annual Report, MRC	Dadri, Ghaziabad, Uttar Pradesh	95	0	i	1
156.	1988	Annual Report, MRC	Bulandshahar/ Rajaranipur, Uttar Pradesh	165	0	1	1
157.	1988	Annual Report, MRC	Bulandshahar Gopalpur, Uttar Pradesh	132	0	1	1
158.	1988	Annual Report MRC	Bulandshahar Dhanpur, Uttar Pradesh	128	0	2	2
159.	1989	Gunasekaran et al.	Koraput, Orissa	8995	5	0	5
160.	1990	S & T Project Report	Shahjahanpur, Uttar Pradesh	4958	0	6	6
161.	1990	Prasad & Sharma	Banda, Shahjahanpur, Uttar Pradesh	69	0	2	2
162.	1990	Kulkarni	Bastar, Madya Pradesh	36278	1	3	4
163.	1991	Neeru Singh & Gyanchand	Mandla, Madhya Pradesh	220	0	1	1

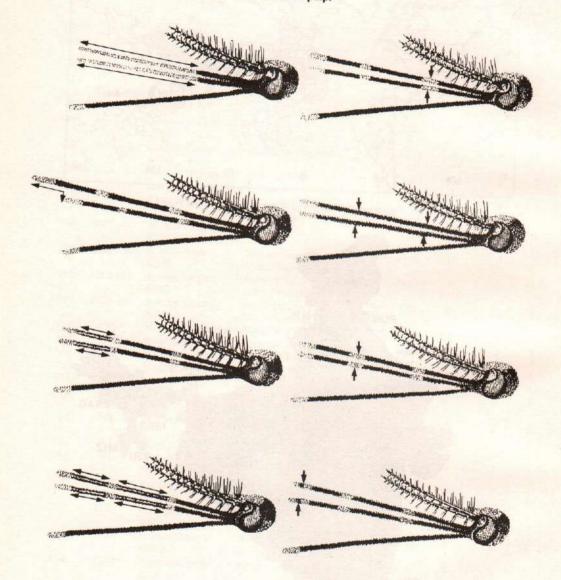


Reported distribution of An. culicifacies in the World.

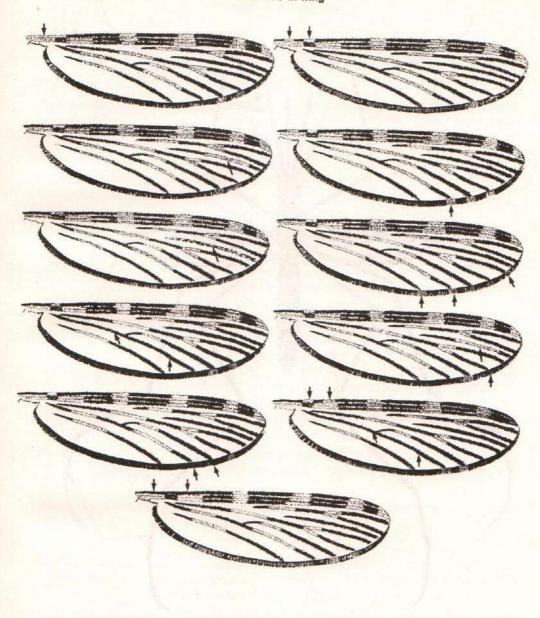


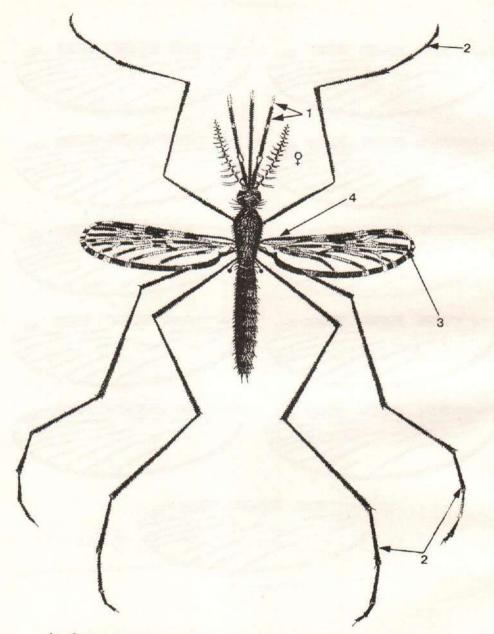
REPORTED IMPORTANT VARIATIONS OF An. culicifacies

Variations in palpi



Variations in wing





An. fluviatilis James 1902, Main identification characters

- Apical pale band nearly equal to the pre-apical dark band.
 Tarsomeres without bands.

- Vein 3 (R4+5) mainly pale.
 Inner costa completely dark.

		Anopheles fluviatilis
Name	:	An. fluviatilis James, 1902. Scientific Memoirs by Officers of the Medical and Sanitary Departments of the Government of India (New Series), 2: 106.
Old name	:	An. listoni Liston.
Derivative		Latin, <i>fluviatilis</i> , of rivers. James in all probability chose this name because he collected its larvae from clear streamlets with grassy edges.
Type form	:	Location not known.
Reported morphological variations	*	4 in maxillary palpi and 4 in wing.
Species complex	:	Three sibling species. (known as S, T and U).
Sitting posture	•	Body and mouth parts at an angle to resting surface.
Resting habit		Adults of the species usually rest in human dwellings and to a lesser extent in cattlesheds during the day. The species also rests outdoors. Collected from an altitude of 1,060 to 3,340 m in Gum Valley of Nepal.
Breeding ecology		Preference for breeding places with a perceptible flow of water such as field channels, slow-moving streams, seepage channels of dams, and irrigation channels. Breeding is also recorded in shallow wells, tanks, ricefields (growing and fallow) along the foothills or undulating terrains, swamps, borrowpits during monsoon period when streams and channels are flushed by heavy rains.
Biting time	•	Peak biting activity occurs between 20.00 and 24.00 hrs, but timing may differ depending on the season and location.
Feeding preference		A predominantly anthropophilic species, sometimes feeds on cattle also.
Flight range	:	About 1 km.
Susceptibility to insecticides	:	Susceptible to DDT, except for a few reports of developing resistance to DDT from Pandharpur and Kurduwadi in Maharashtra.
Relation to disease	:	One of the most efficient vectors of malaria particularly in hills and foothills in India, Iran, Nepal, and Pakistan.
Reported distribution	:	Extensively found in the oriental regions and parts of West Asian subregion, Afghanistan, East and South Arabia, Bahrain, Iran, Iraq, Kazakh (USSR), Oman, and Pakistan. In oriental regions it occurs in Afghanistan, Bangladesh, South China, India, Indochina, Myanmar,

Nepal, Pakistan, Taiwan, and Thailand. In India, occurs in all mainland zones in Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Delhi, Goa, Gujarat, Haryana, Himachal Pradesh, Jammu and Kashmir, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Meghalaya, Orissa, Punjab, Rajasthan, Sikkim, Tamil Nadu, Tripura, Uttar Pradesh Terai, and West Bengal.

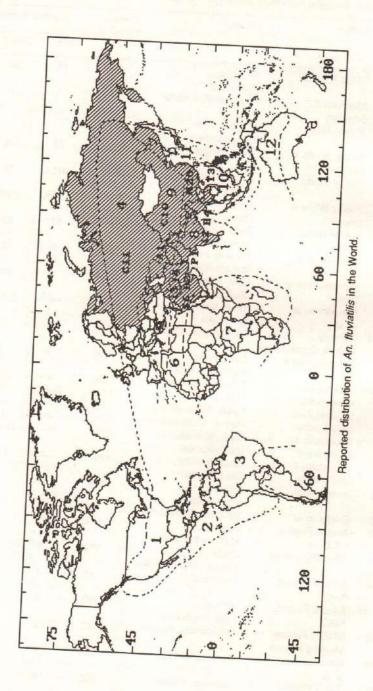
Vector incrimination: Results of studies made so far are summarized in the table.

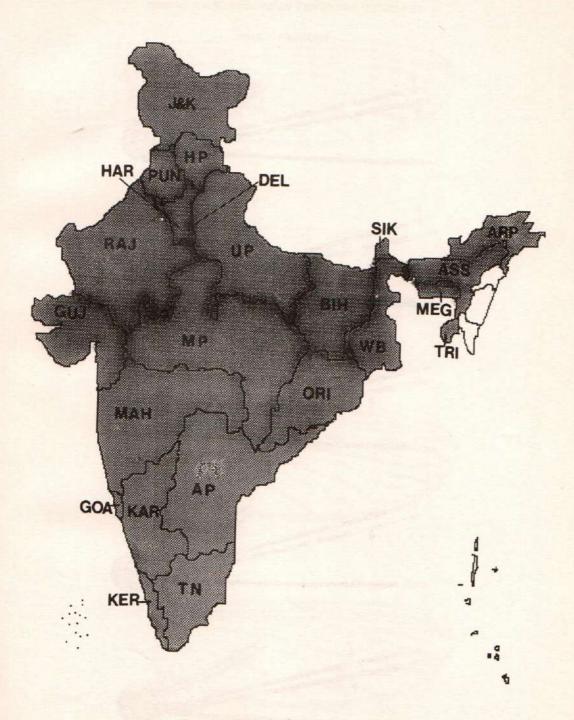
Table: Anopheles fluviatilis: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Nun	ber Positi	ve
					Gut	Gland	Total
1.	1902	James	Assam	NM		_	6%
2.	1914	Perry	Jeypore, Orissa	273	0	4	4
3.	1923	Challam	Assam	315	0	12	12
4.	1927	Covell	Wynaad, Kerala	30	_	_	2
5.	1929	King & Iyer	Mopad, Madras, Tamil Nadu	77	3	0	3
6.	1931	Macdonald & Majid	Karnal, Haryana	23	2	0	2
7.	1934	Measham & Choudhury	Anaimallai hills, Tamil Nadu	203	10	8	18
8.	1937c	Senior White	Jeypore, Orissa	1111	80	39	119
9.	1938	Senior White	Jeypore, Orissa	760	52	23	75
10.	1938	Senior White & Das	Singhbhum hills, Bihar	1031	47	18	65
11.	1938	Annual Report, Malaria Institute	Bazpur, Nainital, Uttar Pradesh	1817	2	0	2
12.	1939	Mathew	S. Travancore, Kerala	2602	616	339	955
13.	1939	Covell & Harbhagwan	Wynaad, Kerala	2781	445	213	658
14.	1939	Covell & Harbhagwan	Chedleth	374	22	11	33
15.	1940	Senior White & Narayana	Singhbhum hills, Bihar	81	1	1	2
16.	1940	Senior White & Adhikari	Satpura, Madhya Pradesh	931	41	26	67
17.	1941	Rao	Bihar Prov.	813	34	16	50
18.	1941	Senior White	Hazaribagh, Bihar	2167	46	31	77
19.	1941	Senior White	Madras Prov., Tamil Nadu	218	8	1	9
20.	1941	Senior White	(Trap collection) Madras Prov., Tamil Nadu	1301	8	3	11
21.	1942	Russell & Rao	(Resting collection) Coonoor, Tamil Nadu	2580	252	262	
22.	1942	Russell & Jacob	Nilgiris Distt.	NM	9.7%	10.1%	514
23.	1943	Viswanathan & Ramachandra Rao	North Kanara, Karnataka	NM	29.6%	37.0%	-
24.	1943	Bombay Malaria Organization	Thane, Maharashtra	574	-	-	23

Table: Anopheles fluviatilis: Results of vector incrimination studies (Contd.)

SI. No.	Year	Author	Locality	Number dissected	Number Positive		
					Gut	Gland	Total
25.	1944	Singh & Jacob	N. Kanara, Karnataka	897	48	64	112
26.	1944	Abraham & Samuels	Nizamabad, Andhra Pradesh	177	1	0	1
27.	1945	Viswanathan	Bombay Prov., Maharashtra	191		-	16
28.	1946	Senior White & Ghosh	West Bengal	2032	77	54	131
29.	1946c	Senior White	E. Cen. Prov., Madhya Pradesh	3552	101	72	173
30.	1946	Senior White & Ghosh	Jeypore hills, Orissa	NM	3.8%	2.6%	-
31.	1947	Senior White et al.	Delhi, Union Territory	767	67	33	100
32.	1948	Godbole et al.	Bombay Prov., Maharashtra	981	0	1	1
33.	1948	Subramanian & Dixit	Cent. Prov., Madhya Pradesh	1324	0	17	17
34.	1948	Ramakrishnan	South Kanara, Karnataka	NM	-	8.7%	-
35.	1948-49	Bombay Malaria Organizatio	Panchmahal, Gujarat	124	0	1	1
36.	1950	Subramanian & Gupta	Cent. Prov., Madhya Pradesh	1335	0	9	9
37.	1950	Viswanathan	Bijapur, Maharashtra	980	_	_	1
38.	1950	Viswanathan	Kolaba, Maharashtra	NM	0.44%	_	_
39.	1950	Viswanathan	Nasik, Maharashtra	1475	_	-	3
40.	1950	Viswanathan	Poona, Maharashtra	6202	77	_	5
41.	1952	Srivastava & Chakrabarti	Terai, Nainital, Uttar Pradesh	NM	_	11.1%	-
42.	1952	Srivastava & Chakrabarti	Terai, Nainital, Uttar Pradesh	NM	-	1.6%	-
43.	1953	Issaris et al.	Terai, Nainital, Uttar Pradesh	7806	0	7	7
44.	1957	Bhatia et al.	Udaipur, Rajasthan	48	0	1	1
45.	1980	Annual Report, NICD	Jagdalpur, Madhya Pradesh	19571	1	1	2
46.	1983	Choudhury et al.	Gadarpur, Nainital, Uttar Pradesh	579	0	2	2
47.	1984	Annual Report, NMEP	Nagaland	44	1	0	1
48.	1984-85		Nainital, Uttar Pradesh	450	0	1	
49.	1986	NMEP, Review Meeting Report	Mayurbhanj, Orissa	81	0	3	3
50.	1989	Gunasekaran et al.	Koraput, Orissa	653	3	6	9
51.	1990	S & T Project Report	Sonapur, Assam	205	0	4	•
52.	1990	Kulkarni	Bastar, Madhya Pradesh	NM	2.08%	1.2%	,

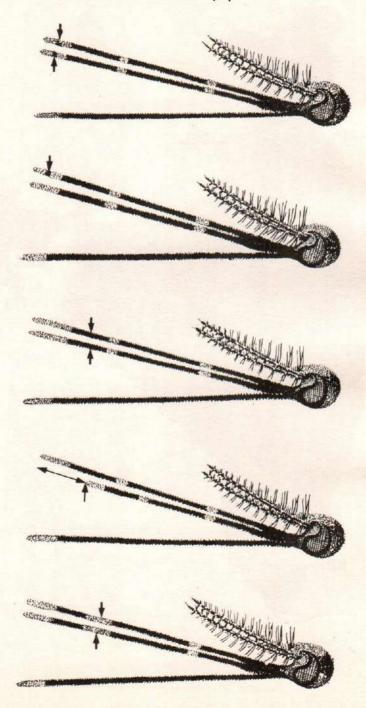




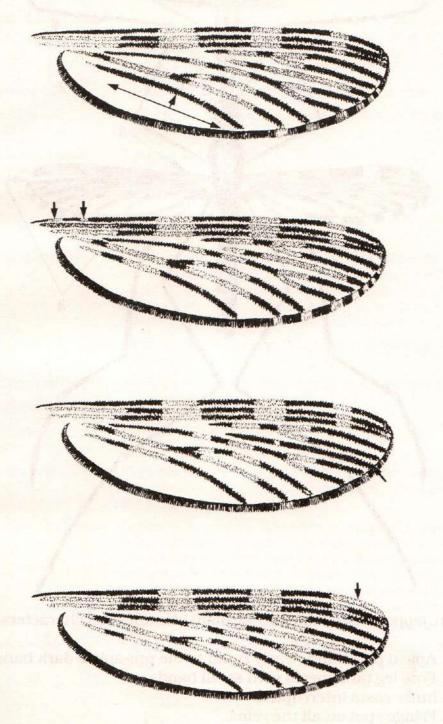
Reported distribution of An. fluviatilis In India.

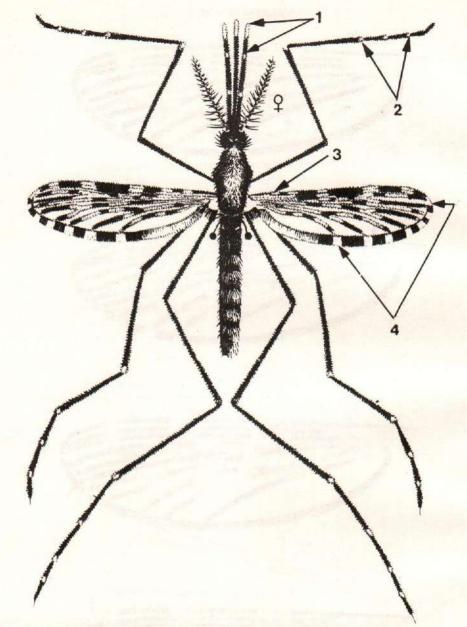
REPORTED IMPORTANT VARIATIONS OF An. fluviatilis

Variations in palpi



Variations in wing





An. jeyporiensis James 1902, main identification characters

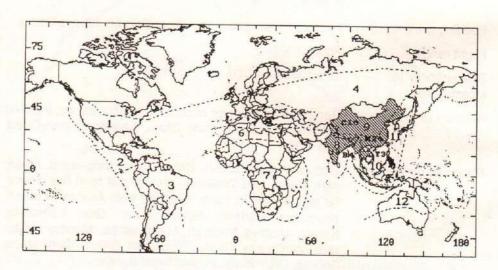
- 1. Apical pale band nearly equal to the pre-apical dark band.
- 2. Fore leg tarsomeres with small bands.
- 3. Inner costa interrupted.
- 4. Fringe spot on all the veins.

	Anopheles jeyporiensis				
Name	:	An. jeyporiensis James, 1902. Scientific Memoirs by Officers of the Medical and Sanitary Departments of the Govt. of India (New Series), 2: 106.			
Derivative	:	Named by James after Jeypore hills in Orissa India.			
Type form	:	Available at the British Museum of Natural History, London.			
Reported morphological variations	:	3 in palpi and 7 in wing			
Sitting posture	•	Body and mouth parts at an angle to resting surface.			
Resting habit	:	Species found largely in cattlesheds but also inside houses. Found at altitudes from 2000 to 6000 m.			
Breeding ecology	:	Grassy edges of slow running streams and channels as well as grassy edges of shallow tanks are ideal breeding places. The species also breeds in seepages, ricefields and a variety of breeding places with clear water, with preference for channels with marginal vegetation.			
Biting time	:	Biting activity starts after 20.00 hrs and continues till midnight.			
Feeding preference	:	Feeds on man and cattle depending on the availability.			
Flight range		About 1 km.			
Susceptibility to insecticides		Susceptible to DDT.			
Relation to disease		Regarded as a vector of secondary importance; infected specimens reported from China, Myanmar, Indo-China and India.			
Reported distribution		Occurs in Bangladesh, China, India, Indo-china, Myanmar, Nepal, and Thailand. The varietal form has a similar distribution. In India, recorded from Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Goa, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Manipur, Mizoram, Meghalaya, Orissa, Rajasthan, Sikkim, Tamil Nadu, Tripura, Uttar Pradesh, and West Bengal.			
Vector incrimination	:	Results of studies made so far are summarized in the table.			

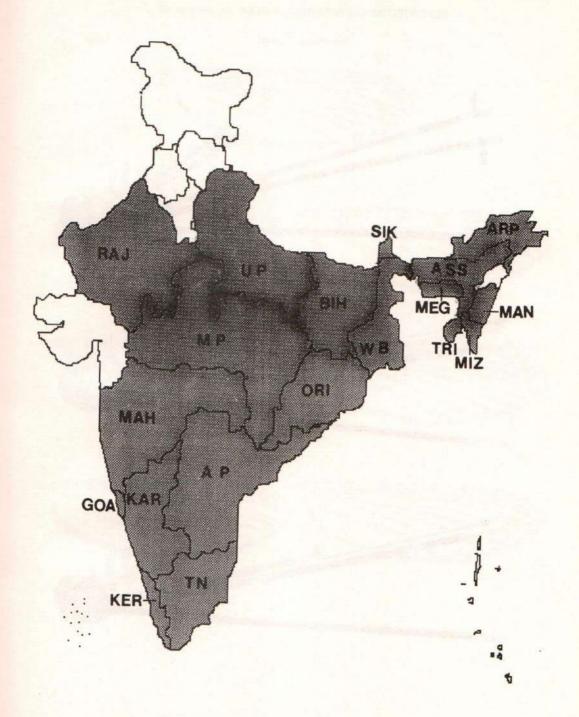
Table: Anopheles jeyporiensis: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected		Number P	ositive
					Gut	Gland	Total
1.	1934	Covell	Mysore, Karnataka	674	2	0	2
2.	1934	lyengar	Kulasekharam, Travancore, Kerala	3833	3	0	3
3.	1934	Nursing et al.	Mysore, Karnataka	1338	2	0	2
4.	1936	Ramsay et al.	Assam	2000	1	0	1
5.	1937c	Senior White	Jeypore hills, Orissa	318	4	0	4
6.	1944	Covell	Assam	2000	1	0	1
7.	1944	Covell	West Bengal	3833*	4	0	4
8.	1944	Covell	Jeypore hills, Orissa	318	4	0	4
9.	1945	Senior White et al.	Delhi, Union Territory	135*	4	0	4
10.	1945	Senior White et al.	Jeypore hills, Orissa	135	4	0	4
11.	1945	Senior White	Jeypore hills, Orissa	248	1	0	1

^{*} Variety candidiensis

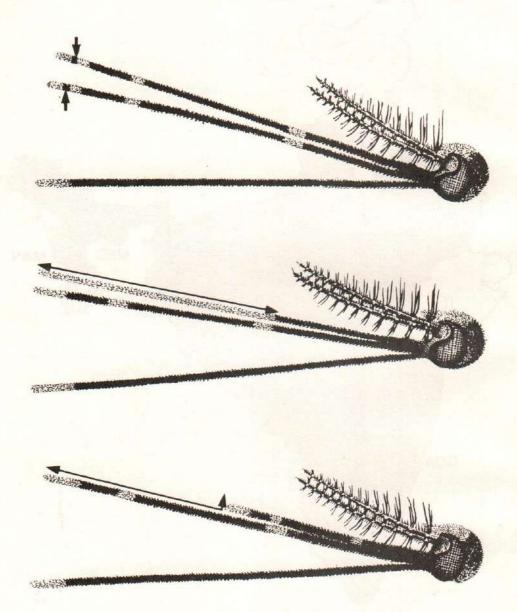


Reported distribution of An. jeyporiensis in the World.

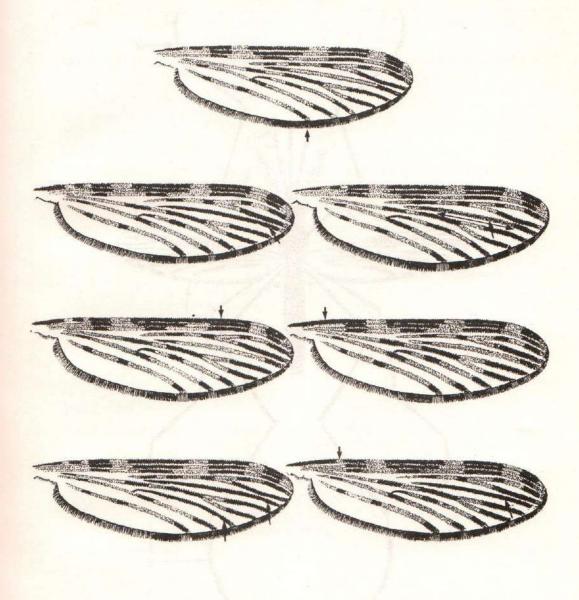


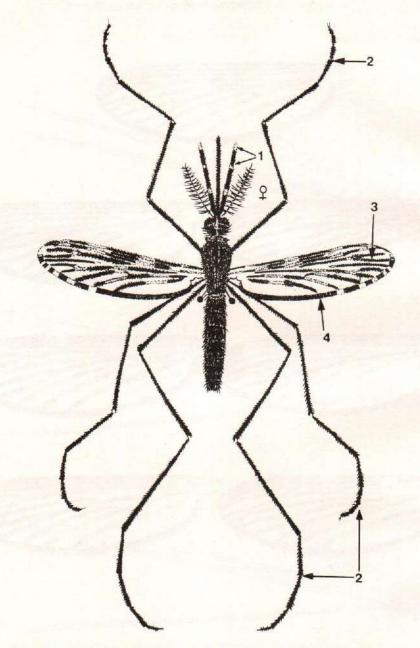
Reported distribution of An. jeyporiensis in India.

REPORTED IMPORTANT VARIATIONS OF An. jeyporiensis



Variations in wing





An. sergentii (Theobald) 1907, main identification characters

- Apical pale band nearly equal to the pre-apical dark band.
 Tarsomeres without bands.
- 3. Vein 3 (R4+5) mainly dark.
- 4. Fringe spot absent on vein 6 (Anal vein).

Anop	heles	sergentii
	110100	our goritin

Name : An. sergentii (Theobald), 1907. A Monograph of the

Culicidae or Mosquitoes, 4: 68-69.

Derivative : Theobald named it after Edmond Sergent, who sent the

specimens for identification.

Type form : Location not known.

Reported morphological variations : 2 in wing

Sitting posture

Body and mouth parts at an angle to resting surface.

Resting habit : Rests in human dwellings and cattlesheds but sometimes

in underground aqueducts.

Breeding ecology : Breeds in small pools and springs often under stones.

Sometimes breeds in ricefields and stagnant water. Prefers flowing water. Also breeds at the edge of lakes, small seepages from springs or marshes or pools associated with larger bodies of water, and seepage

channels.

Biting time : Bites outdoors and indoors before midnight.

Feeding preference : Man and cattle.

Flight range : About 4 km.

Susceptibility to : NA insecticides

Relation to disease : Occasionally found infected with human Plasmodia,

regarded as a cause of some epidemics in Egypt and

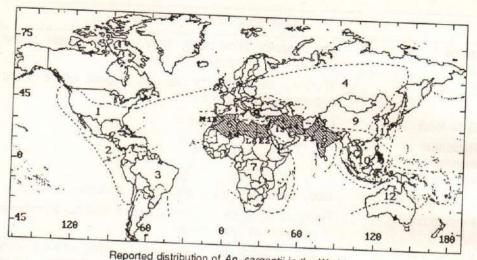
Palestine.

Reported distribution: Wide occurrence in the entire Mediterranean (North

Africa) and West Asian countries, India, and Pakistan. In India, recently recorded from Jabalpur (Madhya Pradesh)

and Koraput (Orissa).

Vector incrimination: NA



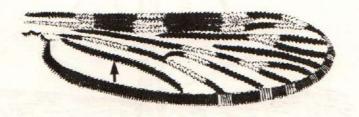
Reported distribution of An. sargentii in the World.

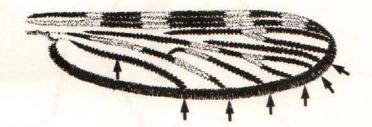


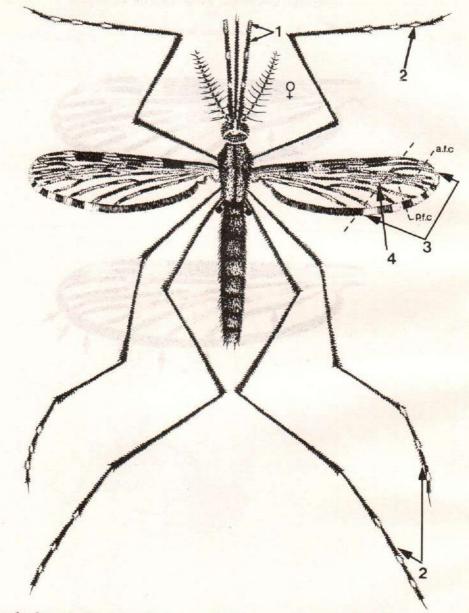
Reported distribution of An. sergentii in India.

REPORTED IMPORTANT VARIATIONS OF An. sergentii

Variations in wing







An. moghulensis Christophers 1924, main identification characters

- 1. Apical pale band nearly equal to the pre-apical dark band
- 2. Tarsomeres with bands
- 3. Fringe spot on all the veins except on vein 6 (Anal vein)
- 4. The distance of the anterior forked cell from the base of the wing is more than posterior forked cell

Ano	pheles	moghu	lensis
, ,,,,	P. 10100		

: An. moghulensis Christophers, 1924. The Indian Journal Name

of Medical Research. 12: 295-301.

: Moghul Empire; Latin, -ensis, locative suffix. Christophers Derivative

named it moghulensis as it is found from areas where the Moghul empire in India existed, i.e. from Quetta (now in Pakistan) Bombay, Belgaum, Satpura Hills, Kasauli,

Pachmarhi hills and from Central Provinces.

: Available at the British Museum of Natural History, Type form

London.

Reported morphological variations

: 1 in wing

: Body and mouth parts at an angle to resting surface.

Sitting posture Resting habit

: An uncommon species and rests in human dwellings and

occasionally in cattlesheds.

Breeding ecology

: Breeds in small rocky hill streams, and seepage waters

of shady streams.

Biting time

: NA

Feeding preference: Human and cattle.

Flight range

NA

Susceptibility to

: NA

insecticides

Relation to disease :

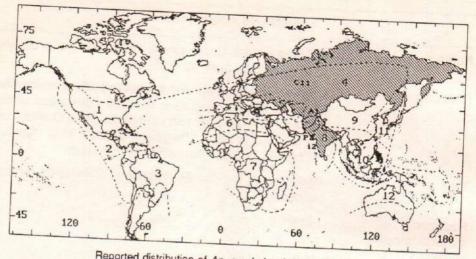
NA

Reported distribution :

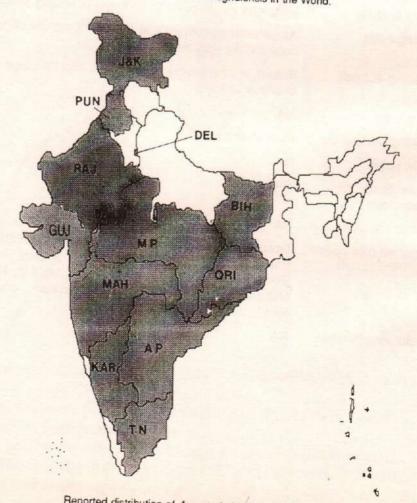
Occurs in Afghanistan, India, Kazakh (USSR), and Pakistan. In India, recorded from Andhra Pradesh, Bihar, Karnataka, Jammu & Kashmir, Madhya Pradesh, Maharashtra, Orissa, Punjab, Rajasthan, and Tamil

Nadu.

Vector incrimination : NA



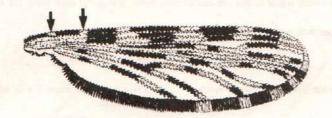
Reported distribution of An. moghulensis in the World.



Reported distribution of An. moghulensis in India.

REPORTED IMPORTANT VARIATIONS OF An. moghulensis

Variation in wing



1.4. An. subpictus, An. sundaicus, An. vagus

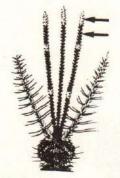
Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1)

Apical pale band nearly equal to the pre-apical dark band; pre-apical dark band 1/4 or 1/5 of apical Foreleg tarsomeres with broad pale bands

Banding pattern of palpi

Apical pale band nearly equal to the pre-apical dark band

Pre-apical dark band 1/4 or 1/5 of the apical pale band



An. subpictus, An. sundaicus



An. vagus

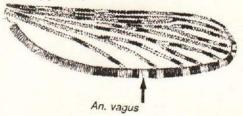
Other characters

Tarsomeres of foreleg with pale bands.

Legs without speckling

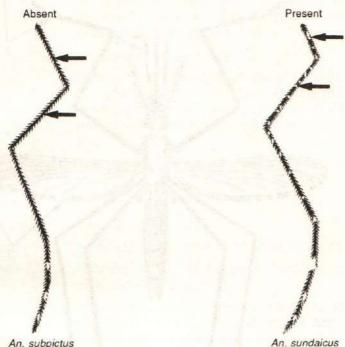


Fringe spot between veins 5(Cu) and 6(Anal) usually present



An. subpictus, An. sundaicus

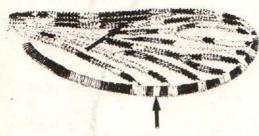
Speckling in legs



An. subpictus

Other characters

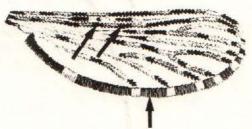
Main costal spot with one dark spot on vein 1(R1). Fringe spot between veins 5(Cu) and 6(Anal) usually present



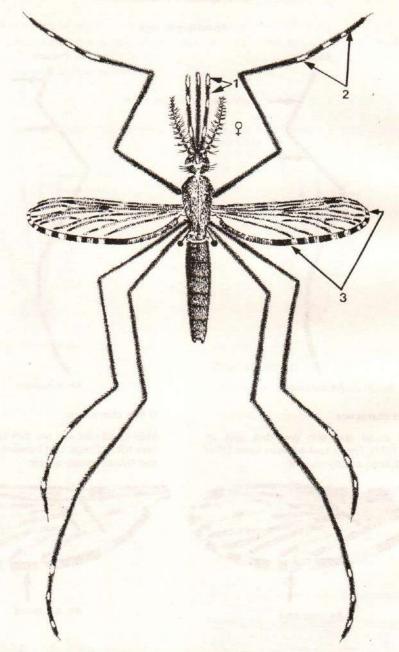
An. subpictus

O ther characters

Main costal spot with two dark spot areas on vein 1(R1). Fringe spot between veins 5(Cu) and 6(Anal) usually absent



An. sundaicus



An. subpictus Grassi 1899, main identification characters

- 1. Apical pale band nearly equal to the pre-apical dark band
- 2. Fore leg tarsomeres with broad bands
- 3. Fringe spot on all the veins

Ano	phel	es	sub	pictus

Name : An. subpictus Grassi, 1899. In: Grassi, Bignami and

Bastianelli, 1899. Atti dell, Accademia Nazionale dei

Lincei. Rendiconti, 8: 100-104.

Old name : An. rossii Giles.

Derivative : In this case sub is used in one of its literal meanings,

i.e. less than or smaller than. Grassi refers to it as "Forma piccola dell' India", comparing it with pictus as pseudopictus which he called the second small form of

Italy.

Type form : Available at the Rome University Museum, Rome.

Reported morphological variations

11 in maxillary palpi and 1 in wing.

Species complex : Two sibling species.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Very commonly found in houses and cattlesheds. Often

collected outdoors.

Breeding ecology : Breeds in a variety of habitats except the highly polluted

or contaminated ones. In flowing or stagnant waters, clear or turbid waters, water with or without vegetation, unshaded or slightly shaded places, wells, borrowpits, channels, lake margins, ponds, tanks, ground pools, fallow and freshly flooded ricefields, cement cisterns, tree

holes, fresh or brackish waters.

Biting time : Bites throughout night but peak biting between 18.00 and

22.00 hrs.

Feeding preference: Feeds predominantly on cattle and other domestic

animals (zoophilic). May feed on man in the absence of

cattle.

Flight range : 1.5-6 km.

Susceptibility to insecticides

: Resistant to DDT in India.

Relation to disease : A non-vector species but recently infected specimens

with Plasmodium parasite recorded from India, Indonesia

and Java.

Reported distribution: Occurs very widely in oriental regions extending from

Afghanistan, Mariana Islands in the east; from China in the north and Sri Lanka in south. In India, occurs in all the mainland zones. Also reported from Andaman Islands

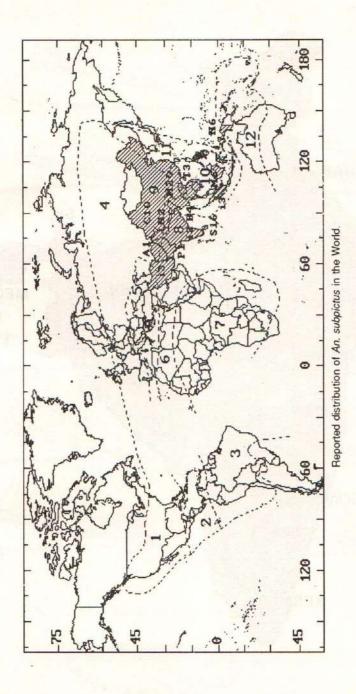
and Lakshadweep.

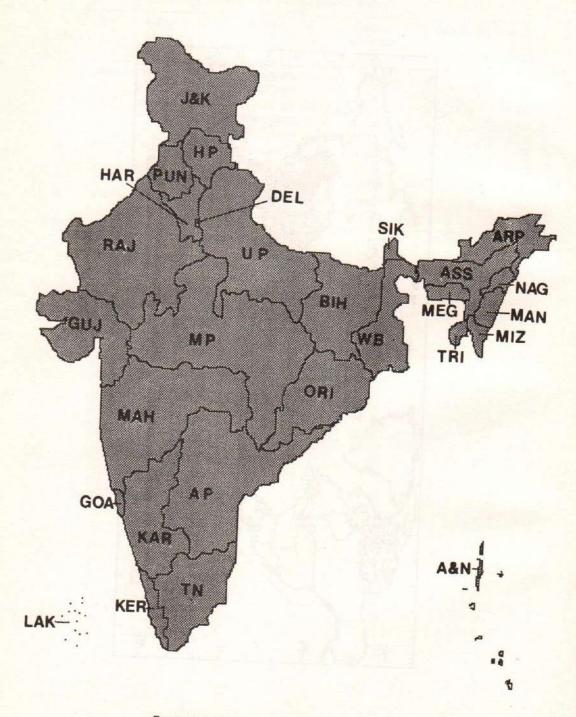
Vector incrimination : Results of studies made so far are summarized in the

table.

Table: Anopheles subpictus: Results of vector incrimination studies

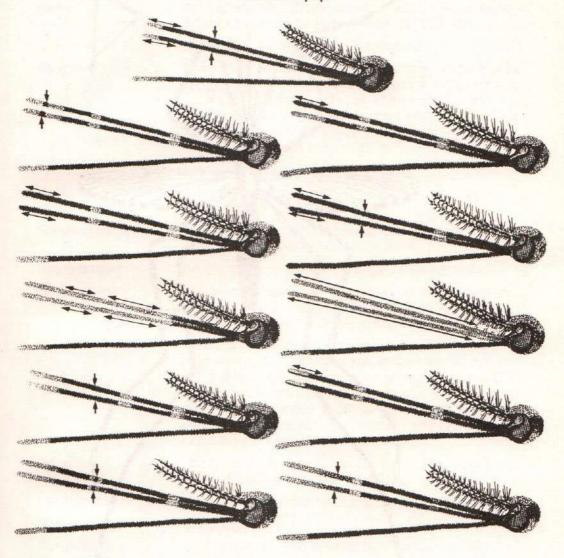
SI. Y	Year	Author	Locality	Number dissected	N	umber Posi	tive
	-3-7-2	of all residences			Gut	Gland	Total
1.	1939	Russell & Jacob	Ennore-Nellore, Tamil Nadu	4897	2	1	3
2.	1939	Russell et al.	Ennore-Nellore Pattukkottai Tamil Nadu	8381	4	1	5
3.	1940	Russell & Rao	Madras, Tamil Nadu	13277	1	1	2
4.	1981	Panicker et al.	South East India	3752	45	2	47
5.	1983	Kulkarni	Bastar, Madhya Pradesh	12107	0	3	3





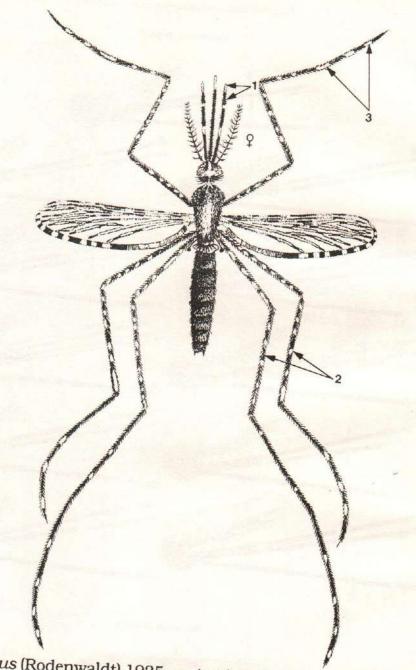
Reported distribution of An. subpictus in India.

Reported important variations of An. subpictus



Variation in wing





An. sundaicus (Rodenwaldt) 1925, main identification characters

- 1. Apical pale band nearly equal to the pre-apical dark band 2. Legs with speckling
- 3. Fore leg tarsomeres with broad bands

Anopheles sundaicus

Name : An. sundaicus (Rodenwaldt), 1925. Geneeskundig

Tijdschrift voor Nederlandsch-Indie. 65: 173-201.

Old name : An. ludlowi var. sundaicus, Rodenwaldt

Derivative : Rodenwaldt collected the specimens from Lesser Sunda

Islands, and also from Java, Sumatra and Borneo and named the species *sundaicus*. The Sunda Islands is a collective name for the Islands in the Malay Archipelago which extend from the Malay Peninsula to Timor, more

or less identical with Indonesia.

Type form : Location not known.

Reported morpho- : 4 in maxillary palpi

logical variations

Species complex : Two sibling species suspected. (One breeds in fresh

water and the other in brackish water.)

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : The species rests indoors specially in human dwelling,

cattlesheds and also outdoors, e.g. in rock crevices in

sand banks, bushes in forest areas, etc.

Breeding ecology : Mainly a brackish water breeder but also breeds in

fresh water. Major breeding places are swamps and pits along bunds, etc. containing stagnant, brackish water. Also breeds in salt-water lagoons, creeks, wells (brackish water or fresh water) overhead tanks, and freshwater pools in coastal areas. Can tolerate salinity from 0.08

to 2.6 per cent and pH from 7.7 to 8.5.

Biting time : Mostly outdoors throughout the night. The peak biting

period is between 21.00 and 2.00 hrs.

Feeding preference: Indiscriminately on cattle or man. Highly anthropophilic.

Flight range : 0.6–9 km.

Susceptibility to : Susceptible to DDT in India but resistant to DDT and

insecticides dieldrin in Indonesia.

Relation to disease : An important vector of malaria throughout its range of

occurrence in coastal areas but with some difference in the degree of transmission. Positive gut and gland specimens recorded from India, Java, Thailand, Malaysia

and Indonesia.

Reported distribution: Occurs in Bangladesh, China, India, Indonesia, Myan-

mar, Sulawesi, and Sunda Island. In India, recorded earlier from Andhra Pradesh, Orissa, Tamil Nadu, and West Bengal; it is now restricted to only Andaman Islands

where it plays a major role in malaria transmission. Recently a focus on this species was also reported from the western region of the country, from Mandvi and Mundra coastal areas of Kutch district (Gujarat).

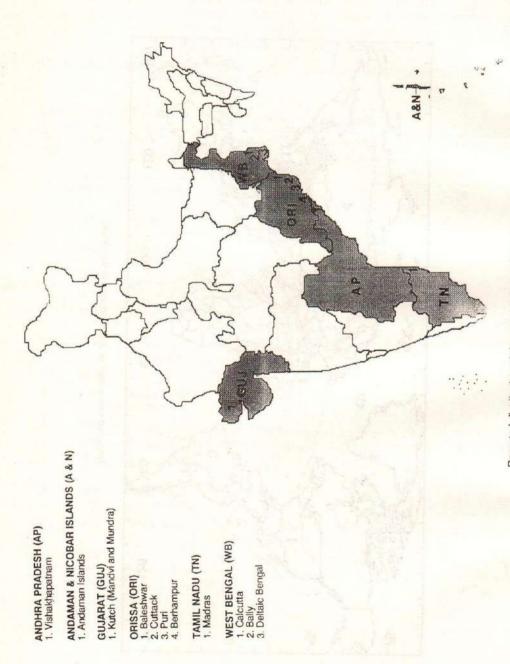
Vector incrimination : Results of studies made so far are summarized in the table.

Table: Anopheles sundaicus: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	N	umber Posi	tive
					Gut	Gland	Tota
1.	1912	Christophers	Andaman Island, Union Territory	53	2	0	2
2.	1914b	Hodgson	Madras, Tamil Nadu	NM	_	1	1
3.	1927	Covell	Andaman Island, Union Territory	98	1	1	2
4.	1931	lyengar	Howrah, West Bengal	71	0	3	3
5.	1931	lyengar	Industrial area, West Bengal	838	71	169	240
6.	1936	Ramsay & Macdonald	West Bengal	1593	0	175	175
7.	1938	Sen	West Bengal	124	2	2	4
8.	1939	Senior White & Adhikari	Chilka lake, Orissa	659	10	5	15
9.	1940	lyengar	West Bengal	19	3	1	4
10.	1942	Panigrahi	Puri town, Orissa	617	5	7	12
11.	1942	Covell & Singh	Chilka lake, Orissa	10714	32	51	83
12.	1947	Senior White et al.	Chilka lake, Orissa	1059	13	12	25
13.	1947	Senior White et al.	N. Vizagapatnam, Andhra Pradesh	1280	60	33	93
14.	1948a	Sen	West Bengal	373	_		29

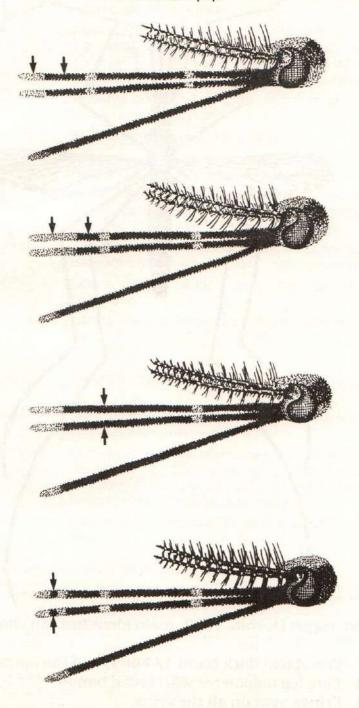


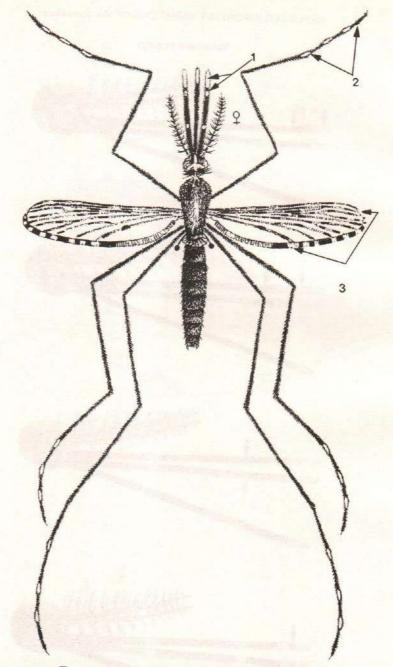
Reported distribution of An. sundaicus in the World.



Reported distribution of An. sundaicus in India.

REPORTED IMPORTANT VARIATIONS OF An. sundaicus





An. vagus Doenitz 1902, main identification characters

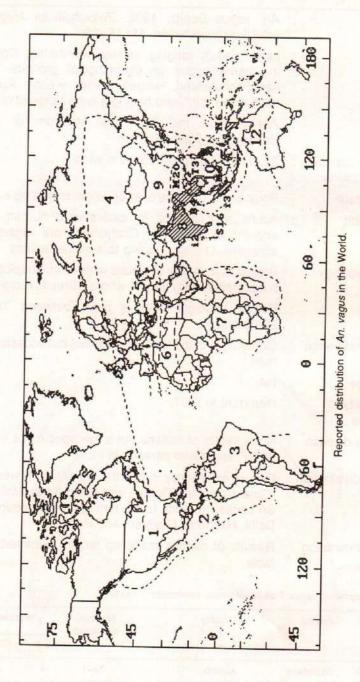
- 1. Pre-apical dark band 1/4 or 1/5 of the apical pale band.
- 2. Fore leg tasomeres with broad bands

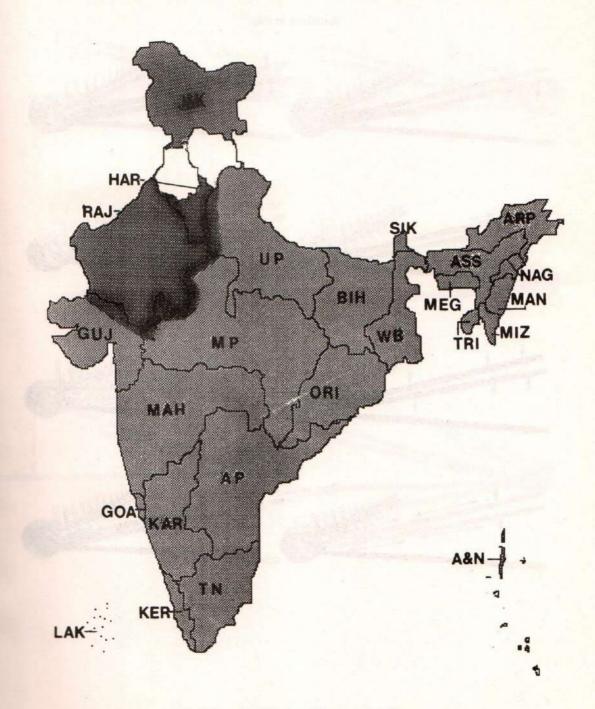
3. Fringe spot on all the veins.

		Anopheles vagus
Name	:	An. vagus Donitz, 1902. Zeitschrift fur Hygiene und Infektionskrankheiten, 41: 15-88.
Derivative		Latin, vagus, ranging, roving, wandering. Donitz gave the name vagus on etymological grounds "vagus = herumschweifend, wegen seiner weiten Ausbreitung nach Osten hin", and having a wide range of distribution.
Type form	:	Available at the Zoological Museum of Humboldt University of Paris, France.
Reported morphological variations	:	21 in maxillary palpi and 6 in wing
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Adults rest indoors in cattlesheds, human dwellings and mixed dwellings. Cattlesheds are regarded more attractive. Outdoor resting to a limited extent.
Breeding ecology	:	Breeds in a variety of places where An. subpictus breeds but prefers muddy waters where it breeds more intensely.
Biting time	:	Before midnight with a peak between 18.00 and 20.00 hrs.
Feeding preference	:	Generally a cattle feeder (zoophilic) but occasionally bites man.
Flight range	:	NA
Susceptibility to insecticides	:	Resistant to DDT.
Relation to disease	:	Not a vector of malaria but a few specimens were found with <i>Plasmodium</i> parasite in India.
Reported distribution	•	Occurs extensively in oriental regions from western India (Gujarat) to New Guinea and Hong Kong, Moluccas and Sri Lanka. In India, found throughout the country except Delhi, Himachal Pradesh, and Punjab.
Vector incrimination	:	Results of studies made so far are summarized in the table.

Table: Anopheles vagus: Results of vector incrimination studies

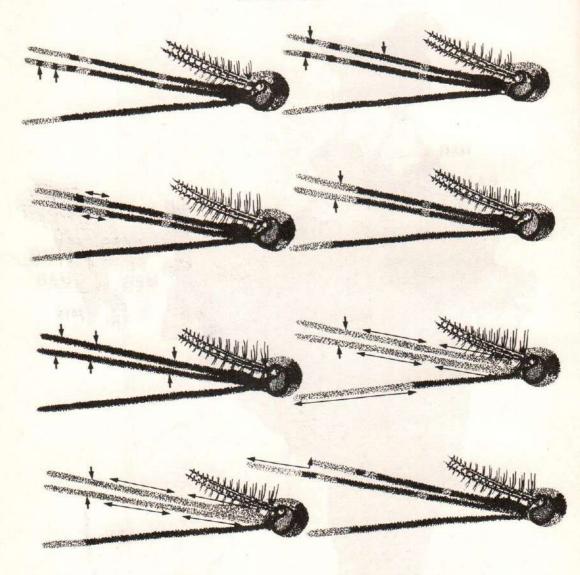
SI. No.	Year	Author	Locality	Number dissected	Number Positive			
		The second second			Gut	Gland	Total	
1.	1929	Strickland	Assam	1341	1	0	1	
2.	1933	Strickland et al.	West Bengal	10452	0	2	2	
3.	1939	Russell et al.	Madras, Tamil Nadu	3128	1	0	1	
4.	1940	Russell & Rao	Madras, Tamil Nadu	6874	0	1	1	

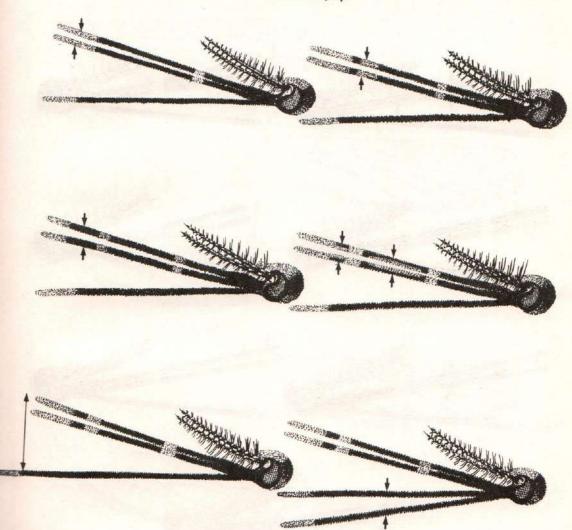


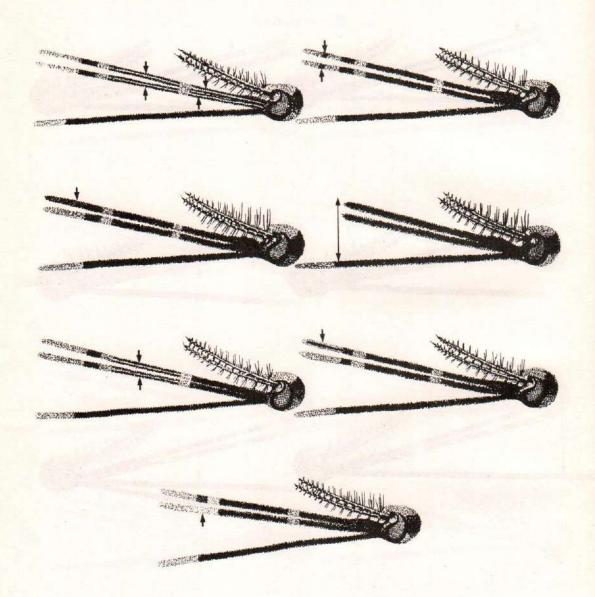


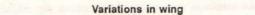
Reported distribution of An. vagus in India.

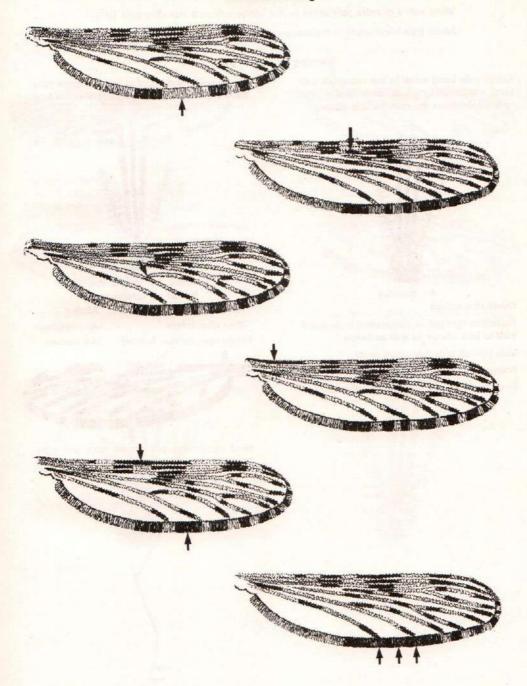
REPORTED IMPORTANT VARIATIONS OF An. vagus









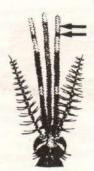


1.5. An. minimus, An. varuna, An. aconitus, An. majidi

Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1) Apical pale band equal to the subapical pale band separated by a dark band

Banding pattern of palpi

Apical pale band equal to the subapical pale band, separated by a small dark band. In most of the specimens the dark band is absent



An. aconitus

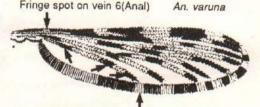
Other characters

Proboscis light yellow (flavescent) in its apical third to half, above as well as below Vein 6(Anal) with three dark spots Fringe spot on vein 6(Anal) Inner costa may or may not be interrupted Apical pale band equal to the subapical pale band separated by a dark band of equal size



An. minimus, An. varuna, An. majidi Bands on tarsomeres

Present Absent
Other characters An. minimus
Fringe spot on vein 6(Anal) An. varuna



Hind tarsomeres with broad dark and pale bands and tip of the tarsomeres pale



An. majidi

An. minimus, An. varuna

Inner costa

Completely dark



An. varuna

Other characters

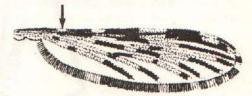
Vein 5.1(Cu1) with 2 dark areas Fringe spot absent on vein 6(Anal)



Proboscis mostly dark but sometimes half of it yellow



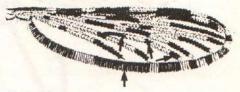
Interrupted at least in one wing



An. minimus

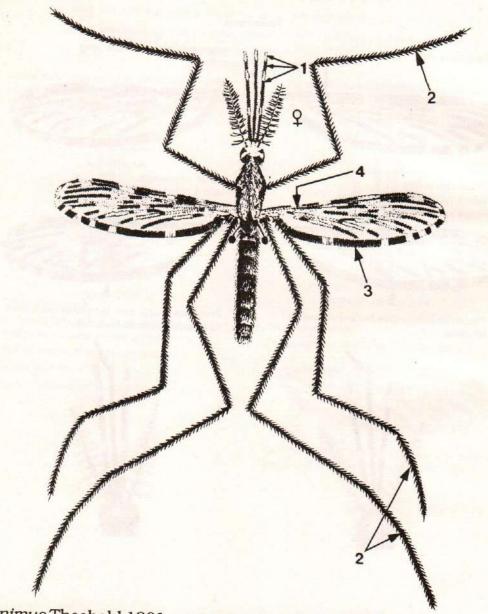
Other characters

Vein 5.1(Cu1) with 3 dark areas Fringe spot absent on vein 6(Anal)



Proboscis mostly dark but sometimes shows flavescence towards lower side





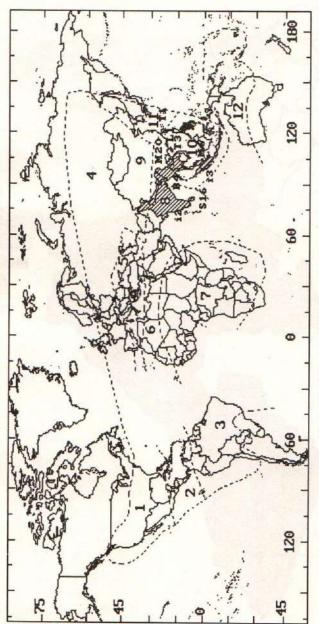
An. minimus Theobald 1901, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band. 2. Tarsomeres without bands.
- 3. Fringe spot absent on vein 6 (Anal vein).
- 4. Inner costa interrupted.

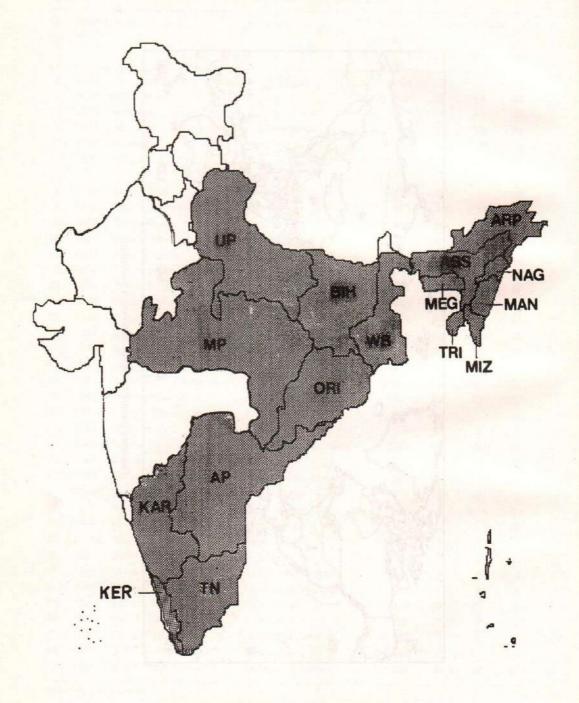
	Anopheles minimus
Name	An. minimus Theobald, 1901. A. Monograph of the Culicidae or Mosquitoes, 1: 186-188.
Derivative	Latin, <i>minimus</i> , smallest, very small; superlative of parvus, small. Theobald so named the species because of its very small size (3 mm).
Type form	Location not known.
Reported morphological variations	1 in wing
Sitting posture	Body and mouth parts at an angle to resting surface.
Resting habit	Rests in houses and cattlesheds during the day. Recenstudies show that this species rests mostly outdoors Collected up to 1,600 m.
Breeding ecology	Larvae are generally found in streams, ditches, channels in tea gardens, drains, etc. which have a perceptible but slow flow of water. Occasionally it also breeds in borrowpits, ricefields and seepages. The species prefers shady places.
Biting time	Peak biting time is between 18.00 and 19.00 hrs outdoors and between 24.00 and 2.00 hrs indoors. Biting time varies from locality to locality and from season to season.
Feeding preference	Prefers human blood and it is one of the mos anthropophilic species of the oriental region.
Flight range	More than 2 km. Migrations recorded up to 12 km.
Susceptibility to insecticides	Susceptible to DDT.
Relation to disease	The species is a primary vector of malaria all ove its range of occurrence in the foothill areas of the oriental region, e.g. Bangladesh, India, Cambodia, Laos Myanmar, Nepal, Thailand, and Vietnam.
Reported distribution	Occurs in Bangladesh, South China, India, Indochina Indonesia, Cambodia, Laos, Malaysia, Myanmar, Nepal Sri Lanka, Thailand, Vietnam, Hong Kong, Ryukyu Island and Taiwan. In India widely prevalent in Arunacha Pradesh, Assam, Madhya Pradesh, Manipur, Meghalaya Mizoram, Nagaland, and Tripura. Also recorded from scattered places in Andhra Pradesh, Bihar, Karnataka Kerala, Orissa, Tamil Nadu, West Bengal, and foothills of Uttar Pradesh.
Vector incrimination	Results of studies made so far are summarized in the table.

Table: Anopheles minimus: Results of vector incrimination studies

SI. No.	1000000	Year Author	Locality	Number dissected	Number Positive		
	SUL				Gut	Gland	Total
1.	1902	Stephens & Christophers	West Bengal	64	0	4	4
2.	1929	Strickland	Assam	1489	39	19	58
3.	1930	Ramsay	Assam	3847	59	27	86
4.	1931	Ramsay	Jorhat, Assam	1221	26	38	64
5.	1931	Manson	Assam	110	0	3	3
6.	1931	Clyde	U.P. Terai, Nainital Uttar Pradesh	1221	26	38	64
7.	1932	Gupta et al.	Assam	1119	91	9	100
8.	1933	Manson & Ramsay	Assam	NM	1.4%	2.1%	_
9.	1933	Gupta et al.	Assam	934	41	12	53
10.	1936	Paul et al.	Assam	NM	0.67%	0.67%	1.35%
11.	1937	Niogi & Khan	Jalpaiguri, West Bengal	8385	_	- 0.07 /6	513
12.	1937c	Senior White	Jeypore hills, Orissa	201	17	8	25
13.	1938	Senior White & Das	Singhbhum hills, Bihar	334	23	13	36
14.	1938	Senior White	Jeypore hills, Orissa	7195	367	295	662
15.	1939	Gilroy	Darjeeling, West Bengal	421	35	6	41
16.	1939	lyengar	West Bengal	719	25	18	43
17.	1940	lyengar	West Bengal	172	20	16	36
18.	1940	Senior White & Narayana	Singhbhum hills, Bihar	124	12	7	19
19.	1941	Anderson & Viswanathan	Assam	14092	393	391	784
20.	1941	Viswanathan et al.	Assam & Meghalaya	5102	83	72	155
21.	1941	Clark & Choudhury	Assam	408		_	13
22.	1942	Khan	Assam	94	2	3	5
23.	1944	Senior White & Rao	Singhbhum hills, Bihar	2762	62	36	98
24.	1944	Covell	India	201	17	8	25
25.	1945	Senior White et al.	Jeypore hills, Orissa	161	15	7	22
26.	1946	Senior White & Ghosh	West Bengal	178	2	0	2
27.	1947	Puri & Krishnaswami	West Bengal	NM		3.1%	_
28.	1948	Ray	West Bengal	1090	_	_	32
29.	1948a	Sen	West Bengal	17	0	1	1
30.	1955	Misra & Dhar	Tripura	99	0	2	2
31.	1956	Misra	Arunachal Pradesh	7	0	1	1
32.	1982	Bhatnagar et al.	Dimapur (Nagaland)	12	1	1	1
33.	1984	Annual Report NMEP	Boko, Assam	1256	0	45	45
34.	1985	Das & Baruah	Mizoram	58	0	2	2
35.	1987	Dutta & Baruah	Arunachal Pradesh	39	0	1	1
36.	1989	S & T Project Report	Sonapur, Assam	3543	2	121	123
17.	1990	S & T Project Report	Sonapur, Assam	5019	3	148	151



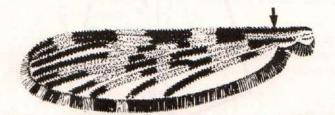
Reported distribution of An. minimus in the World.

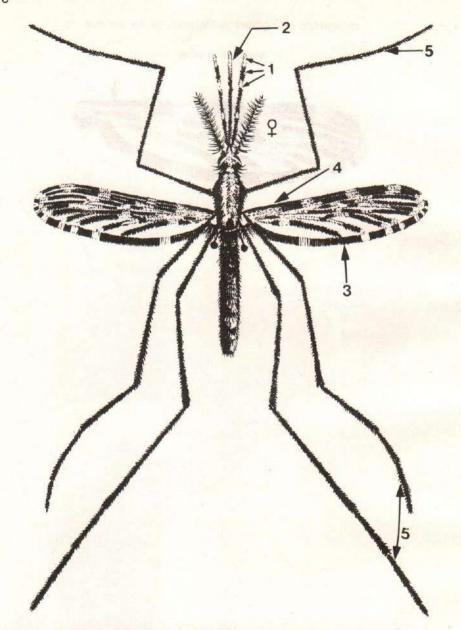


Reported distribution of An. minimus in India.

REPORTED IMPORTANT VARIATIONS OF An. minimus

Variation in wing





An. varuna lyengar 1924, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band.
- 2. Proboscis with flavescent.
- 3. Fringe spot absent on vein 6 (Anal vein)
- 4. Inner costa completely dark
- 5. Tarsomeres without bands

Anon	ha	Inn	varuna
Allopi	161	62	varuna

Name

: An. varuna lyengar, 1924. The Indian Journal of Medical Research, 12: 23-29.

Derivative

: Sanskrit, varuna, the God of Rain. This species is so named as it is the most prevalent during monsoon in Bengal, and breeds principally in rainwater collections.

Type form

: Location not known.

Reported : 1 in maxillary palpi and 2 in wing morphological variations

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests in houses, cattlesheds, and mixed dwellings. Also outdoors in foothill areas.

Breeding ecology : Breeds in a variety of places, both in stagnant and flowing waters. Breeds profusely in freshwater tanks, ponds, rice-

fields, drains, irrigation channels, wells, etc. with algal and other aquatic vocatation

other aquatic vegetation.

Biting time : Bites man mostly between midnight and dawn depending

on place. Biting time on cattle is between 18.00 and

22.00 hrs in Myanmar.

Feeding preference: Both cattle and human but prefers cattle.

Flight range : About 1 km.

Susceptibility to : NA insecticides

Relation to disease: A secondary vector of malaria in foothills of India,

Myanmar and Sri Lanka.

Reported distribution: Found in Bangladesh, South China, India, Myanmar, Sri

Lanka, and Thailand. In India, recorded from Andhra Pradesh (coastal), Arunachal Pradesh, Assam, Bihar, Goa, Gujarat, Kerala, Madhya Pradesh, Meghalaya, Orissa, Tamil Nadu, and Uttar Pradesh. Also recorded

from Lakshadweep and Andaman Islands.

Vector incrimination : Results of studies made so far are summarized in the

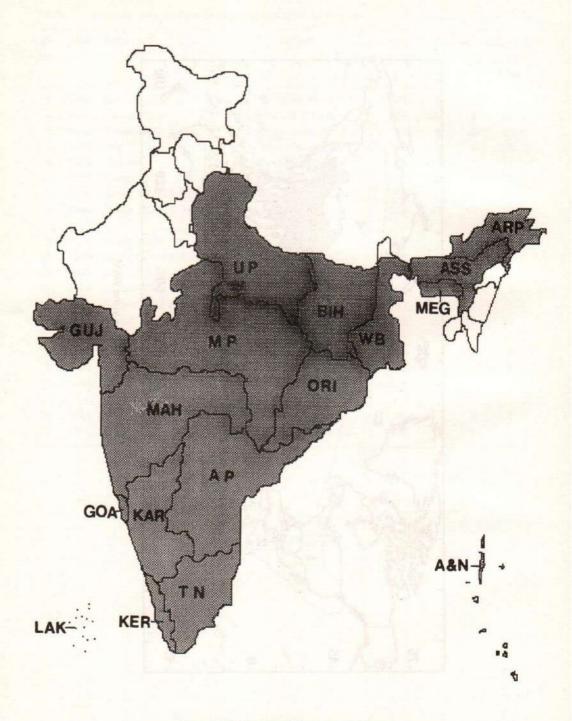
table.

Table: Anopheles varuna: Results of vector incrimination studies

SI.		Author	Locality	Number	Number Positive		
					Gut	Gland	Total
1.		., ongu	Hoogly, West Bengal	25	0	•	1997
2.	1938	Corner Million	Jeypore hills, Orissa	225	20	1	1
3.	1938	Senior White & Das	Singhbhum hills, Bihar	189	8	9	29
4.	1939	Roy	Calcutta, West Bengal	113	05700	2	10
5.	1939	Mathew	S. Travancore, Kerala	429	0	4	4
6.	1940	lyengar	West Bengal	THE REAL PROPERTY.	10	7	17
7.	1940	Senior White & Narayana	Singhbhum hills, Bihar	511	1	0	1
8.	1940	Senior White & Adhikari	E. Satpuras,	153	8	3	11
			Madhya Pradesh	386	13	12	25
9.	1941	Senior White	Madras, Tamil Nadu	In a little of			
10.	1943	Senior White & Rao	Coastal, Andhra Pradesh	59	1	0	1
11.	1943	Senior White		11000	0	2	2
12.	1943	Senior White & Rao	Hazaribagh, Bihar	131	3	2	5
13.	1943	Senior White & Rao	Madras, Tamil Nadu	762	0	1	1
14.	1945	Senior White et al.	Madras, Tamil Nadu	10567	0	1	1
15.	1945	Senior White et al.	Delhi, Union Territory	229	22	14	36
16.	1946	Senior White & Ghosh	Singhbhum hills, Bihar	2188	3	0	3
17.	1948	Sen	Bariguda, West Bengal	289	7	1	8
18.	1980	Annual Report,	West Bengal	1000	1	4	5
		NICD NICD	Jagdalpur,	19571	1	0	1
19.	1984	Mani et al	Madhya Pradesh				37
20.	1990	Kulkarni	Tamil Nadu	69	1	0	1
	1990	Nulkarni	Bastar, Madhya Pradesh	44	0	1	1



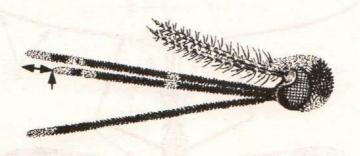
Reported distribution of An. varuna in the World.



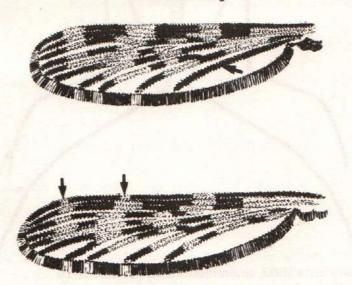
Reported distribution of An. varuna in India.

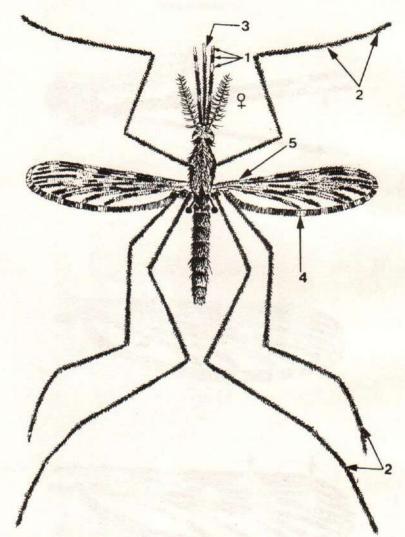
REPORTED IMPORTANT VARIATIONS OF An. varuna

Variation in palpi



Variations in wing





An. aconitus Donitz 1902, main identification characters

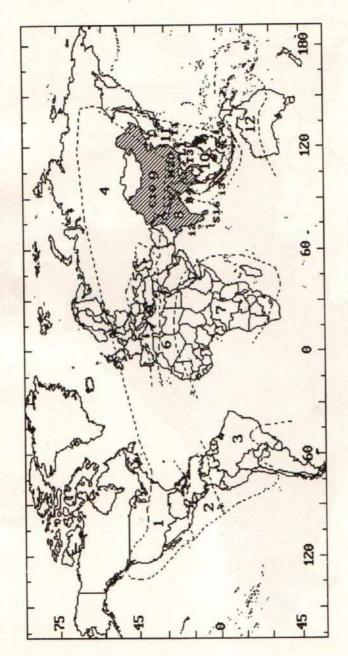
- 1. Apical and subapical pale bands equal and separated by a small dark band.
- 2. Tarsomeres without bands
- 3. Proboscis with flavescent
- 4. Fringe spot on vein 6 (Anal vein)
- 5. Inner costa interrupted

		Anopheles aconitus
Name	:	An. aconitus Dönitz, 1902. Zeitschrift Für Hygiene und Infektionskrankheiten. 41: 15–18.
Derivative	:	Greek, a, without; konis, dust, ashes; itus, adjectival suffix; Dönitz' meaning here is unequivocal.
Type form		Available at the Zoologisches Museum der Humboldt Universitaet, Berlin, West Germany.
Reported morphological variations	:	1 in maxillary palpi and 1 in wing
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Rests in houses and cattlesheds, but mostly in cattlesheds; also on bushes and banks of streams.
Breeding ecology	;	The species breeds in clean-water tanks with grassy edges, ponds, streams, water drains, river bed pools, and ricefields (at least 45 cm long). In Malaysia and Indonesia it is a swamp breeder.
Biting time		Biting starts at 18.00 hrs with a peak at 24.00 hrs and terminates at 01.00 hrs.
Feeding preference	:	Predominantly feeds on cattle but bites man to some extent.
Flight range	:	About 1 km.
Susceptibility to insecticides	:	Resistant to organochlorine pesticides.
Relation to disease		In India, regarded as a secondary vector next to An. annularis in the Orissa coastal plains. Of some importance in transmission in Java, Sumatra, Indonesia, and Malaysia.
Reported distribution	1:	Occurs throughout the oriental regions from India to Indonesia and Indochina, in south to Sri Lanka. In India, recorded from Andaman Islands, Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Delhi, Gujarat, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Manipur, Meghalaya, Mizoram, Nagaland, Orissa, Tamil Nadu, Tripura, Uttar Pradesh, and West Bengal.

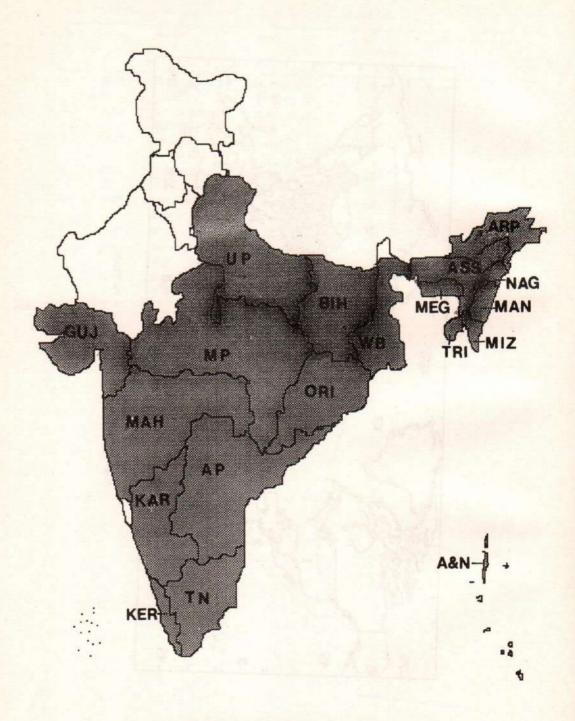
Vector incrimination : Results of studies made so far are summarized in the table.

Table: Anopheles aconitus: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive			
					Gut	Gland	Tota	
1.	1937c	Senior White	Jeypore hills, Orissa	107	1	0	1	
2.	1939	Senior White & Adhikari	Orissa, Chilka lake	481	1	0	1	
3.	1941	Anderson & Viswanathan	Assam	1145	1	0	1	
4.	1941	Viswanathan et al.	Assam	254	1	0	1	
5.	1943	Senior White et al.	Coastal Orissa	951	3	2	5	
6.	1943	Das	West Bengal	24	2	1	3	
7.	1945	Senior White et al.	Jeypore hills, Orissa	68	1	0	1	
8.	1989	Gunasekaran et al.	Koraput, Orissa	935	1	0	1	



Reported distribution of An. aconitus in the World.



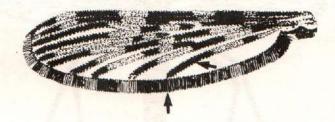
Reported distribution of An. aconitus in India.

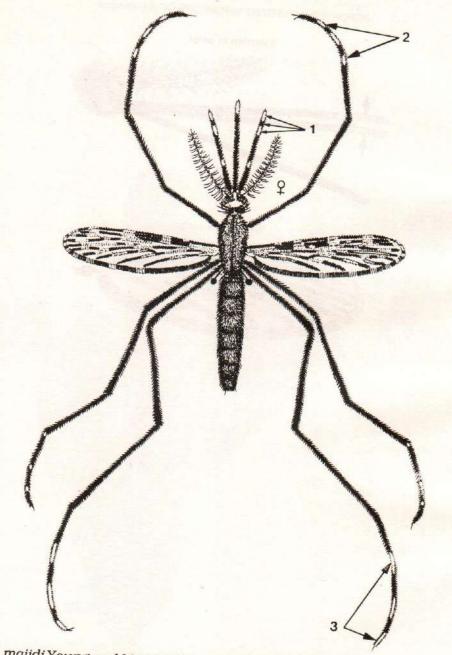
REPORTED IMPORTANT VARIATIONS OF An. aconitus

Variation in palpi



Variation in wing





An. majidi Young and Majid 1928, main identification characters

- Apical and subapical pale bands equal and separated by a dark band.
 Fore leg tarsomeres with broad bands
 Hind leg tarsomeres banded as shown in figure

Ano	phe	les	ma	iidi
, ,,,,	D	00		

: An. majidi Young and Majid, 1928. The Indian Journal of Name Medical Research, 16: 469-471.

: Species named after one of the junior authors, Majid, who Derivative

first collected it.

: Available at the National Institute of Communicable Type form

Disease, Delhi 110 054, India.

Reported morphological variations

: 1 in leg

Sitting posture

: Body and mouth parts at an angle to resting surface.

Resting habit

: A very rare species, rests in houses and cattlesheds, also

outdoors.

Breeding ecology

: Breeds in grassy slow-running streams. Breeding also recorded from open drains in tea gardens and fallow

ricefields.

Biting time

: NA

Feeding preference: Man and cattle.

Flight range

: NA

Susceptibility to

: NA

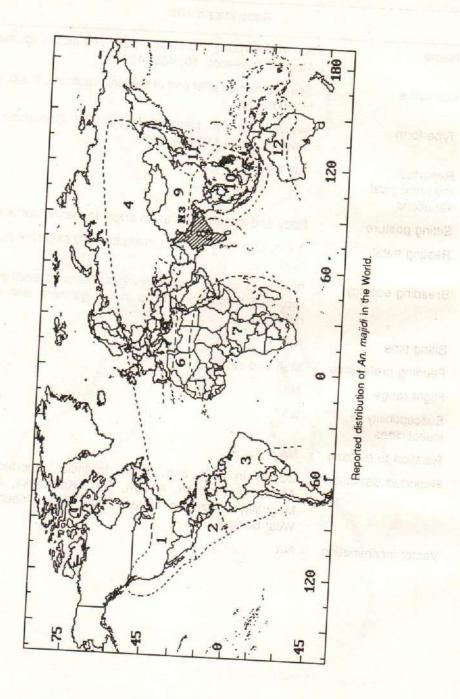
insecticides

Relation to disease: NA

Reported distribution: Occurs in India and Nepal. In India, reported from Arunachal Pradesh, Assam, Goa, Karnataka, Kerala, Meghalava, Mizoram, Orissa, Tamil Nadu, Tripura, and

West Bengal.

Vector incrimination : NA





Reported distribution of An. majidi in India.

REPORTED IMPORTANT VARIATIONS OF An. majidi

Variation in leg



1.6. An. stephensi, An. maculatus, An. willmorei, An. theobaldi, An. pseudowillmori

Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1)

Two apical broad pale bands of the palpi separated by a small dark band

Legs with speckling

Banding on foreleg tarsomeres

Absent

An. stephensi

1

Present

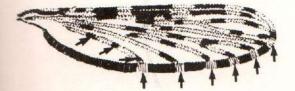
An. maculatus, An. willmorei, An. theobaldi, An. pseudowillmori

Other characters

Palpi with speckling restricted to middle and basal dark band



Fringe spot present on all the veins and vein 6(Anal) with 3 dark areas



Thorax with broad scales

An. maculatus, An. willmori, An. theobaldi, An. pseudowillmori

Palpi with or without speckling



Hind-leg tarsomeres

Fifth and fourth tarsomeres completely white



An. theobaldi

Fifth and fourth tarsomeres white but a dark band present on 4th tarsomeres



An. maculatus, An. willmori, An. pseudowillmori

An. theobaldi

Other characters

Base of costa darker than An. maculatus

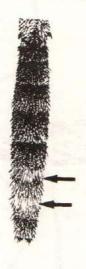


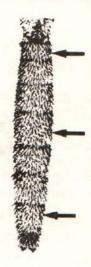
An. maculatus, An. willmori, An. pseudowillmori,

Scaling on abdomen

Broad scales restricted The whole area of to only two segments, i.e. 6 and 7

tergites 2-7 with broad golden scales





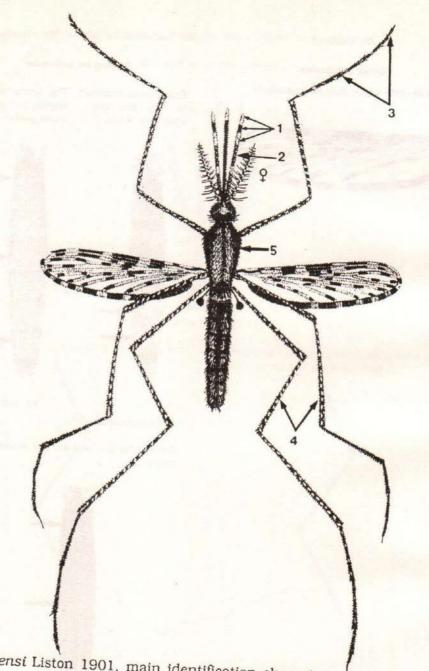
An. willmori

An. maculatus

The tergites 2-7 without any broad golden scales



An. pseudowillmori



An. stephensi Liston 1901, main identification characters

1. Apical and subapical pale bands equal and separated by a dark band. 2. Palpi with speckling.

3. Fore leg tarsomeres without broad bands.

4. Legs with speckling.

5. Thorax with broad scales.

Anophele	es st	ephensi
A SHARE THE RESERVE OF THE STREET, SAN		001101

Name : An. stephensi Liston, 1901. The Indian Medical Gazette,

36: 361-366, 441-443.

Derivative Liston named it after J.W.W. Stephens.

Type form Location not known.

Reported morphological variations

Species complex

25 in maxillary palpi and 9 in wing.

Earlier (Sweet & Rao, 1937) classified it into two varieties on the basis of ridges on egg float. These were named An. stephensi stephensi or type form and An. stephensi mysorensis. Type form is mainly urban whereas mysorensis is rural. Recent genetic studies have revealed three races, i.e. An. stephensi stephensi, An. stephensi mysorensis and An. stephensi intermediate.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Adults rest in houses, cattlesheds and barracks, etc.

Breeding ecology In urban areas, predominantly breeds in wells, overhead or ground-level water tanks, cisterns, tanks, coolers, roof gutters, and other artificial containers. Scanty breeding is also reported in recent years from ricefields, and polluted

and brackish water habitats. Larvae are shade lovers.

Biting time : Peak biting activity is generally between 22.00 and 24.00 hrs but varies from area to area and also from

season to season.

Feeding preference: Predominantly on cattle in rural areas and human in

urban areas.

Flight range : 0.8-2.5 km.

Susceptibility to : Resistant to DDT, HCH and malathion in many places in insecticide India.

Relation to disease Major urban malaria vector in India, Pakistan, Iran, and

Reported distribution : Afghanistan, Bangladesh, China, India, Indochina, Iran, Iraq, Myanmar, Nepal, Pakistan, Taiwan, and Thailand. In India, found in Andaman & Nicobar Islands, Andhra

Pradesh, Bihar, Delhi, Goa, Gujarat, Haryana, Himachal Pradesh, Karnataka, Kashmir, Madhya Pradesh, Maharashtra, Orissa, Punjab, Rajasthan, Sikkim, Tamil Nadu,

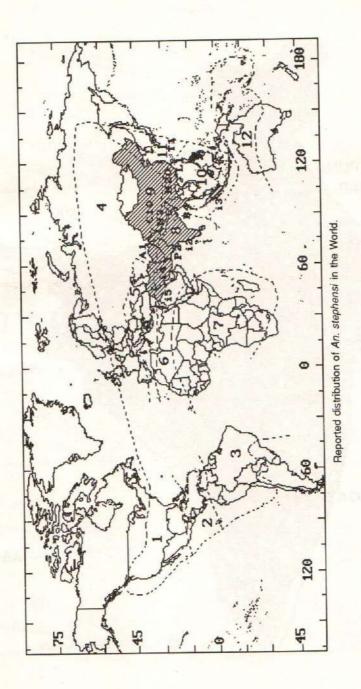
Uttar Pradesh, and West Bengal.

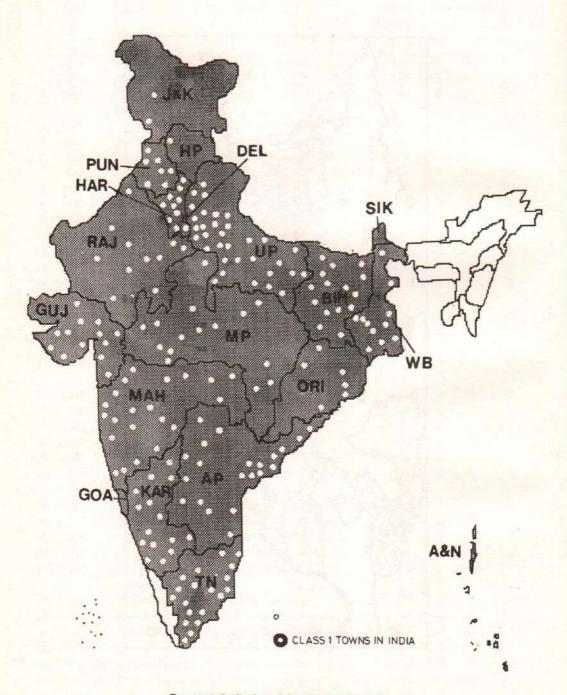
Wector incrimination : Results of studies made so far are summarized in the

table.

Table: Anopheles stephensi: Results of vector incrimination studies

S. No.	Year	ear Author	Locality	Number dissected	Number positive		
					Gut	Gland	Total
1.	1911	Bently	Bombay, Maharashtra	2445	91	30	121
2.	1914a	Hodgson	Delhi	110	2	0	2
3.	1921	Christophers and Shortt	India	17	0	2	2
4.	1927	Covell	Bombay, Maharashtra	2445	91	30	121
5.	1927	Challam	Bombay, Maharashtra	151	2	5	7
6.	1928	Covell	Bombay, Maharashtra	671	17	12	29
7.	1929	King and Iyer	Mopad, Madras, Tamil Nadu	166	9	1	10
8.	1930	Banerjee	Lucknow, Uttar Pradesh	75	7	5	12
9.	1931	Sweet and Rao	Mysore, Karnataka	2710	2	0	2
10.	1934	Nursing et al.	Mysore, Karnataka	277	3	0	3
11.	1938	Afridi et al.	Kutch State, Gujarat	238	2	4	6
12.	1938	Roy et al.	Madras Prov., Tamil Nadu	166	_	9	9
13.	1941	Rao	Bihar Prov., Bihar	423	5	7	12
14.	1943	Singh	Ahmedabad, Gujarat	155	0	2	2
0.00	1040	and Jacob	Annedabad, Odjarat	133	O	-	-
15.	1943	Senior White and Rao	Madras Prov., Tamil Nadu	151	6	4	10
16.	1943	Senior White	Vizagapatnam,	225	6	4	10
		and Rao	Andhra Pradesh				
17.	1945	Subbarao and	Madras Prov., Tamil Nadu	692	1	11	12
		Apparao					
18.	1946	Siddons	Calcutta, West Bengal	1730	4	6	10
19.	1946	Bhaskar Rao et al.	Bellay, Karnataka	902	1	1	2
20.	1948	Godbole et al.	Bijapur, Karnataka	1706	0	1	1
21.	1950	Viswanathan	Maharashtra	4706	0	1	1
22.	1950	Viswanathan	Bijapur, Kamataka	1272	0	1	1
23.	1962	Neogy and Sen	Durgapur Steel Plant, West Bengal	1613	0	1	1
24.	1962	Neogy and Sen	Burdwan, West Bengal	193	0	1	1
25.	1967	Nair and Samnotra	Broach town, Gujarat	87	0	2	2
26.	1977	Pattanayak et al.	Delhi, Union Territory	225	0	1	1
27.	1979	Das et al.	Salem, Tamil Nadu	145	2	1	3
28.	1980	Hati and	Calcutta, West Bengal	NM	0	2.5%	2.5%
		Mukhopadhyay	odiodila, rvodi beligai	1000000	ŭ	2.076	2.0 /6
29.	1982	Sharma et al.	Alwar, Rajasthan	170	0	1	1
30.	1983	Annual Report, MRC	Arthala, Uttar Pradesh	119	0	1	1
31.	1983-84	Annual Report, MRC	Delhi, Union Territory	131	0	1	1
32.	1983	Choudhury	Delhi, Union Territory		0	1	
33.	1984-85	Annual Report, MRC	Delhi, Union Territory	866	0		1
34.	1985-86	Annual Report, MRC	Delhi, Union Territory	2165		1	1
35.	1987	Choudhury and Sen	The same of the sa	469	0	1	1
			Calcutta, West Bengal	982	0	1	1
36.	1987	Hati et al.	Calcutta, West Bengal	104	0	1	- 1

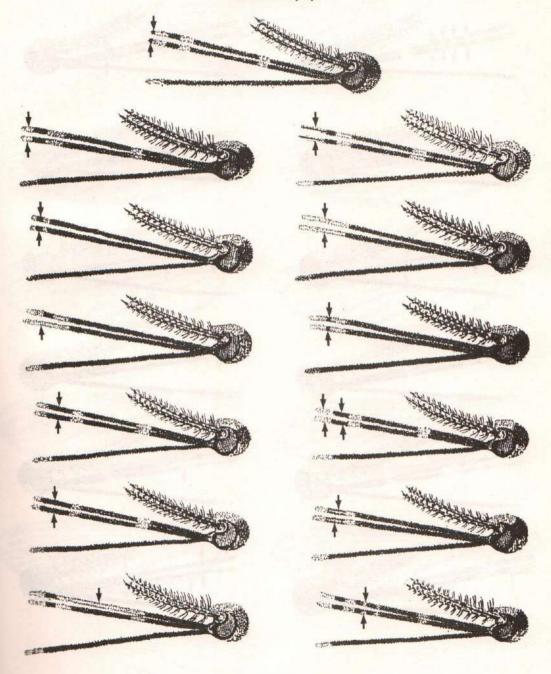


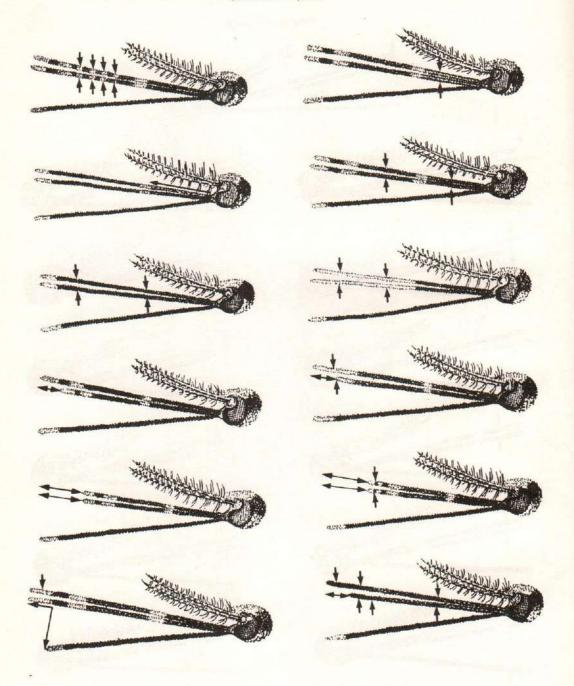


Reported distribution of An. stephensi in India.

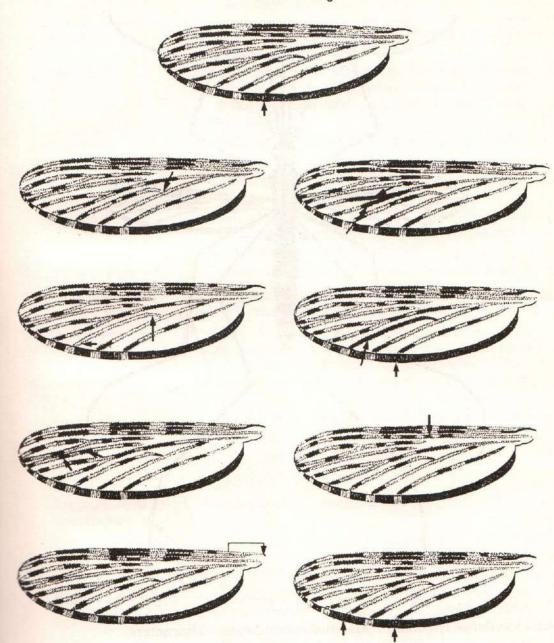
REPORTED IMPORTANT VARIATIONS OF An. stephensi

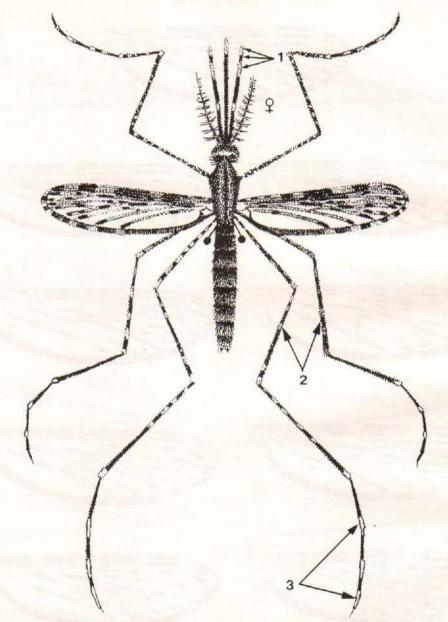






Variations in wing





An. maculatus Theobald 1901, main identification characters

- Apical and subapical pale bands equal and separated by a dark band.
 Legs with speckling.
- 3. Hind leg tarsomeres banded as shown in figure

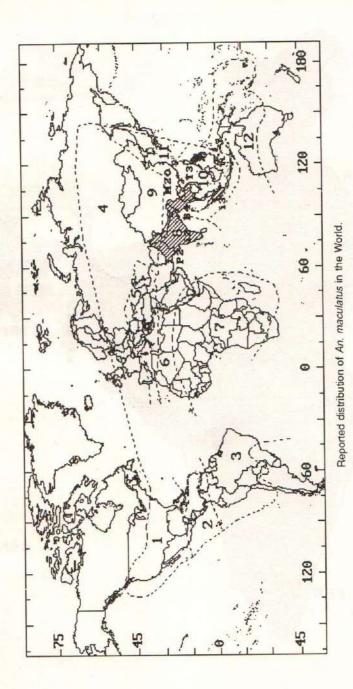
		Anopheles maculatus
Name	:	An. maculatus Theobald, 1901. A Monograph of the Culicidae or Mosquitoes, 1: 171-174.
Derivative	:	Although described as <i>maculata</i> by Theobald, it is listed as <i>maculatus</i> in Knight and Stone catalog 1977. In Latin, <i>maculatus</i> means spotted, (<i>maculare</i> to spot, to stain; <i>macula</i> , spot).
Type form	:	Available at the British Museum of Natural History, London.
Reported morpho- logical variations	:	4 in maxillary palpi, 2 in wing and 1 in leg
Species complex	•	Six species (An. maculatus, An. pseudowillmori, An. will-morei, An. sawadwongporni, An. notanandai, An. dravidieus; Rattanarithikul and Green, 1986).
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Adults generally rest outdoors in forest areas but also rest in human dwellings and cattlesheds.
Breeding ecology	:	Primarily a stream breeder, also breeds in ponds, tanks, ricefields and riverbed pools. Prefers bright sunlit breeding places and shade has a deleterious effect. Breeding is more pronounced in pre- and post-monsoon months.
Biting time	:	Biting commences soon after dusk and the peak reaches between 21.00 and 24.00 hrs, both outdoors and indoors.
Feeding preference	:	Man and cattle, but prefers cattle.
Flight range		Up to 2.2 km.
Susceptibility to insecticides	:	Resistant to DDT in Arunachal Pradesh.
Relation to disease	:	Role in malaria transmission suspected in foothills of northeastern states of India. Very important vector of malaria in Malaysia.
Reported distribution	n:	Found widely from Pakistan to Indonesia. In India, occurs in all foothill regions of the country, viz., Andhra Pradesh, Arunachal Pradesh, Assam, Bihar, Haryana, Himachal Pradesh, Jammu & Kashmir, Karnataka, Kerala, Madhya Pradesh, Maharashtra, Manipur, Meghalaya, Mizoram, Nagaland, Orissa, Punjab, Rajasthan, Sikkim, Tamil Nadu, Tripura, Uttar Pradesh, and West Bengal. Also occurs in Andaman & Nicobar Islands.

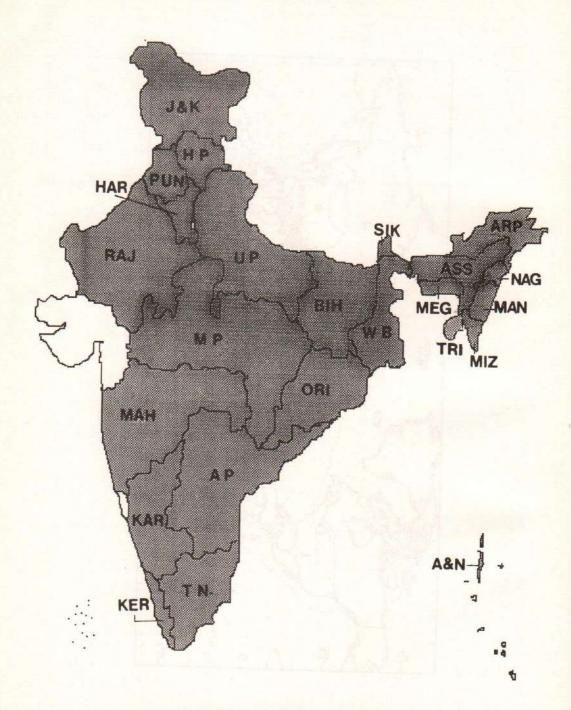
Vector incrimination : Results of studies made so far are summarized in the table.

Table: Anopheles maculatus: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive		
					Gut	Gland	Total
1.	1927	Covell	India	24	7	4	11
2.	1941	Anderson & Viswanathan	Assam	8483	24	5	29
3.	1941	Viswanathan et al.	Assam	1573	13	1	14
4.	1990	Kulkarni	Bastar, Madhya Pradesh	NM	0	1	1

NM-Not Mentioned

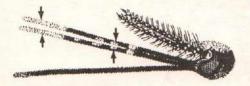


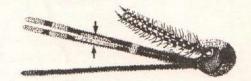


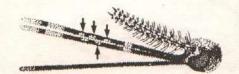
Reported distribution of An. maculatus in India.

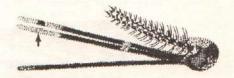
REPORTED IMPORTANT VARIATIONS OF An. maculatus

Variations in palpi









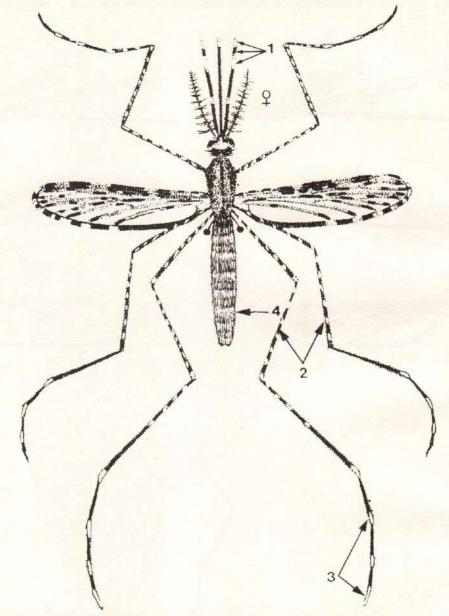
Variations in wing





Variation in leg





An. willmori (James) 1903, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band.
- 2. Legs with speckling.
- 3. Hind leg tarsomeres banded as shown in figure
- 4. Abdomen covered with golden scales.

Anopheles willmori

Name : An. willmori (James), 1903. In: Theobald, 1903. A Monograph of the Culicidae or Mosquitoes, 3: 100.

Derivative : James named it after Lieut. Willmori, IMS, who collected the species from Kashmir at 4800 ft. Previously it was the variety of An. maculatus but Rattanarithikul and Green, 1986 raised it to species level.

Type form : Available at the British Museum of Natural History, London.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests outdoors in forest areas but also prefers to rest in human dwellings and cattlesheds in foothill areas.

Breeding ecology : A stream breeder, also breeds in ponds, tanks, and pools in the bed of rivers. Prefers bright sunlit breeding places

as does An. maculatus.

Biting time : Biting starts just after the dusk and peak reaches between

21.00 and 24.00 hrs both outdoors and indoors.

Feeding preference : Man and cattle both.

: NA

Flight range : Up to 2 km.

Susceptibility to

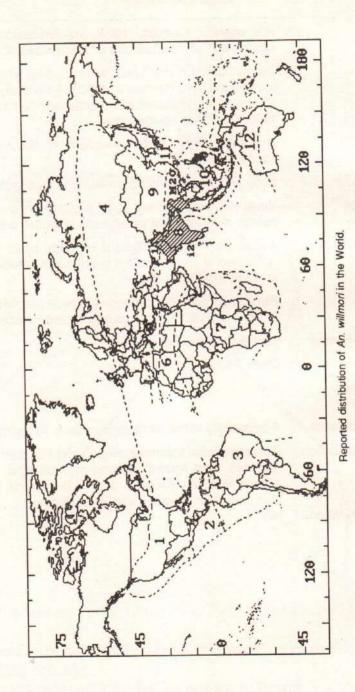
insecticides

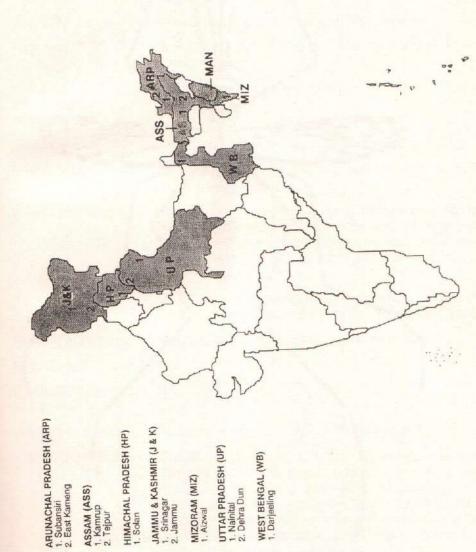
Relation to disease : A secondary vector of malaria in Gum Valley of Nepal.

Reported distribution: Occurs in India, Myanmar, Nepal, and Pakistan. In India, recorded from foothill districts of Arunachal Pradesh, Assam, Himachal Pradesh, Jammu & Kashmir, Mizoram,

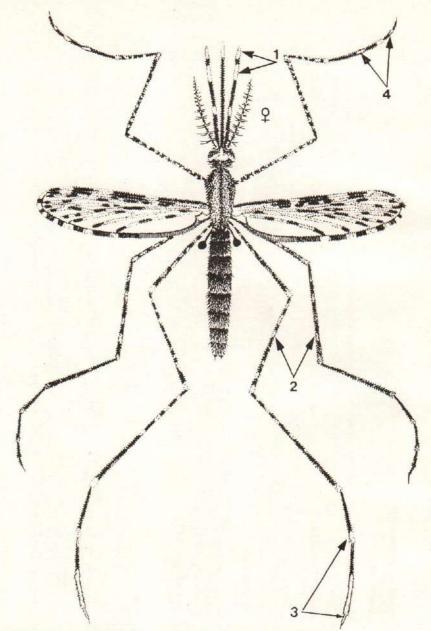
Manipur, Uttar Pradesh, and West Bengal.

Vector incrimination: NA





Reported distribution of An. willmon in India.



An. theobaldi Giles 1901, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band.
- 2. Legs with speckling
- 3. Hind leg tarsomeres banded as shown in figure
- 4. Fore leg tarsomeres with broad bands

Anopheles theobaldi

Name : An. theobaldi Giles, 1901. Entomologist's Monthly

Magazine, 37: 196-198.

Derivative : Named by Giles in 1901 after F. V. Theobald.

Type form : Available at the British Museum of Natural History,

London.

Reported morphological variations

: 2 in maxillary palpi and 1 in leg

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Adults generally rest outdoors but also rest in human

dwellings and cattlesheds.

Breeding ecology : Forest species and basically a stream breeder, also

breeds in ponds, tanks, ricefields and riverbed pools. Prefers bright sunlit breeding places, and shade has a deleterious effect. Breeds more commonly in pre- and

post-monsoon months.

Biting time : Bites outdoors and indoors, biting commences soon after

dusk, with peak activity between 21.00 and 24.00 hrs.

Feeding preference: Man and cattle, but prefers to feed on cattle.

Flight range : NA

Susceptibility to :

: NA

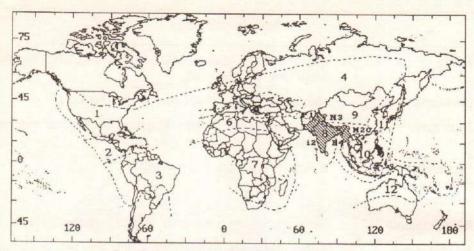
Relation to disease : So far no evidence in malaria transmission.

Reported distribution: Occurs in Bangladesh, India, Myanmar, Nepal and

Pakistan. In India, found scattered in many areas throughout the country, but mainly concentrated in the central and western parts of the Peninsula. Recently this species has been collected from many localities of Uttar Pradesh such as Nainital, Chamoli and Pithoragarh and

in West Bengal from Darjeeling district.

Vector Incrimination: NA



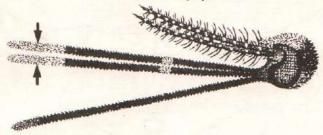
Reported distribution of An. theobaldi in the World.

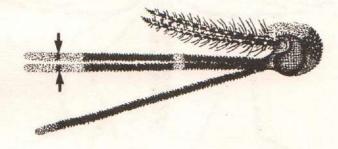


Reported distribution of An. theobaldi in India.

REPORTED IMPORTANT VARIATIONS OF An. theobaldi

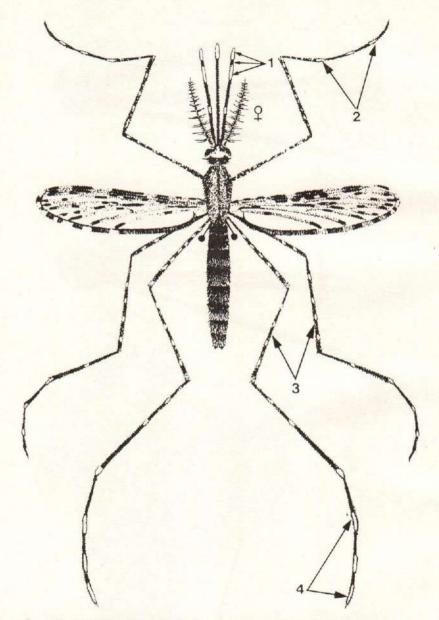
Variations in palpi





Variation in leg





An. pseudowillmoriTheobald 1910, main identification characters

- 1. Apical and subapical pale bands equal and separated by a dark band
- 2. Fore leg tarsomeres with broad bands
- 3. Legs with speckling
- 4. Hind leg tarsomeres banded as shown in figure

Anopheles pseudowillmori

Name : An. pseudowillmori Theobald 1910. A Monograph of the

Culicidae or Mosquitoes, 5: 65-67.

Derivative : The species closely resembles with the An. willmori but

differs in the wing marks and scaling on the abdomen. Previously the species was synonym of An. maculatus but Rattanarithikul and Green, 1986, raised it to species

level.

Type form : Available at the Zoological Survey of India, Indian

Museum, Calcutta, India.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : NA

Breeding ecology NA

Biting time : NA

Feeding preference: NA Flight range

NA Susceptibility to : NA

insecticides

Relation to disease : It is a non vector species in India but recently Christopher

A. Green et al. 1991 incriminated it as a Vector of malaria

in Thailand.

Reported distribution: Occurs in China, India, Nepal, Thailand and Vietnam.

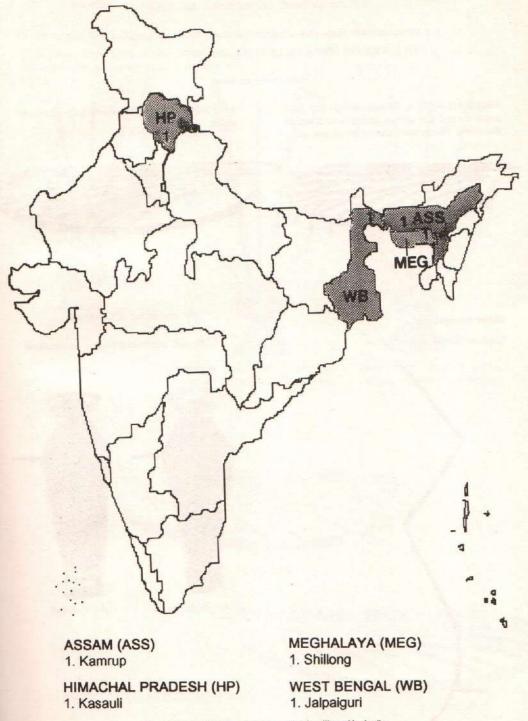
In India, recorded from Assam, Himachal Pradesh,

Meghalaya and West Bengal.

Vector incrimination : NA



Reported distribution of An. pseudowillmori in the World.



Reported distribution of An. pseudowillmori in India.

1.7. An. turkhudi, An. multicolor, An. dthali

Wing with 4 or more pale areas on the costa, subcosta including vein 1(R1) Tip of palpi dark (Apical band dark with pale hairs)

Pale areas on wing

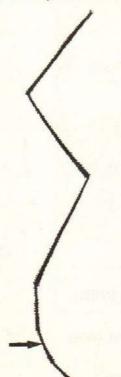
Wing veins 1(R1) to 6(Anal) without any pale spots except sometimes at the bifurcation of the veins. Fringe spot absent on all the veins Wing veins 1(R1) to 6(Anal) with pale and dark spots



An. dthali

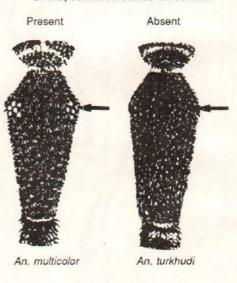
Other characters

Legs uniformly dark in colour



An. multicolor, An. turkhudi

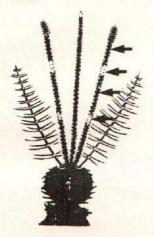
Broad, scattered scales on scutum



An. dthali

Other characters

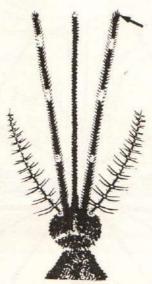
Palpi thin and uniform in thickness with two small pale bands



An. multicolor

Other characters

Tip of the palpi has dark tuft of spines

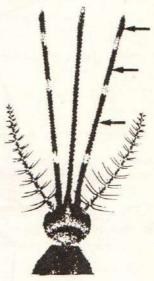


Inner costa somewhat pale, vein 6(Anal) with three dark spots

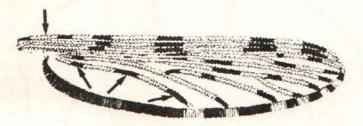


Palpi long, thin, cylindrical, and smooth

An. turkhudi



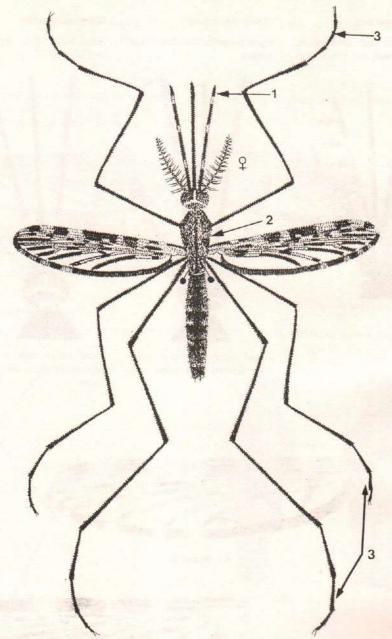
Inner costa dark in colour, wing vein 6(Anal) with two pale spots.



An. multicolor



An. turkhudi



An. turkhudi Liston 1901, main identification characters

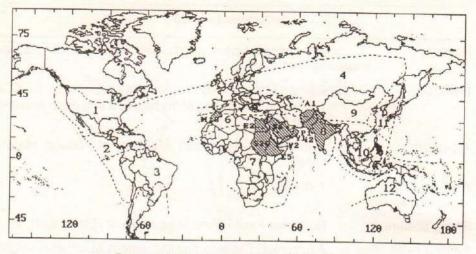
- 1. Tip of the palpi dark (Apical band dark with pale scales).
- 2. Mesonotum without broad scales.
- 3. Tarsomeres without bands

		Anopheles turkhudi
Name	:	An. turkhudi Liston, 1901. The Indian Medical Gazette, 36: 441-443.
Derivative	:	Captain Liston in 1901 named this species after Dr. D. A. Turkhud in recognition of his beautiful and exact drawings of the mosquito.
Type form	:	Available at the British Museum of Natural History, London.
Reported morphological variations	:	1 in wing
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit		Adults rest in houses and cattlesheds at altitudes ranging from 500 to 1,800 m. In Pakistan the species was collected at an altitude of 2800 m.
Breeding ecology	:	Larvae found in shallow pools on the ground among green algae, and in stream pools with a rich growth of green algae. The species also breeds in ricefields.
Biting time	:	
Feeding preference	:	Indiscriminately on man and cattle.
Flight range	:	About 1.0 km.
Susceptibility to insecticides	:	NA
Relation to disease	:	NA
Reported distribution	:	Found in countries West of India up to Aden, Afghanistan, Egypt (Sinai), Eritrea, Ethiopia, Israel, Morocco, Pakistan, Saudi Arabia, Somaliland, Sudan, and Yemen. In India, found in Andhra Pradesh, Delhi, Gujarat, Haryana, Himachal Pradesh, Jammu and Kashmir, Karnataka, Madhya Pradesh, Maharashtra, Punjab, Rajasthan, Tamil Nadu, and Uttar Pradesh.

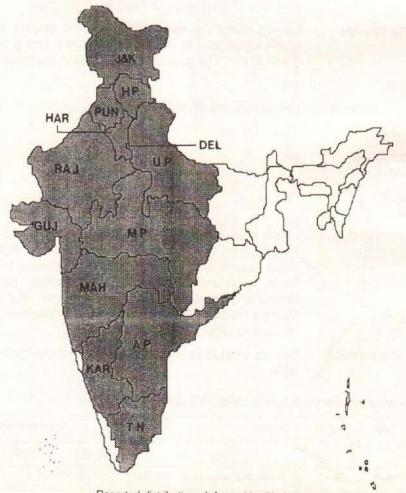
		The Start Haddon.	
Vector incrimination	:	Results of studies made so far are summarized in the	

Table: Anopheles turkhudi: Results of vector incrimination studies

SI. No.	Year	Author	Locality	Number dissected	Number Positive		
					Gut	Gland	Total
1.	1949	Bhatt	Nasik, Maharashtra	417	0	1	1



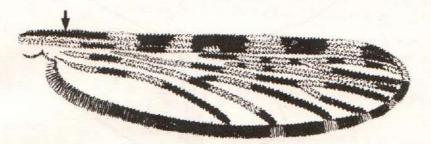
Reported distribution of An. turkhudi in the World.

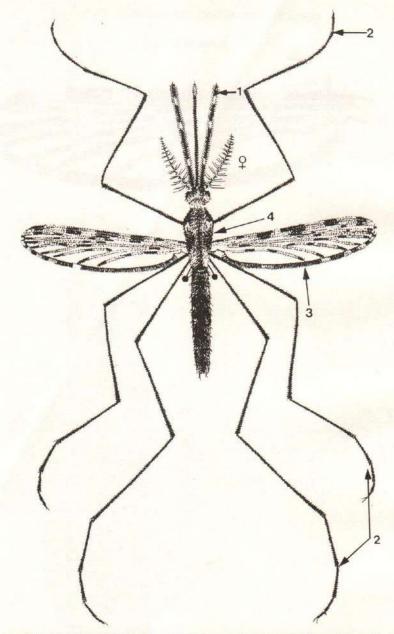


Reported distribution of An. turkhudi in India.

REPORTED IMPORTANT VARIATIONS OF An. turkhudi

Variation in wing





An. multicolor Cambouliu 1902, main identification characters

- 1. Tip of the palpi dark (Apical band dark with pale hairs)
- 2. Tarsomeres without bands
- 3. Fringe spot absent on vein 6 (Anal vein)
- 4. Mesonotum with broad scattered scales

Ano	nhel	es	multio	color
11110	PIICI	00	FILLALTIC	JUIUI

Name : An. multicolor Cambouliu, 1902. Compte Rendu de 1

Academie des Sciences, Paris, 135: 704-706.

Derivative Latin, multus, much or many; color, hue. Impressed with

many colours of this beautiful mosquito, the author gave

the name multicolor.

Type form : Available at the Faculty of Medicine, University of Paris,

Paris, France.

Body and mouth parts at an angle to resting surface. Sitting posture

Resting habit Rests in houses and cattlesheds.

Breeding ecology : Breeds in disused or abandoned wells and can withstand

salinity up to 6%.

Biting time Before midnight.

Feeding preference Man and animals.

Flight range Reported up to 13 km.

Susceptibility to NA insecticides

Relation to disease : In Sahara desert it is considered a vector on

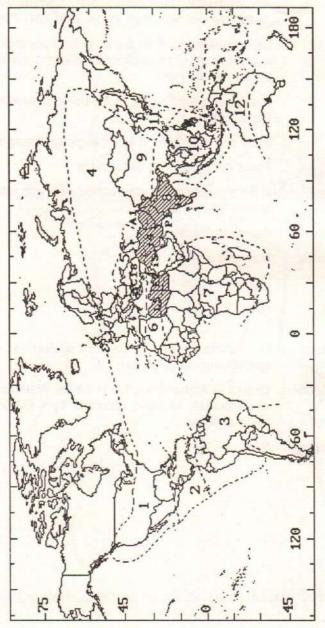
epidemiological grounds.

Reported distribution: Occurs in Afghanistan, North Africa, West Asia, Cyprus.

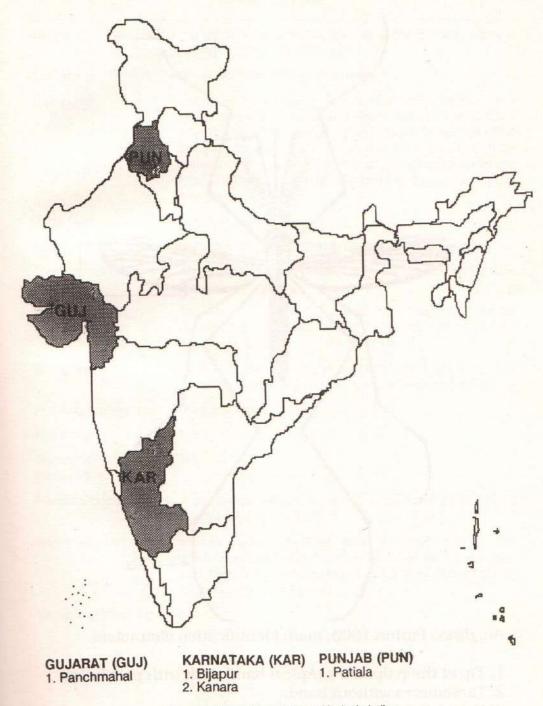
and Pakistan. In India, recorded from Gujarat, Punjab,

and Karnataka.

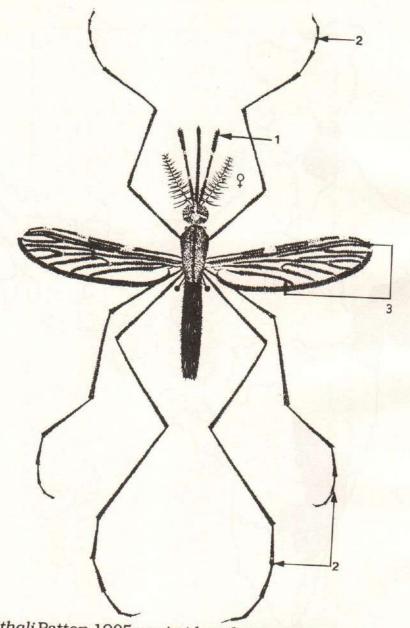
Vector incrimination : NA



Reported distribution of An. multicolor in the World.



Reported distribution of An. multicolor in India.



An. dthali Patton 1905, main identification characters

- 1. Tip of the palpi dark (Apical band dark with pale hairs)
- 2. Tarsomeres without bands
- 3. Vein 1 (RI) to 6 (Anal vein) completed dark

Anopheles dthali

Name : An. dthali Patton, 1905. Journal of the Bombay Natural

History Society. 16: 623-627.

Old name : An. rhodesiensis Theo (Eastern).

Derivative : Described by Patton in The Culicid Fauna of the Aden

Hinterland, Their Haunts and Habits. Named after D'thala village where their breeding was found in springs. The village is located at 13° 45' N, 44° 33' E on the D'thala plateau on the northern slopes of the Jehat range in the People's Democratic Republic of Yemen near the border

with Saria (Yemen).

Type form : Location not known.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Adults rest indoors in houses and cattlesheds; prefer dark

places.

Breeding ecology : Breeds in margins of rivers and streams covered

by pebbles; ricefields, water seepages are important

breeding places.

Biting time : Peak man-biting period is between 20.00 and 21.00 hrs

in southern Iran. No information available from India.

Feeding preference: Man and cattle.

Flight range : NA

Susceptibility to : NA

insecticides

Relation to disease : A secondary vector of malaria in southern parts of Iran

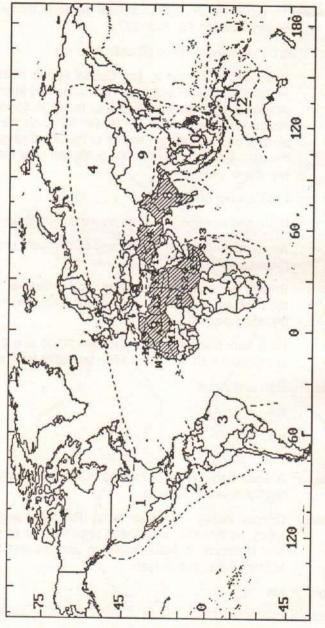
but not a vector in India.

Reported distribution: Occurs widely in West Asia (Pakistan and to the

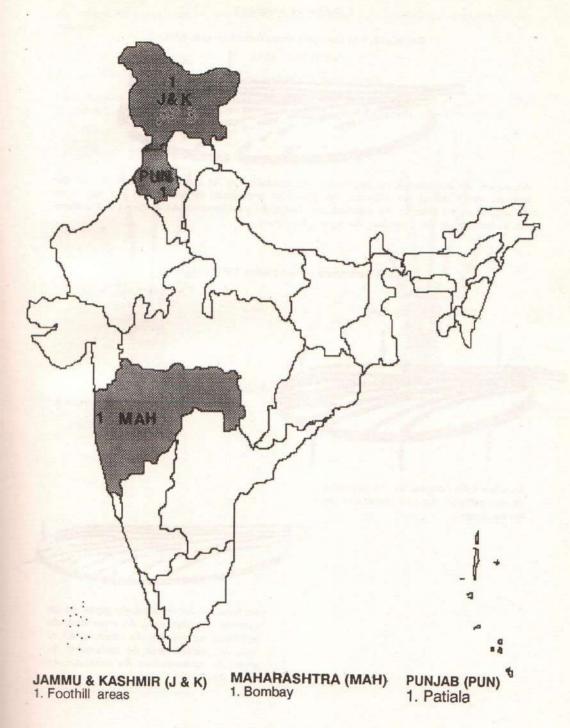
west); northern Mediterranean region as far as Ethiopia, and Morocco; in India, found in Jammu and Kashmir,

Maharashtra, and Punjab.

Vector incrimination : NA



Reported distribution of An. otthali in the World.



Reported distribution of An. dthali in India.

2. Subgenus Anopheles

Costa with 3 or less pale areas including vein 1(R1)



An. aitkenii, An. bengalensis, An. pinjaurensis, An. insulaeflorum, An. culiciformis, An. sintoni, An. barianensis, An. lindesayi, An. nilgiricus, An. gigas, An. annandalei, An. interruptus, An. nigerrimus, An. argyropus, An. sinensis, An. crawfordi, An. nitidus, An. peditaeniatus, An. barbirostris, An. ahomi, An. barbumbrosus, An. umbrosus, An. roperi, An. baileyi

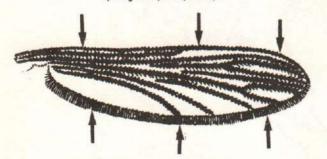
WING

Pale and dark spots on wing veins 1(R1) to 6(Anal)

Absent

(Wing completely dark)

Present



An. aitkenii, An. bengalensis, An. pinjaurensis, An. insulaeflorum, An. culiciformis, An. sintoni, An. barianensis



An. lindesayi, An. nilgiricus, An. gigas, An. annandalei, An. interruptus, An. nigerrimus, An. argyropus, An. sinensis, An. crawfordi, An. nitidus, An. peditaeniatus, An. barbirostris, An. ahomi, An. barbumbrosus, An. umbrosus, An. roperi, An. baileyi An. aitkenii, An. bengalensis, An. pinjauransis, An. insulaeflorum, An. culiciformis, An. sintoni, An. barianensis

PALPI AND HEAD

Palpi and size of the head scales

Palpi completely dark and equal to the length of proboscis. Head scales small in size

Palpi completely dark but slightly shorter than the proboscis. Head scales fairly broad



An. aitkenii, An. bengalensis, An. insulaeflorum, An. pinjaurensis, An. barianensis



An. culiciformis, An. sintoni (Sec. 2.1)

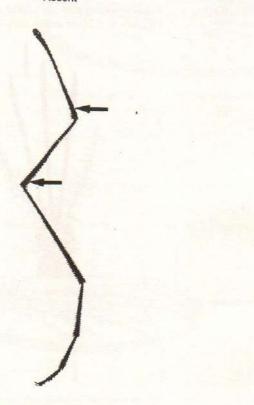
An. aitkenii, An. bengalensis, An. insulaeflorum, An. pinjaurensis, An. barianensis

HIND LEG

Pale band towards the end of hind femur and a pale ring on tibia at termination

Absent

Present



An. aitkenii, An. bengalensis, An. insulaeflorum, An. pinjaurensis (Sec. 2.2)



An. barianensis

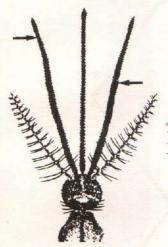
An. lindesayi, An. nilgiricus, An. gigas, An. baileyi, An. annandalei, An. interruptus, An. nigerrimus, An. argyropus, An. sinensis, An. crawfordi, An. nitidus, An. peditaeniatus, An. barbirostris, An. ahomi, An. barbumbrosus, An. umbrosus, An. roperi

PALPI

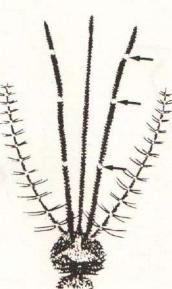
Palpi completely dark

Palpi dark but with very small pale bands at the joints

Tip of the palpi pale (four-banded palpi)



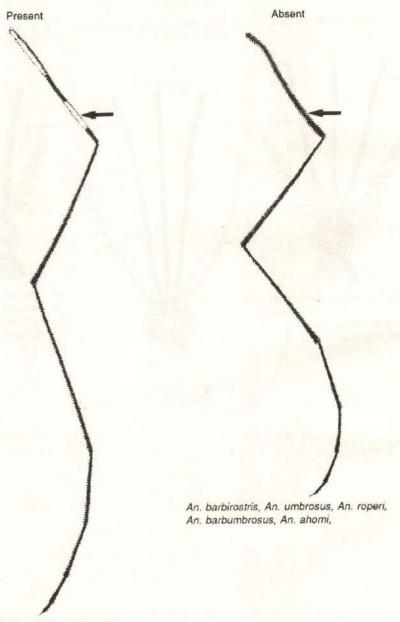
An. barbirostris, An. ahomi, An. barbumbrosus, An. roperi, An. umbrosus, An. lindesayi, An. nilgiricus



An. nigerrimus, An. nitidus, An. crawfordi, An. sinensis, An. argyropus, An. peditaeniatus, (Sec. 2.4)

An. gigas, An. baileyi, An. annandalei, An. interruptus, An. barbirostris, An. ahomi, An. barbumbrosus, An. roperi, An. umbrosus, An. lindesayi, An. nilgiricus

HIND LEG Pale area on hind femur



An. lindesayi, An. nilgiricus (Sec. 2.3)

An. barbirostris, An. barbumbrosus, An. roperi, An. umbrosus, An. ahomi

ABDOMEN

Tuft of dark scales on the ventral side of 7th abdominal segment

Absent

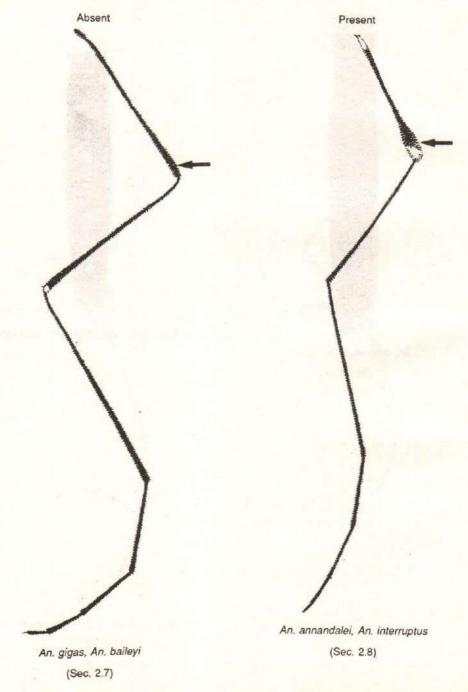
An. umbrosus, An. roperi (Sec. 2.5) Present



An. barbirostris, An. ahomi, An. barbumbrosus (Sec. 2.6)

HIND LEG

Apex of hind femur with a prominent tuft of white and black scales visible to naked eye



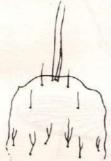
2.1. An. culiciformis, An. sintoni

Wing completely dark

Palpi shorter than proboscis, head scales fairly broad

Further identification of these two species is based on larval characters, branches, and shape of frontal (5c, 6c, 7c; clypeal hairs 5, 6, 7) lateral, subantennal (11c; clypeal hair11) and prothoracic (9p to 12p; prothoracic hairs 9, 10, 11, 12) hairs

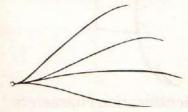
Frontal hairs (5c, 6c, 7c; clypeal hairs 5, 6, 7) with one or two branches



Subantennal hair (11c; clypeal hair 11) club shaped



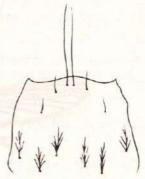
Prothoracic hair (9p to 12p; prothoracic hairs 9, 10, 11, 12) very thin but not spine-like



Lateral hair, long (present on abdominal segment 3 to 6 with very few short branches

An. culiciformis

Frontal hairs (5c, 6c, 7c; clypeal hairs 5, 6, 7), with more branches (feathered)



Subantennal hair (11c; clypeal hair 11) not club shaped but with a number of branches distally

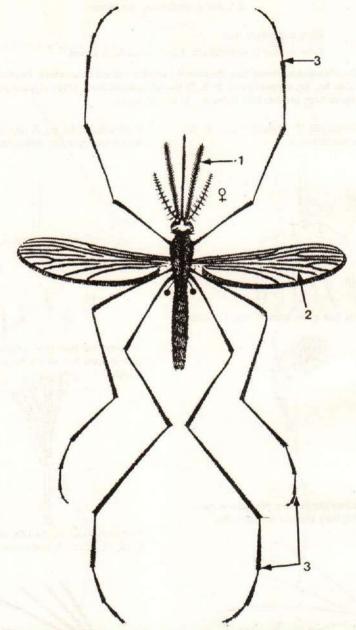


Prothoracic hair (9p to 12p; prothoracic hairs 9, 10, 11, 12) thick and spine-like



Lateral hair, with very long branches

An. sintoni



An. culiciformis Cogill 1903, main identification characters

- 1. Palpi completely dark but slightly shorter than proboscis
- 2. Wing completely dark
- 3. Legs completely dark

Anopheles culiciformis

: An. culiciformis Cogill, 1903. Journal of the Bombay Name

Natural History Society, 15: 327-336.

: Latin, Culex (genitive culicis), a gnat, midge; forma, Derivative

shape, figure, form; -is, adjectival suffix. Cogill treats culiciformis and aitkenii together, named culiciformis as it resembles Culex in sitting posture, has clear wings and

abdominal segments light in colour.

: Available at the British Museum of Natural History, Type form

London.

: Five species (An. alongensis, An. kyondawensis, Species complex

An. sintonoides, An. sintoni, and An. culiciformis).

: Body parallel to resting surface. Sitting posture

: Rests indoors preferably in human dwellings and Resting habit

outdoors mostly in tree holes.

: Forest species, breeds in tree holes and in fallen or cut Breeding ecology

bamboos.

: Before midnight. Biting time

Feeding preference : Man and to some extent wild animals.

: NA Flight range

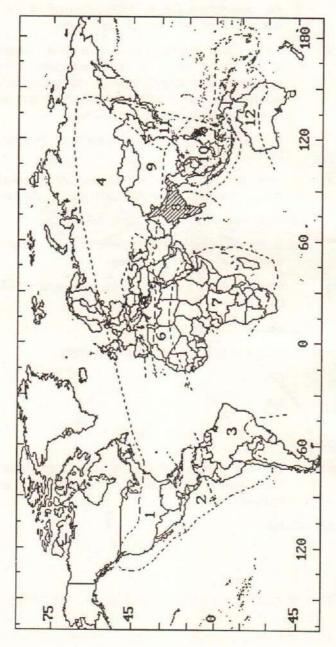
: NA Susceptibility to insecticides

Relation to disease : NA

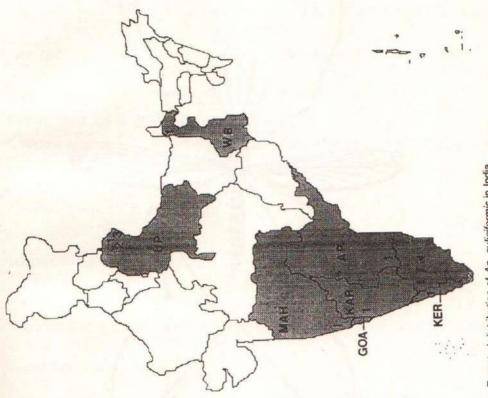
Reported distribution: In India, Andhra Pradesh, Goa, Karnataka, Kerala,

Maharashtra, Tamil Nadu, Uttar Pradesh, and West

Bengal.



Reported distribution of An. culiciformis in the World.



UTTAR PRADESH (UP)
1. Chamoli
2. Almora
3. Pithoragarh

WEST BENGAL (WB) 1. Darjeeling

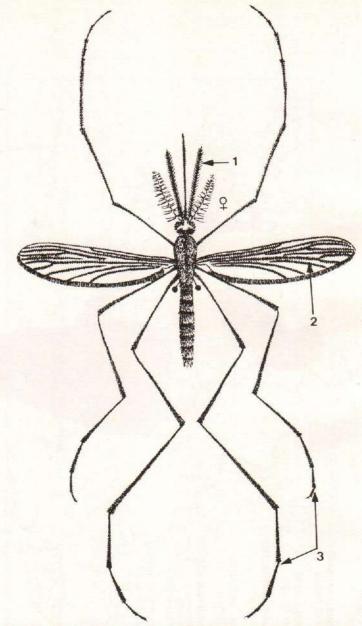
MAHARASHTRA (MAH)

1. Ratnagiri

KARNATAKA (KAR) 1. North Kanara

GOA (GOA) 1. Goa KERALA (KER) 1. Malappuram TAMIL NADU (TN)
1. Nilgiris

ANDHRA PRADESH (AP) 1. Chittoor Reported distribution of An. culiciformis in India.



An. sintoni Puri 1929, main identification characters

- 1. Palpi completely dark but slightly shorter than proboscis
- 2. Wing completely dark
- 3. Legs completely dark

Anopheles sintoni

Name : An. sintoni Puri, 1929. The Indian Journal of Medical

Research, 17: 397-404.

Derivative : Puri named it after Major J.A. Sinton, V.C., IMS, Director,

Malaria Survey of India.

Type form : Available at the British Museum of Natural History,

London.

Sitting posture : Body parallel to resting surface.

Resting habit : Rests outdoors and indoors.

: NA

Breeding ecology : Forest species, breeds in tree holes and fallen or cut

bamboos.

Biting time : Before midnight.

Feeding preference: NA

Flight range : NA

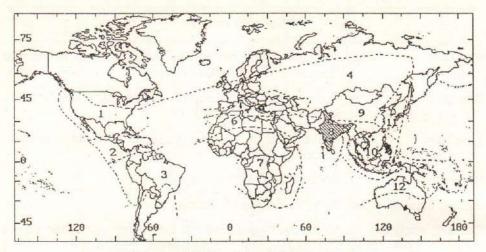
Susceptibility to

insecticides

Relation to disease : NA

Reported distribution: Recorded from the west coast of Kerala, Tamil Nadu, and

West Bengal.



Reported distribution of An. sintoni in the World.



Reported distribution of An. sintoni in India.

2.2 An. aitkenii, An. bengalensis, An. insulaeflorum, An. pinjaurensis* An. barianensis**

Wing completely dark

Head scales small in size

Femur and tibia with a pale band at their termination

Setae on prescutellar space of thorax

Present

An. insulaeflorum

The species can be confirmed by the larval characters

Larval characters

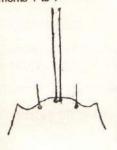
Inner clypeal hair (2c; clypeal hair 2) simple and rarely bifid; palmate hair (1-III, hair no. 1 of abdominal segment III) present on abdominal segments 1 to 7



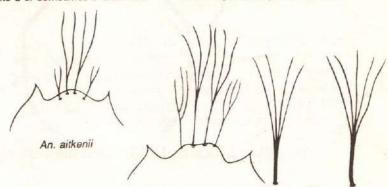
An. bengalensis An. aitkenii

Further identification based on larval characters

Branches of inner clypeal hair



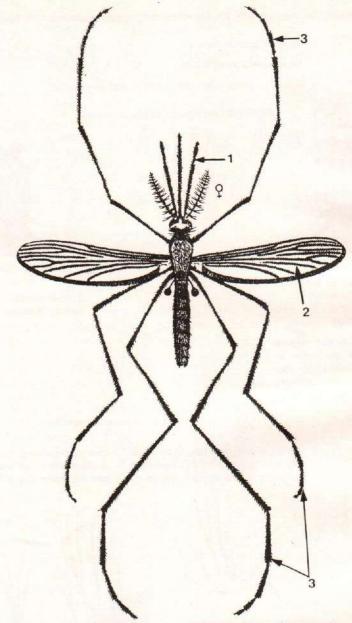
Inner clypeal hair (2c; clypeal hair 2) dividing about 1/4 way from base into 2 or sometimes 3 branches Inner clypeal hair (2c; clypeal hair 2) dividing about half way from base into 2-9 (commonly 4-7) branches



An. bengalensis

^{*}Identification of this species based on male genitalia; phalosome, very long, somewhat expanded at the tip

^{**}Identification of this species is given on page number 278



An. aitkenii James 1903, main identification characters

- Palpi completely dark
- 2. Wing completely dark
- 3. Legs completely dark

Ano	nha	lac	aitk	anii
AIIU	DITE	162	allh	CIIII

Name : An. aitkenii James, 1903. In: Theobald, A Monograph of

the Culicidae or Mosquitoes, 3: 22-23.

Derivative : James named it after E.H. Aitken (1851-1909) who

collected this species in the Goa frontier and Karwar

(Bombay Presidency).

Type form : Available at the British Museum of Natural History,

London.

Species complex : Ten species (An. palmatus, An. pinjaurensis, An. fragilis,

An. tigertti, An. aitkenii, An. bengalensis, An. borneensis, An. acaci, An. insulaeflorum, and An. stricklandi).

Sitting posture : Body parallel to resting surface.

Resting habit : Rests in shades of jungles and rarely enters houses.

Breeding ecology : Breeds in a variety of places such as small pools and

seepages in the jungle, tea-garden drains which are shaded by tea plants, swamps, marshes, channels, river, rock pools, streams heavily shaded with trees, wells, etc.

Biting time : Exophilic and bites during the day.

Feeding preference: Wild animals but sometimes man.

Flight range : NA

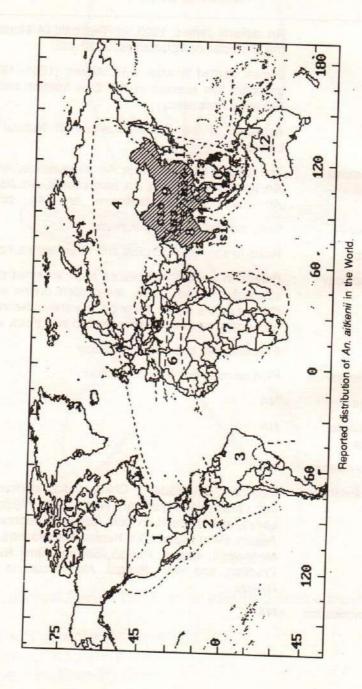
Susceptibility to : NA insecticides

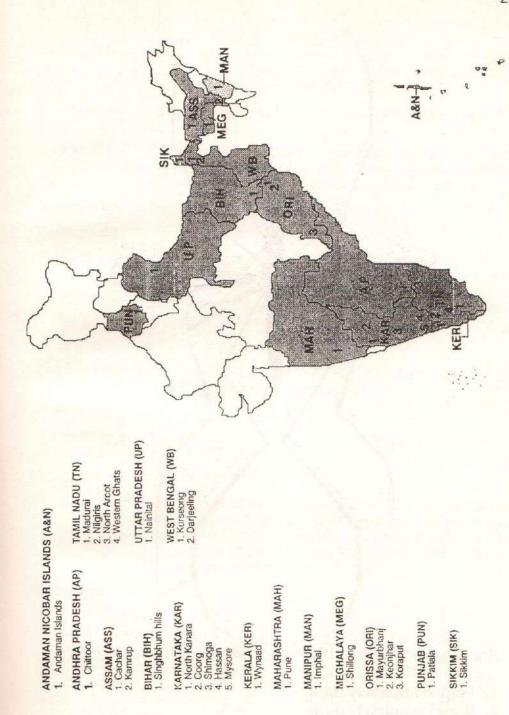
Relation to disease : NA

Reported distribution: Occurs in Bangladesh, China, India, Indonesia, West

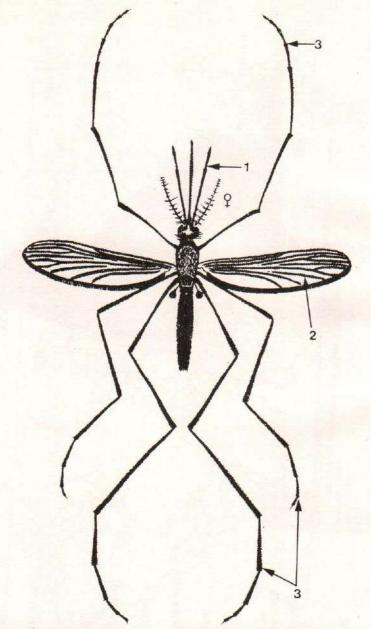
Iran, Malaysia, Myanmar, Nepal, the Philippines, Sri Lanka, and Thailand. In India, occurs in Andhra Pradesh, Assam, Bihar, Karnataka, Kerala, Maharashtra, Manipur, Meghalaya, Orissa, Punjab, Sikkim, Tamil Nadu, Uttar Pradesh, and West Bengal. Also occurs in Andaman

Islands.





Reported distribution of An. aitkenii in India.



An. bengalensis Puri 1930, main identification characters

- 1. Palpi completely dark
- 2. Wing completely dark
- 3. Legs completely dark

Anopheles beng	anlanair	
Alloulieles Della	Jaielisis	i

: An. bengalensis Puri, 1930. The Indian Journal of Name

Medical Research, 18: 953-956.

Derivative : Species named after Bengal, from where it was first

collected.

Type form : Paratype available in the National Institute of Communi-

cable Diseases, Delhi, India. Original type form deposited

in the British Museum of Natural History, London.

Sitting posture : Body parallel to resting surface.

Resting habit : A forest-loving species, rests outdoors.

Breeding ecology Breeds in seepages, springs or slow running streams

> with dead leaves and floating debris of vegetation. Also breeds in isolated pools along streams, and bamboo groves; otherwise the breeding sites are similar to those

of An. aitkenii.

Biting time Bites during day-time in jungles.

Feeding preference No record of feeding habit in India but in Thailand and

Malaysia, mostly feeds on man.

Flight range NA

: NA Susceptibility to

insecticides

Relation to disease : NA

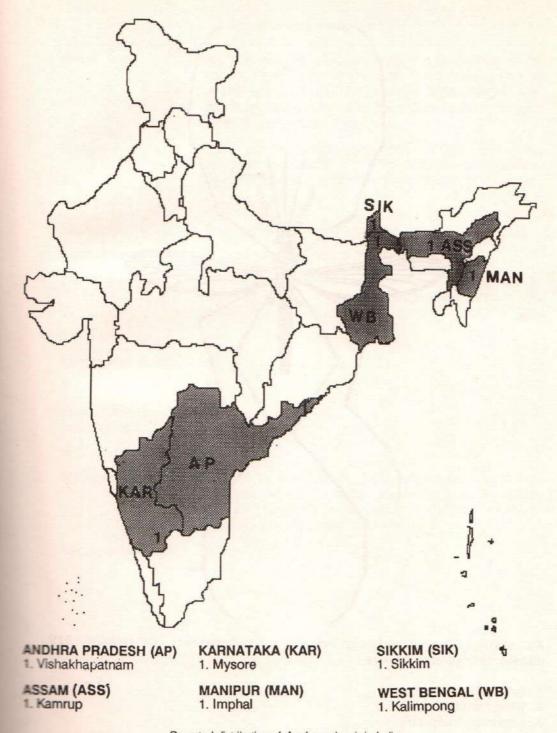
Distributed in Bangladesh, South China, India, Japan, Reported distribution:

Malaysia, Myanmar, the Philippines, Ryukyu Islands, Taiwan, Thailand, and Vietnam. In India, recorded from Andhra Pradesh, Assam, Karnataka, Manipur, Sikkim,

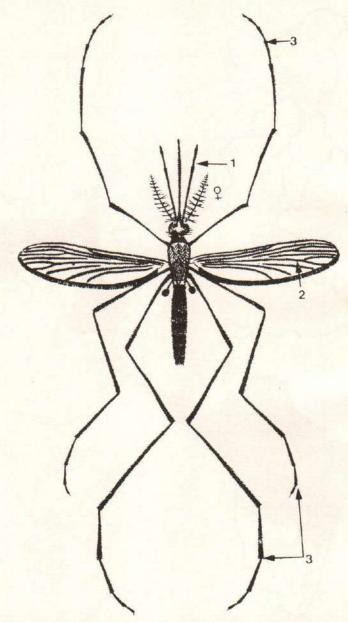
and West Bengal.



Reported distribution of An. bengalensis in the World.



Reported distribution of An. bengalensis in India.



An. insulaeflorum (Swellengrebel and Swellengrebel, de Graaf) 1919, main identification characters

- 1. Palpi completely dark
- 2. Wing completely dark
- 3. Legs completely dark

Anopheles insulaeflorum

Name : An. insulaeflorum (Swellengrebel and Swellengrebel-de

Graaf), 1919 (1920). Addenda to Description of Larvae of Netherlands Indian Anophelines. *Mededelingen van den Burgerlijken Geneeskundigen Dienst in Nederlansch-*

Indie, 9 (addendum): 1-3.

Derivative : Latin, insula, island; flos, floris, flowers. The word

insulaeflorum means "of the island of flowers"; chosen

by the author as it is found in some islands of Moiuccas.

Type form : Location not known.

Sitting posture : Body parallel to resting surface.

Resting habit : A forest species, rests outdoors.

Breeding ecology : Breeds in ground pools, rock pools, streams, margins,

arecanut garden trenches, seepage pools, etc. in the

shade.

Biting time : NA

Feeding preference: NA

Flight range : NA

Susceptibility to

insecticides

100 100

NA

Relation to disease: NA

Reported distribution: Occurs in Bangladesh, India, Indonesia, Cambodia,

Malaysia, Myanmar, Nepal, the Philippines, Sri Lanka, Taiwan, Thailand, and Vietnam. In India, found in Assam,

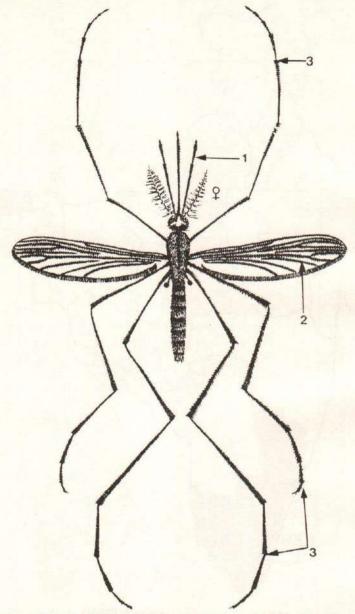
Karnataka, Kerala, and Tamil Nadu.



Reported distribution of An. insulaeflorum in the World.



Reported distribution of An. insulaeflorum in India.



An. pinjaurensis Barraud 1932, main identification characters

- 1. Palpi completely dark
- 2. Wing completely dark
- 3. Legs completely dark

Further identification is based on male genetilia characters

Anopheles pinjaurensis

Name : An. pinjaurensis Barraud, 1932. Records of the Malaria

Survey of India, 3: 353-355.

: Barraud named it pinjaurensis because it was caught at Derivative

Pinjaur Patiala State, near Kalka, (now in Haryana).

: Available at the National Institute of Communicable Type form

Diseases, Delhi, India.

Sitting posture : Body parallel to resting surface.

Breeding ecology : NA

Biting time : NA

: NA Feeding preference

: NA Flight range

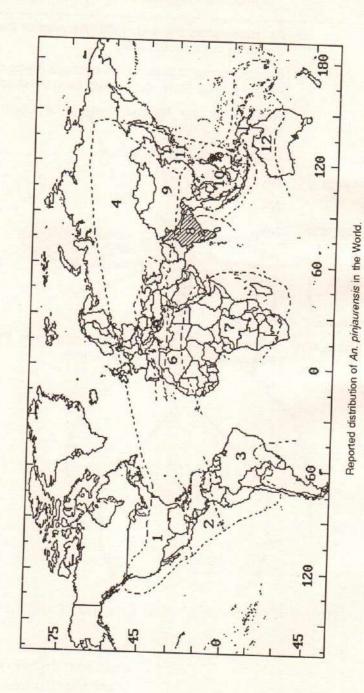
Susceptibility to

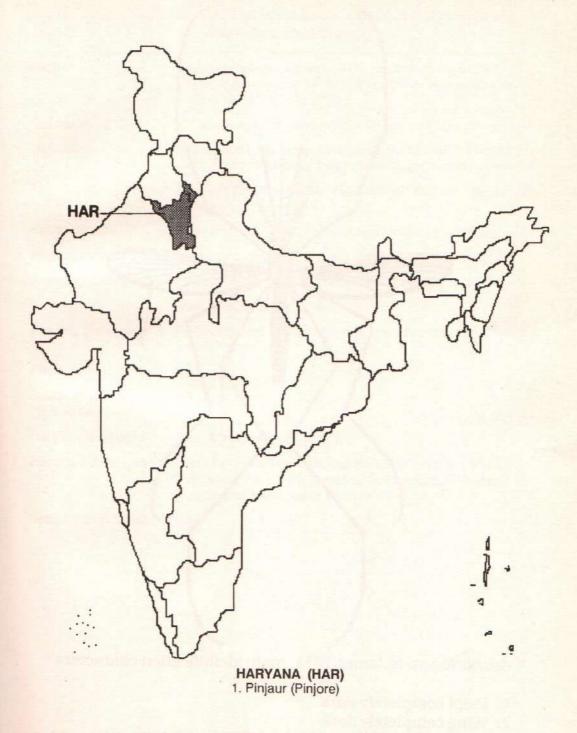
: NA insecticides

Relation to disease : NA

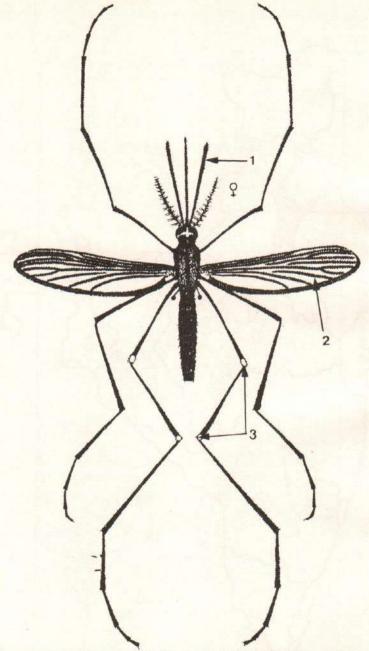
Reported distribution: A single male specimen collected from Pinjaur(now

Pinjore, near Kalka,), Haryana, India.





Reported distribution of An. pinjaurensis in India.



An. barianensis James 1911, main identification characters

- 1. Palpi completely dark
- 2. Wing completely dark
- 3. Pale band at the termination of femur and tibia of hind leg

Anopheles barianensis

Name : An. barianensis James, 1911. In: James and Liston, 1911, A Monograph of the Anopheline Mosquitoes of

India, 2nd edition, 128 pages.

Old name : An. plumbeus var. barianensis James.

Derivative : Named after the locality Barian in Murree hills, Punjab,

India (now in Pakistan) from where first collected.

Type form : Available at the British Museum of Natural History,

London

Sitting posture : Body and mouth parts at an angle to resting surface.

: Rests outdoors in tree holes, also indoors. Resting habit

Breeding ecology : Forest species, breeds mainly in tree holes.

Biting time : Soon after dusk.

Feeding preference: Preferably man and wild animals. Also cannibalistic at the

aquatic stage.

NA

Flight range : NA

Susceptibility to : NA

Relation to disease :

insecticides

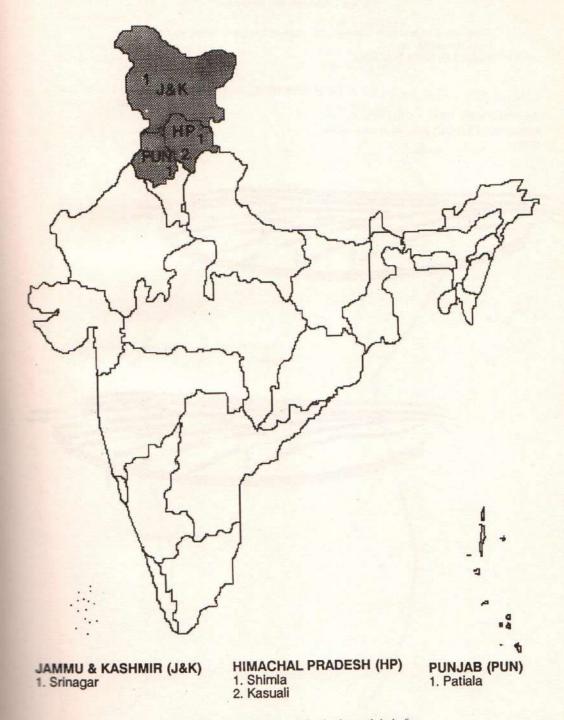
Reported distribution: Occurs in India, Pakistan, and the USSR. In India, known

only from the northern states i.e. Himachal Pradesh,

Punjab, and Jammu and Kashmir.



Reported distribution of An. barianensis in the World.



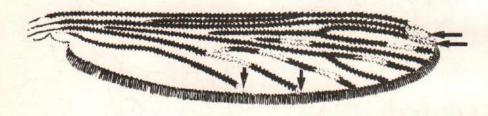
Reported distribution of An. barianensis in India.

2.3. An. lindesayi, An. nilgiricus

Wing with 3 or less pale areas on the costa, subcosta including vein1 (R1) Palpi completely dark
Femur with broad pale areas

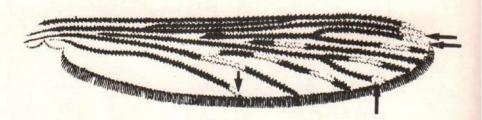
Tip of wing veins

Tip of wing veins 1(R1), 2.1(R2), 6(Anal), and sometimes of 5.2(Cu2) pale, tip of other veins dark



An. nilgirius

Tip of at least one other vein, usually 4.2(M2) pale



An. lindesayi

Other characters

Hind femur with a pale ring, nearly equal to the diameter of the femur

Base of hind femur with a pale ring of about equal width above and below



Remigum with yellow scales

An. lindesayi

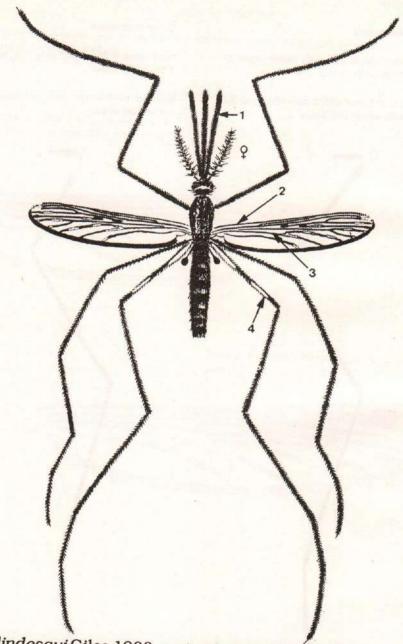
Other characters

Hind femur with a broad pale ring, occupying 1/5th of the femur

Base of hind femur more extensively pale below than above



Wing veins 3(R4+5) and 6(Anal) are mainly dark



An. lindesayi Giles 1900, main identification characters

- 1. Palpi completely dark
- 2. Inner costa completely dark
- 3. Stem of vein 5 (CU) completely dark
- 4. Hind femur with a pale band

Anopheles lindesayi

Name : An. lindesayi Giles, 1900. A Handbook of the Gnats or Mosquitoes Giving the Anatomy and Life History of the

Culicidae, 1st edition, 374.

Derivative : Giles named it after Captain Victor Edward Hugh

Lindesay who sent this specimen to him from Bakloh,

located in Punjab (now in Himachal Pradesh).

Type form : Available at the British Museum of Natural History,

London.

Species complex : Three species (An. gigas, An. wellingtonianus and

An. lindesayi).

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : A wild and high altitude species, recorded up to 2700 m.

Rests outdoors or in human dwellings and chicken coops.

Breeding ecology : Stream bed pools are the common breeding sites

although it also breeds in ground pools, mountain

streams, ricefields and borrowpits.

Biting time : Bites outdoors and indoors, biting commences during the

day in jungles and also at dusk near breeding sites.

Feeding preference: Preferably wild animals and man.

Flight range : NA

Susceptibility to : NA

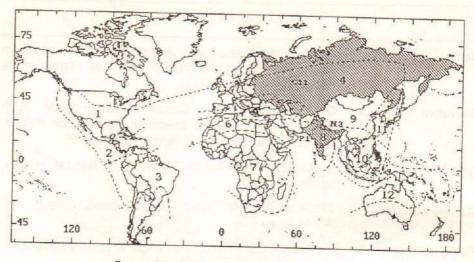
insecticides

Relation to disease: NA

Reported distribution: Occurs in India, Myanmar, Nepal, Pakistan, and USSR. In

India, recorded from Assam, Himachal Pradesh, Jammu and Kashmir, Manipur, Punjab, Sikkim, Tamil Nadu, Uttar

Pradesh, and West Bengal.



Reported distribution of An. lindesayi in the world.

Reported distribution of An. lindesayi in India.

ASSAM (ASS)

1. Kamrup

HIMACHAL PRADESH (HP)

- Kangra
 Bilaspur
- 3. Solan

JAMMU & KASHMIR (J & K)

- Srinagar
 Guimarg

MANIPUR (MAN) 1. Imphal

PUNJAB (PUN) 1. Ferozepur 2. Patiala

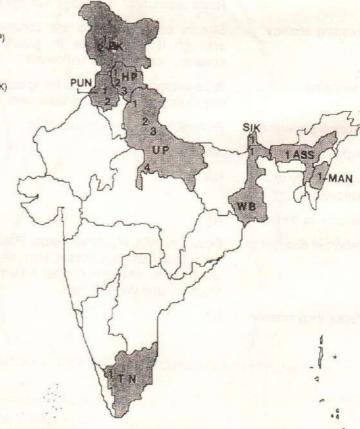
SIKKIM (SIK) 1. Sikkim

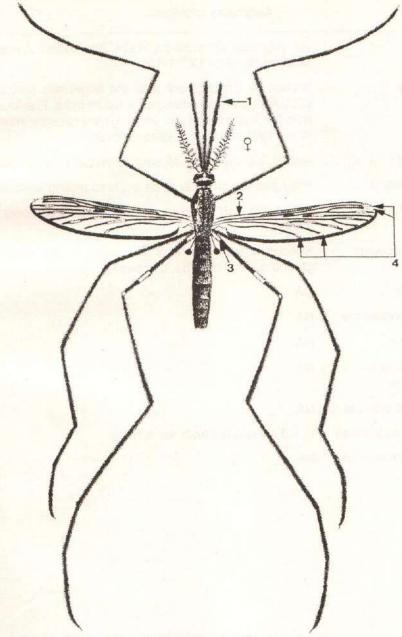
TAMIL NADU (TN) 1. Nilgiris

UTTAR PRADESH (UP)

- Almora
 Chamoli
- 3. Dehra Dun
- 4. Nainital

WEST BENGAL (WB) 1. Darjeeling





An. nilgiricus Christophers 1924, main identification characters

- Palpi completely dark
 Inner costa completely dark
 Hind femur with a pale band
 Tip of veins 1(R1), 2.1(R2), 6(Anal vein) and sometimes of 5.2 (CU2) pale.

Anopheles nilgiricus

Name An. nilgiricus Christophers, 1924. The Indian Journal of

Medical research 12: 11-13.

Derivative : Named by Christophers after the Nilgiri hills (2000 m),

2000 km South of Himalayas in South India. Previously it was a subspecies of An. lindesayi but recently Harrison

et al., 1991 raised it to species level.

Type form : Available at the British Museum of Natual History London.

Sitting posture Body and mouth parts at an angle to resting surface.

Resting habit A wild and high altitude species rests both outdoors and

indoors.

Breeding ecology Species breeds in stream bed pools. It also breeds in

ground pools, ricefields, borrowpits.

Biting time NA

Feeding preferance: NA

Flight range NA

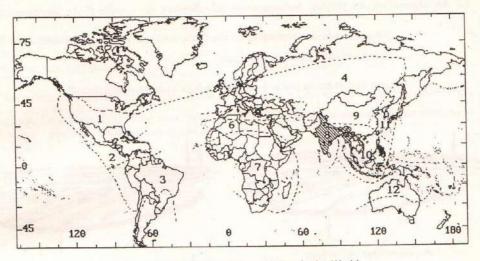
Susceptibility to insecticides

Relation to disease : NA

Reported distribution: In India, reported from Tamil Nadu.

: NA

Vector incrimination:



Reported distribution of An. nilgiricus in the World.



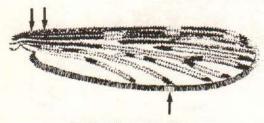
Reported distribution of An. nilgiricus in India.

2.4. An. nigerrimus, An. nitidus, An. argyropus, An. sinensis, An. crawfordi, An. peditaeniatus

Wing with 3 or less pale areas on the costa, subcosta including vein 1(R1) Four band palpi (tip of the palpi pale)

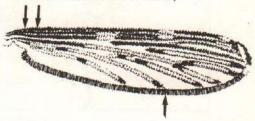
Basal half of costa (inner costa) and fringe spot on vein 5.2(Cu2)

Inner costa with pale scales and fringe spot



An. nigerrimus, An. nitidus

Inner costa without pale scale and fringe spot absent (exception in An. sinensis)



An. sinensis, An. peditaeniatus, An. argyropus, An. crawfordi

Size of basal dark mark on wing vein 5(Cu)

Small (0.2 x length of the stem of vein 5(Cu))



An. nitidus

Long (0.4 x length of the stem of vein 5(Cu))



An. nigerrimus

An. nitidus, An. nigerrimus

Other characters

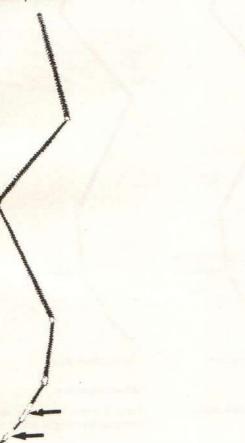
Humeral pale spot present

Hind tarsomeres with broad pale bands, fourth segment with basal pale band

Other characters

Humeral pale spot absent

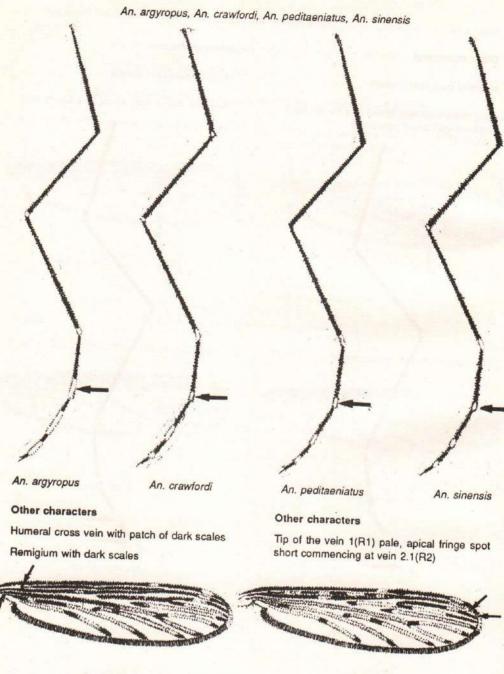
Hind tarsomeres with apical pale bands only



An. nitidus



An. nigerrimus



An. argyropus

An. crawfordi

An. peditaeniatus, An. sinensis

Other characters

Humeral cross vein with pale scales

Remigium with pale scales

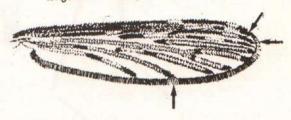


An. peditaeniatus

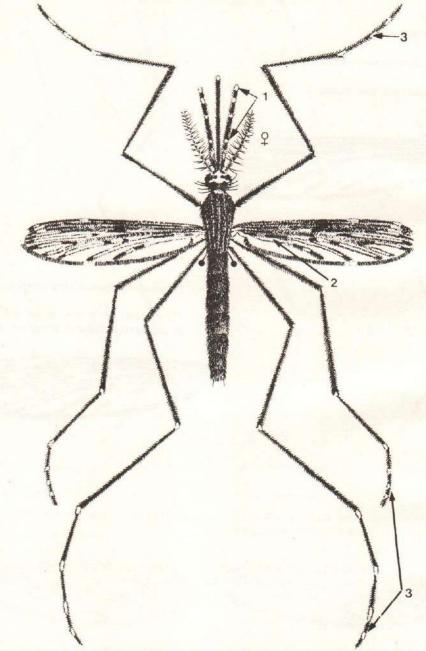
Other characters

Fringe spot present at wing vein 5.2(Cu2)

Tip of vein 1(R1) dark, apical fringe spot longer commencing at or above vein 1(R1)



An. sinensis



An. nigerrimus Giles 1900, main identification characters

- 1. Palpi with 4 small pale bands
- 2. Basal dark area of vein 5 (CU) is 1/2 of the stem
- 3. Tarsomeres with broad bands

Ano	phei	es	nigerrimus
, ,,,,	P1101	00	ingominuo

Name : An. nigerrimus Giles, 1900. A Handbook of the Gnats

or Mosquitoes giving the Anatomy and Life History of

Culicidae, 374 pages.

Old name : An. sinensis Wied (oriental).

Derivative : Latin, niger, black; -imus, superlative suffix. Giles gave

this name because of the intensely black colour of the

mosquito.

Type form : Available at the British Museum of Natural History,

London.

Reported morphological variations

: 3 in maxillary palpi and 1 in wing

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests mainly indoors in cattlesheds and human

dwellings, also outdoors in good numbers.

Breeding ecology : Breeds in various types of still waters with aquatic

vegetation and is particularly associated with ricefields (with well-grown plants). Also breeds in lakes, grassy pools, tanks, moats, swamps, borrowpits, drains, edges of slow-moving waters, shady ponds, usually with good

aquatic vegetation.

Biting time : Bites outdoors and indoors and usually at dusk.

Sometimes bites in day time during full sunshine.

Feeding preference : Generally recognized as a zoophilic but also feeds readily

on man.

Flight range : Up to 1 km.

Susceptibility to insecticides

Incipient resistance to DDT in Orissa (India).

Relation to disease: A well-known vector of malaria in Indonesia and

Malaysia. Also transmits filariasis (*B. malayi*) in India, Malaysia, Thailand, and Sri Lanka. In India, incriminated for *B. malayi* from West Bengal and for *W. bancrofti* from

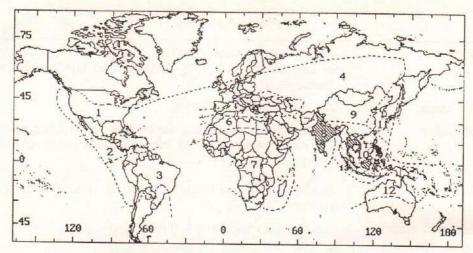
coastal Orissa.

Reported distribution: Distributed throughout the oriental region. In India,

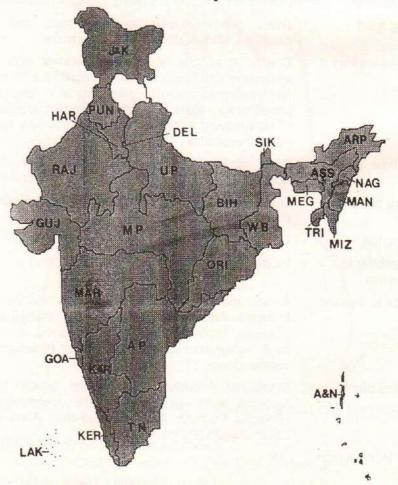
recorded from all parts of the mainland except Himachal Pradesh. Also recorded from Andaman and

Lakshadweep Islands.

Vector incrimination : NA

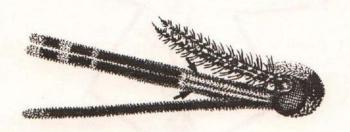


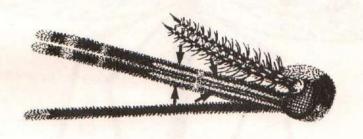
Reported distribution of An. nigerrimus in the World.

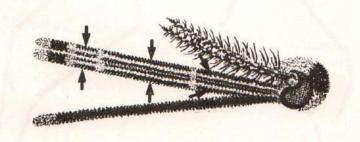


Reported distribution of An. nigerrimus in India.

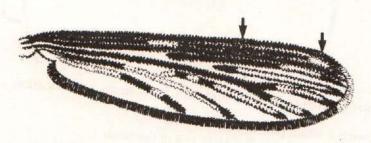
Variations in palpi

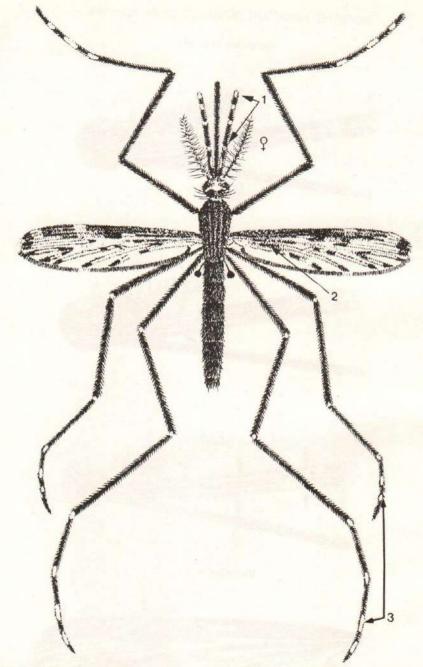






Variation in wing





An. nitidus Harrison, Scanlon and Reid 1973, main identification characters

Palpi with 4 small pale bands
 Basal dark area of vein 5 (CU) is 1/4 of the stem

3. Tarsomeres with broad bands

Anopheles nitidus

Name : An. nitidus Harrison, Scanlon and Reid, 1973. Mosquito Systematics, 5: 263-268.

Derivative : Latin, *nitidus*, shining, bright, glistening. This is a new name for *indiensis* of Reid. Renamed *nitidus* owing to the presence of elegant, neat, bright, sharp wing pattern.

Type form : Available at the British Museum of Natural History, London.

London.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests both indoors and outdoors.

Breeding ecology : Breeds in swamps, marshes in jungles, seepages, large

mine pits, rock pools, ditches, ricefields, and elephant

footprints in the forest.

Biting time : Before midnight.

Feeding preference: Generally a cattle feeder, but sometimes also bites man.

Flight range : NA

Susceptibility to : NA

insecticides

Relation to disease : NA

Reported distribution: Occurs in India, Indonesia, Cambodia, Malaysia,

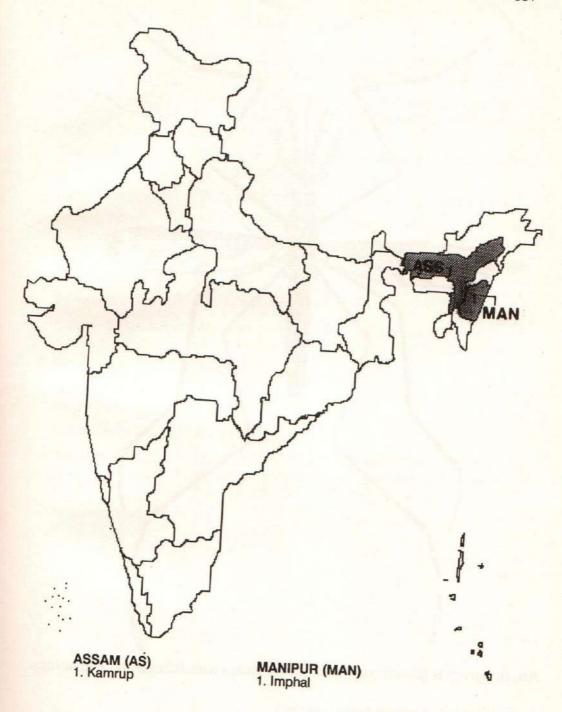
Thailand, and North and South Vietnam. In India,

recorded from Assam, and Manipur only.

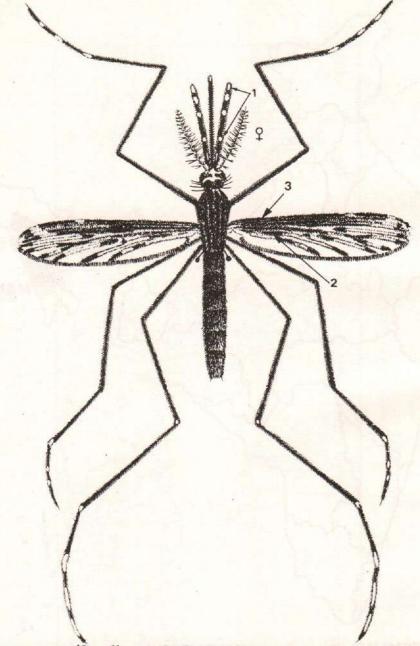
Vector incrimination : NA



Reported distribution of An. nitidus in the World.



Reported distribution of An. nitidus in India.



An. argyropus (Swellengrebel) 1914, main identification characters

- 1. Palpi with 4 small pale bands
- 2. Basal dark area of vein 5 (CU) is 3/4 of the stem
- 3. Inner costa completely dark

Anopheles argyropus

Name : An. argyropus (Swellengrebel), 1914. Geneeskundig

Tijdschrift Voor Nederlandsch, Indië, 54: 334-337.

Derivative : Greek, argyros, silver; pous, foot. Swellengrebel named

it argyropus because of the presence of white markings

on hind tarsomeres 3, 4 and 5.

Type form : Plesiotypes available at the British Museum of Natural

History, London.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Rests in houses and cattlesheds. Outdoor resting also

recorded.

Breeding ecology : Breeds in ricefields and large deep swamps with

vegetation.

Biting time : Before midnight.

Feeding preference: Attracted more towards cattle than man. Specimens can

also be collected in good numbers with the light trap.

Flight range : NA

Susceptibility to : 1

insecticides

: NA

Relation to disease : NA

Reported distribution: Distributed in India, Indonesia, Cambodia, Malaysia,

Myanmar, Malaya (Java), Thailand, and Vietnam. In

India, recorded from Assam region only.

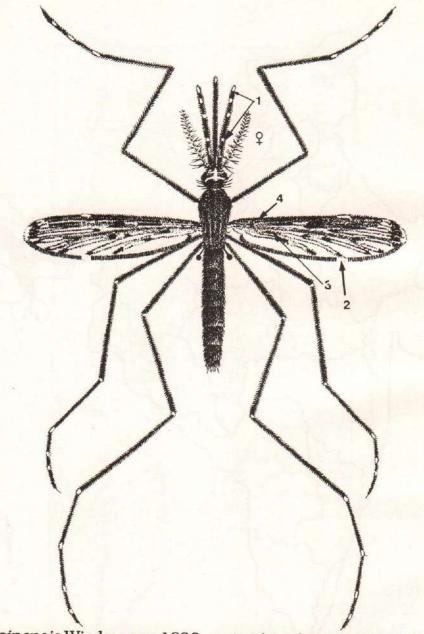
Vector incrimination: NA



Reported distribution of An. argyropus in the World.



Reported distribution of An. argyropus in India.



An. sinensis Wiedemann 1828, main identification characters

- 1. Palpi with 4 small pale bands
- 2. Fringe spot on vein 5.2 (CU2)
- 3. Basal dark area of vein 5 (CU) is 1/6 of the stem
- 4. Inner costa completely dark

Anopheles sinensis

Name

: An. sinensis Wiedemann, 1828. Aussereuropäische Zweiflugelige Insekten, vol. 1, 608 pages

Derivative

The species was collected in China by Wiedemann's friend Dr. Trentepohl from various points in the Far East including Canton and Macao. The Latin Sinae (Greek Sinai), the Chinese, appears to be the base for Wiedemann's locative adjective, sinensis.

Type form

: Available at the Natural History Museum, Vienna, Austria.

Sitting posture

Body and mouth parts at an angle to resting surface.

Resting habit

Regarded as exophilic but also rests in human dwellings

and cattlesheds.

Breeding ecology

Ricefields are the most important breeding sites. Other breeding places are marshes, streams, seepages, and various types of ground pools and ponds with water hyacirith.

Biting time

: Before midnight.

Feeding preference: Highly zoophilic but bites man also.

Flight range

: NA

Susceptibility to insecticides

· NA

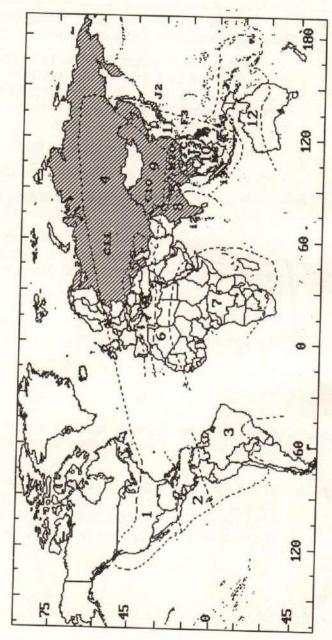
Relation to disease :

Not a vector of malaria in India but a main vector of malaria in China, Indochina, Indonesia, Myanmar, and

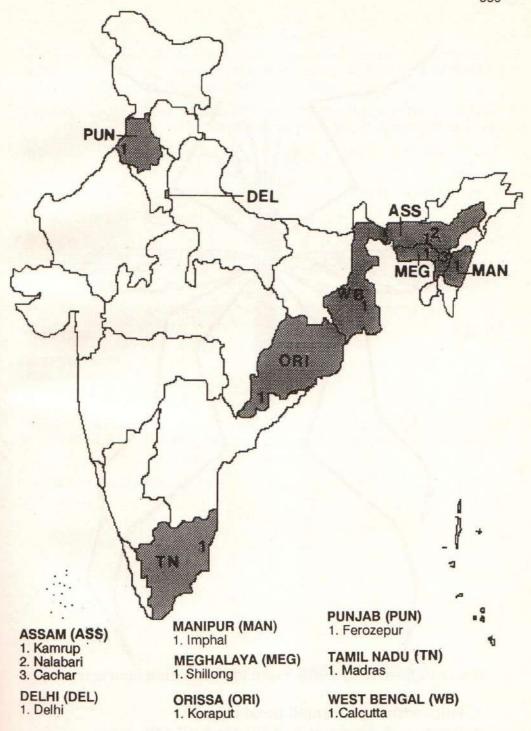
Taiwan.

Reported distribution: Found in China, Formosa, India, Indonesia, Japan, Malaya, Myanmar, Siberia, Sumatra, and Thailand. In India, recorded from Assam, Delhi, Manipur, Meghalaya Mizoram, Orissa, Punjab, and Tamil Nadu.

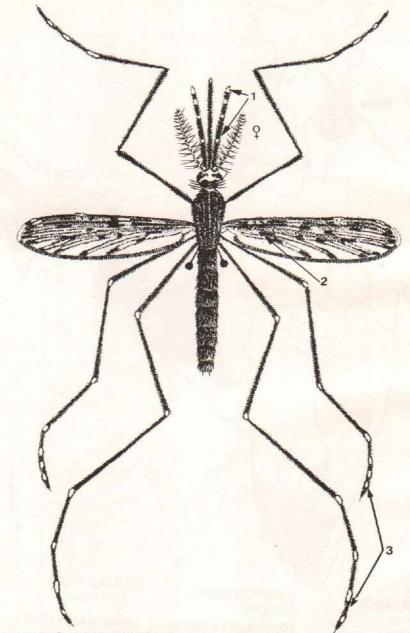
Vector incrimination: NA



Reported distribution of An. sinensis in the World.



Reported distribution of An. sinensis in India.



An. crawfordi Reid 1953, main identification characters

- 1. Palpi with 4 pale small bands
- 2. Basal dark area of vein 5 (CU) is 1/6 of the stem
- 3. Tarsomeres with broad bands

Anopheles crawfordi

: An. crawfordi Reid, 1953. Bulletin of Entomological Name

Research, 44: 5-76.

: Named by Reid after Dr. Robert Crawford who published Derivative an excellent work on the pupae of Malayan anophelines.

: Available at the British Museum of Natural History, Type form

London.

: Body and mouth parts at an angle to resting surface. Sitting posture

: Rests outdoors in wet forests. Resting habit

: Particularly breeds in marshes, ground pools and Breeding ecology

swamps.

: Before midnight. Biting time

More attracted to man than monkeys and cattle. Feeding preference

: NA Flight range

: NA Susceptibility to insecticides

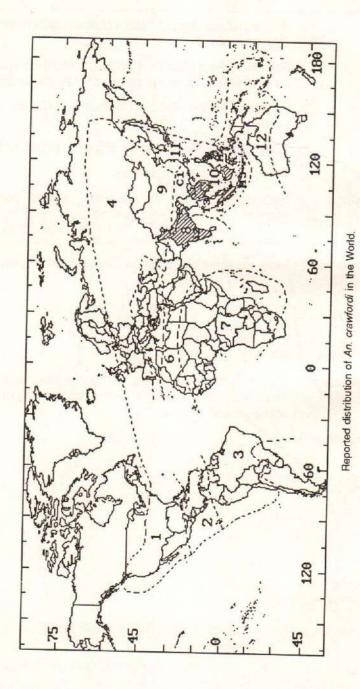
Relation to disease : NA

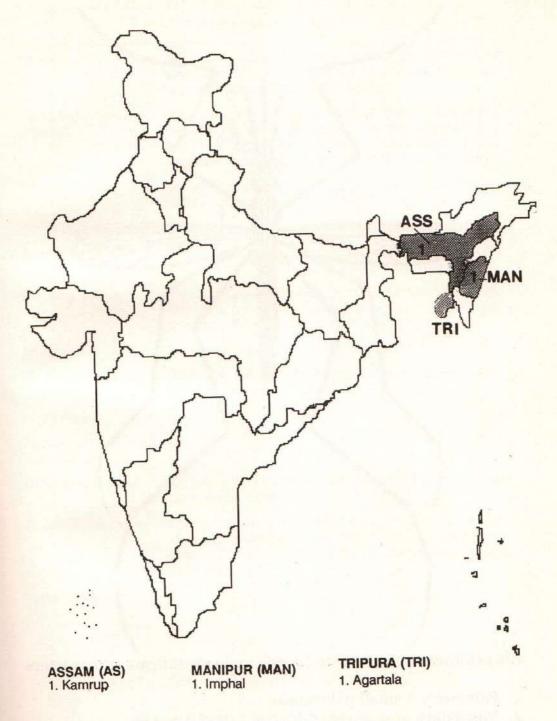
Reported distribution: Occurs in India, Indonesia, Cambodia, Malaya, Malaysia,

Thailand, and Vietnam. In India, recorded from Assam,

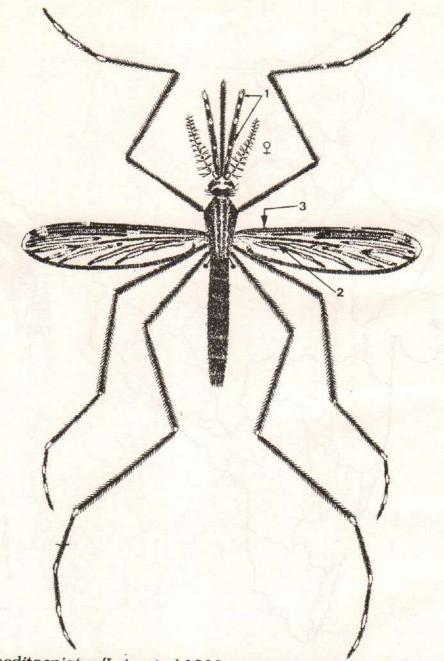
Manipur, and Tripura.

Vector incrimination : NA





Reported distribution of An. crawfordi in India.



An. peditaeniatus (Leicester) 1908, main identification characters

1. Palpi with 4 small pale bands

2. Basal dark area of vein 5 (CU) is 1/3 of the stem

3. Inner costa completely dark

Anopheles peditaeniatus

: An. peditaeniatus (Leicester), 1908. The Culicidae of Malaya. Studies of the Institute for Medical Research. Name Federated Malay States 3 (3): 18-261.

: Latin pes, pedis, foot; taenia, band, ribbon; atus, suffix, meaning 'having the nature of'. Leicester gave the name peditaeniatus because of the presence of creamy yellow bands on the metatarsus of fore-, mid-, and hind legs.

Syntypes available at the British Museum of Natural Type form History, London.

: Body and mouth parts at an angle to resting surface. Sitting posture

: Prefers cattlesheds to human dwellings. Adults collected at an altitude of 570 m in Sri Lanka and 540 m in Thailand. Resting habit

: Breeds mainly in ricefields, but also in marshes, ditches, seepages, ponds, swamps, temporary pools, margins of Breeding ecology streams, animal footprints, shallow wells, etc.

: Bites outdoors and indoors before midnight. Biting time

Cattle are the overwhelmingly preferred host, but also Feeding preference

feeds on humans.

: NA Flight range : NA Susceptibility to

insecticides

Derivative

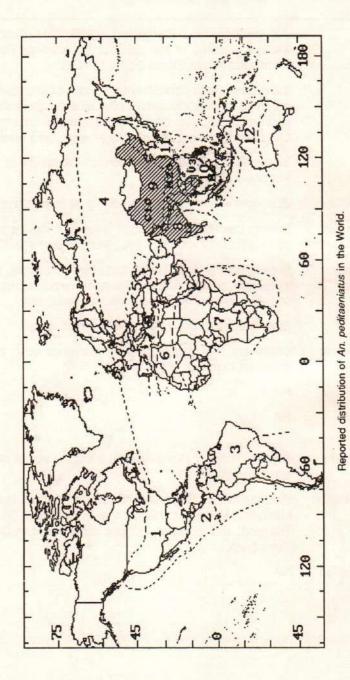
A good vector of B. malayi and a natural vector of animal Relation to disease :

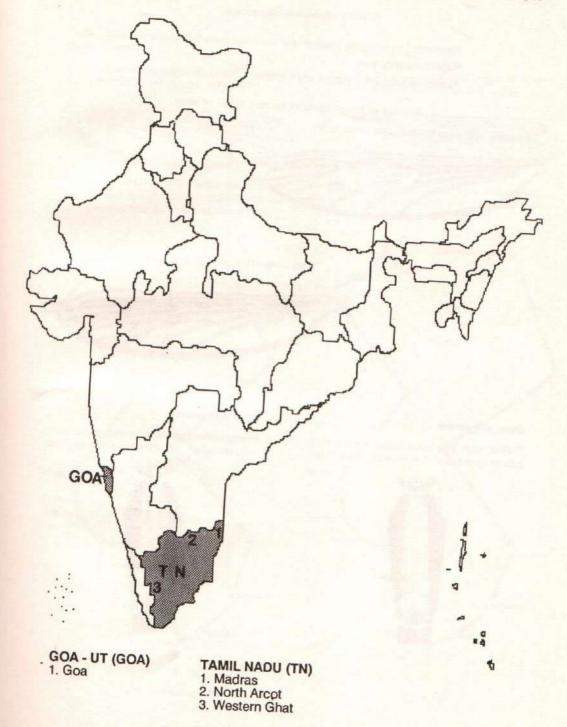
filaria genus Setaria.

Distributed in Bangladesh, China, India, Indonesia, Malaysia, Myanmar, Nepal, the Philippines, Sri Lanka, Reported distribution: Thailand, and Vietnam. In India, recorded from Goa, and

Tamil Nadu.

Vector incrimination: NA





Reported distribution of An. peditaeniatus in India.

2.5. An. umbrosus, An. roperi

Wing with 3 or less pale areas on the costa, subcosta including vein 1(R1)
Palpi completely dark

Abdominal segment 7 without any tuft of dark scales on venter side

Number of fringe spots at apical portion of wing

Two veins 1(R1) and 3(R4+5) with fringe spot



An. umbrosus

Three veins 1(R1), 2.1(R2) and 3(R4+5) with fringe spot



An. roperi

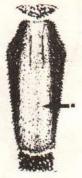
Other characters

Thorax with propleural setae (mesonotum pale in centre and dark towards lateral side)



Thorax without propleural setae (mesonotum with mottled pattern of light and dark patches)





Other characters

Extreme base of costa and bases of veins 1(R1) and 5(Cu) without any pale scales



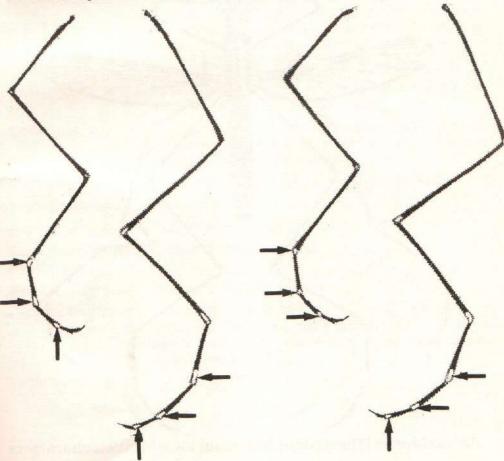
Foreleg tarsomeres with narrow apical pale bands; Hind tarsomeres with pale bands up to 3 times as long as wide and above joints

Other characters

Extreme base of costa and bases of veins 1(R1) and 5(Cu) with some pale scales

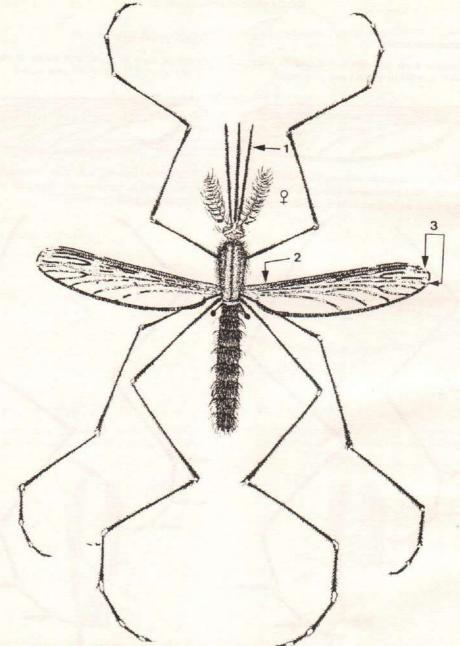


Foreleg tarsomeres with very minute pale bands; Hind tarsomeres with third pale band lying across joints



An. umbrosus

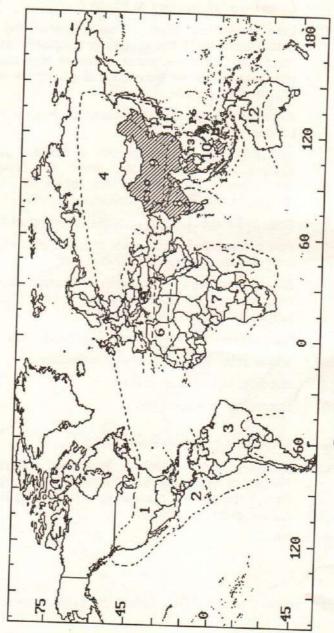
An. roperi



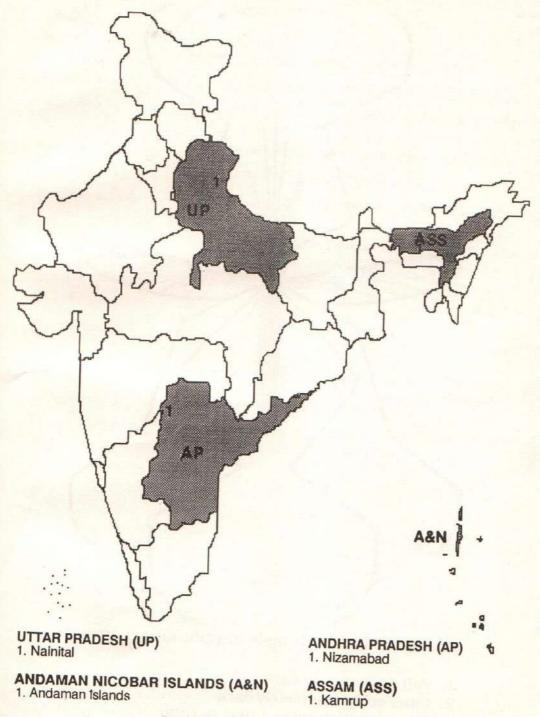
An. umbrosus (Theobald) 1903, main identification characters

- 1. Palpi completely dark
- 2. Inner costa completely dark
- 3. Fringe spot on veins 1 (R1) and 3(R4+5)

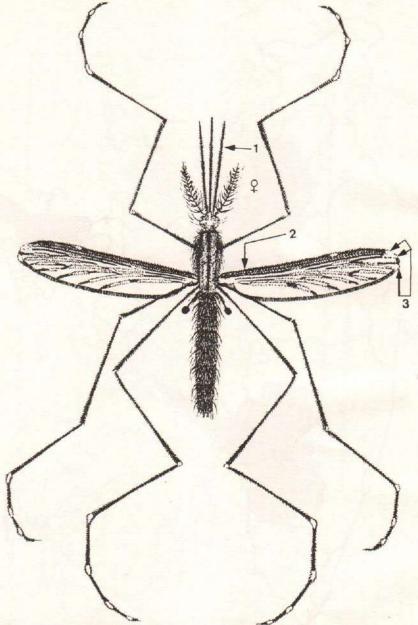
Anopheles umbrosus			
Name	:	An. umbrosus (Theobald), 1903. A Monograph of the Culicidae or Mosquitoes, 3: 87-88.	
Derivative	:	Latin, <i>umbrasus</i> , shady. Theobald gave the name <i>umbrosus</i> because the species is completely dark, i.e. head, palpi, antennae, thorax, abdomen, legs, costa and remainder of the first and second longitudinal wing veins are completely black.	
Type form	:	Available at the British Museum of Natural History, London.	
Species complex	:	Twelve species (An. hunteri, An. sinilissimus, An. bre- vipalpis, An. separatus, An. letifer, An. brevirostris, An. umbrosus, An. baezai, An. roperi, An. collessi, An. whartoni, and An. albotaeniatus).	
Sitting posture	1	Body and mouth parts at an angle to resting surface.	
Resting habit	:	Forest species generally rests outdoors in forest areas under shrubs; also rests indoors in huts.	
Breeding ecology		Breeds in stagnant shallow waters in swamp forest areas. Also breeds in shaded pools and running streams of jungles in Thailand and Malaysia.	
Biting time	:	Bites actively throughout the day in forest areas.	
Feeding preference	:	Mainly zoophilic but also bites man readily.	
Flight range	:	Reported dispersal up to 2 km.	
Susceptibility to insecticides		Susceptible to insecticides.	
Relation to disease	:	A vector of malaria in Indonesia.	
Reported distribution	:	Occurs in India, Indochina, Indonesia, Malaysia, the Philippines, Singapore, and Thailand. In India, recorded from Andaman and Nicobar Islands, Andhra Pradesh, Assain, and Uttar Pradesh.	
Vector incrimination	:	NA	



Reported distribution of An. umbrosus in the World.



Reported distribution of An. umbrosus in India.



An. roperi Reid 1950, main identification characters

- 1. Palpi completely dark
- 2. Inner costa completely dark
- 3. Fringe spot on veins 1 (R1), 2.1(R2)

Anopheles roperi

Name : An. roperi Reid, 1950. Transactions of the Royal Entomological Society of London, 101: 281-318

Derivative : Reid named the species after Richard Snelling Roper.

Type form : Available at the British Museum of Natural History, London.

Editabil

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : An outdoor resting species (tree holes).

Breeding ecology : Breeds in shaded pools, swamps, ditches and flooded

pools.

Biting time : In forests, bites outdoors throughout night and day with

a peak in late evenings.

Feeding preference : Bites both monkeys and men.

Flight range : NA

Susceptibility to : NA insecticides

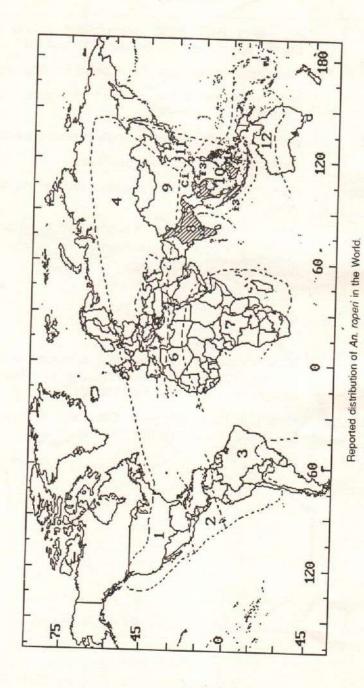
Relation to disease : No information is available about its relation to disease

in India. However, natural infections of Plasmodium

parasite have been found in Malaysia.

Reported distribution: Occurs in India, Indonesia, Cambodia, Malaysia, and

Thailand. In India, collected from Assam only.





Reported distribution of An. roperi in India.

2.6. An. ahomi, An. barbirostris, An. barbumbrosus

Wing with 3 or less pale areas on the costa, subcosta including vein 1(R1) Palpi completely dark

Abdominal segment 7 with a tuft of dark scales on venter side

Size of apical tringe spot (fringe spot of wing vein 3(R4+5)); Scales on sternite 2-6 of abdominal segments

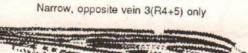
Wide at least from 3(R4+5) to 4.1(M1)



Scales absent on sternite



An. ahomi, An. barbumbrosus





Scales on sternite



An. barbirostris

An. ahomi, An. barbumbrosus

Fringe spot on vein 2.1(R2)

Absent



An. ahomi

Present



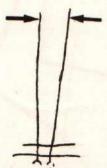
An. barbumbrosus

Both species can be confirmed by the inner clypeal hair on the head of larvae

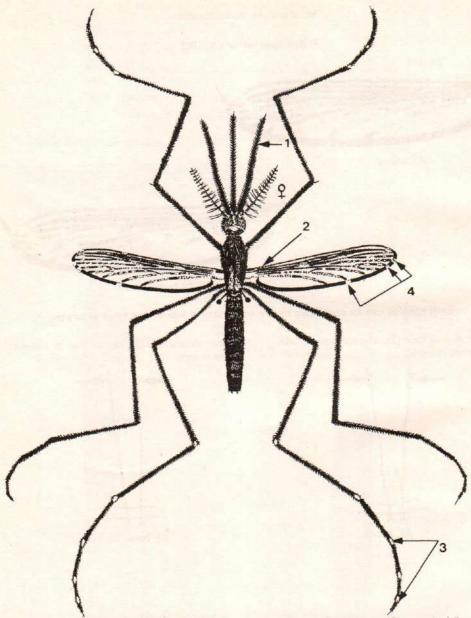
Inner clypeal hair (2c; clypeal hair 2) with branches (12-36)

An, ahomi

Inner clypeal hair (2c; clypeal hair 2) without any branch



An. barbumbrosus



An. ahomi Chowdhury 1929, main identification characters

- 1. Palpi completely dark
- 2. Inner costa with pale scales
- 3. Hind leg tarsomeres with bands
- 4. Fringe spot on vein 3 (R4+5), 4.1 (M1) and 5.2 (CU2)

Further identification is based on larval characters

Anopheles ahomi

Name : An. ahomi Chowdhury, 1929. The Indian Journal of Medical Research, 16: 986-988.

Derivative : Chowdhury named the species ahomi after Ahom or Aham, a powerful tribe of Shan descent, invaded the

Assam valley in the 13th century.

Type form : Location not known.

: NA

Species complex : A member of barbirostris subgroup of barbirostris

complex.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : NA

Breeding ecology : Breeds in pools and streams in tea gardens.

Biting time : NA

Feeding preference: NA

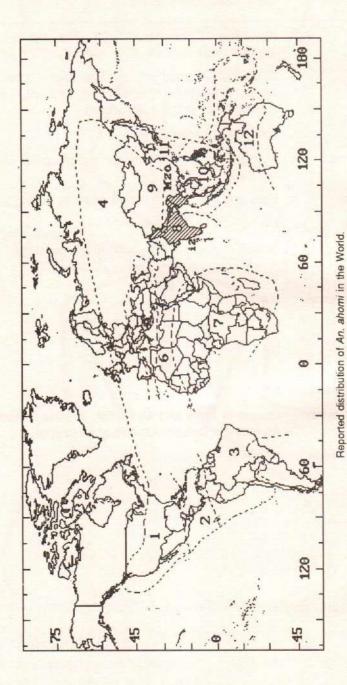
Flight range NA

Susceptibility to insecticides

Relation to disease : NA

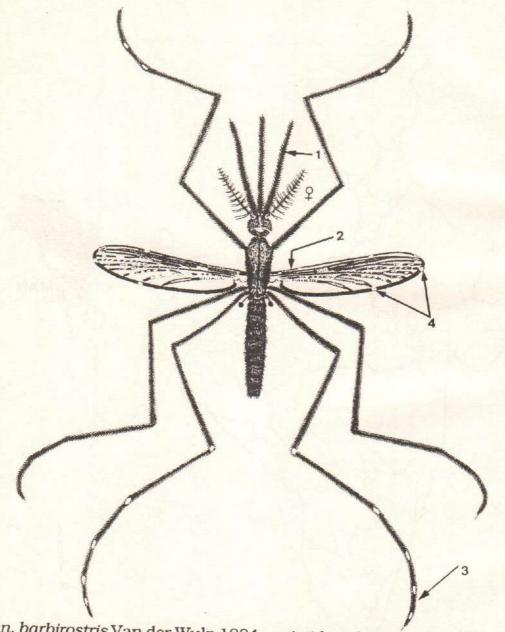
Reported distribution: Occurs in India and Myanmar. In India, recorded from

Arunachal Pradesh, Assam, and Manipur.





Reported distribution of An. ahomi in India.



An. barbirostris Van der Wulp 1884, main identification characters

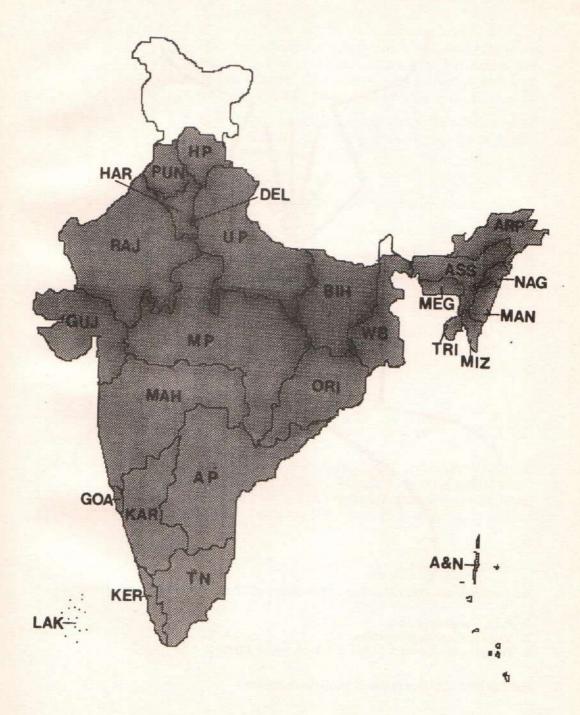
- 1. Palpi completely dark
- 2. Inner costa with pale scales
- 3. Hind leg tarsomeres with broad bands
- 4. Fringe spot on vein 3 (R4+5) and 5.2 (CU2)

Anopheles barbirostris		
Name	:	An. barbirostris Van der Wulp, 1884. Notes from the Leyden Museum, 6: 248-256.
Derivative	:	Latin, barba, beard; rostrum, beak, bill, snout; -is, adjectival ending. Van der Wulp named the species barbirostris, because the proboscis is quite long and hairy and the rostrum and palpi are longer than the head and thorax taken together, both clothed with brown, scaly hairs; the joints of palpi are hardly distinguishable in this species.
Type form	:	Available at the State Museum of Natural History, Leyden, Netherlands.
Species complex		Eleven species (An. barbirontris, An. campestris, An. donaldi, An. tranciscoi, An. hodgkini, An. pollicaris, An. ahomi, An. vanus, An. barbumbrosus, An. manalangi, and An. reidi).
Sitting posture	:	Body and mouth parts at an angle to resting surface.
Resting habit	:	Rests indoors in cattlesheds and human dwellings, also rests outdoors in jungles.
Breeding ecology	:	Breeds in ponds, small ground pools with vegetation or organic matter, borrowpits, ricefields, slow-running streams, shallow wells, tanks, and even salt sumps.
Biting time	:	Man and cattle before midnight. In jungles, bites during day-time also.
Feeding preference	:	Zoophilic but in the absence of animals bites man readily
Flight range	:	Up to 1 km.
Susceptibility to insecticides	:	Resistant to DDT.
Relation to disease	:	A secondary vector of malaria and a recognized vector of human filaria in Indonesia. In India, found positive for filariasis (B. malayı) infection.
Reported distribution	1:	Occurs in Bangladesh, North Borneo, China, India

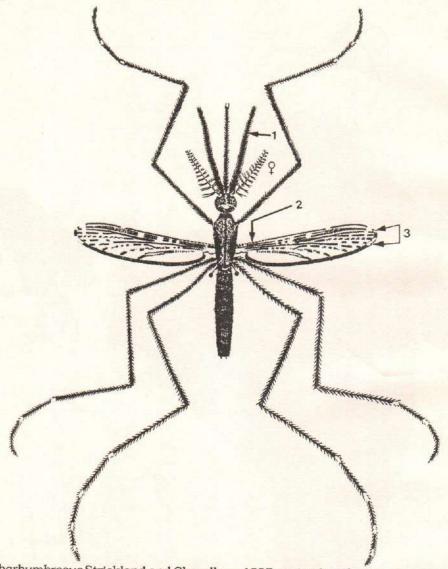
Indonesia, Cambodia, Kalimantan, Laos, Malaysia, Moluccas, Myanmar, Nepal, Sri Lanka, Thailand, Timar and Vietnam. In India, recorded throughout the country.



Reported distribution of An. barbirostris in the World.



Reported distribution of An. barbirostris in India.



An. barbumbrosus Strickland and Chowdhury 1927, main identification characters

- Palpi completely dark
- 2. Inner costa with pale scales
- 3. Fringe spot on vein 2.1 (R2), 3 (R4+5), and 4.1 (M1)

Further identification is based on larval characters

Anopheles barbumbrosus

Name : An. barbumbrosus Strickland and Chowdhury, 1927. An

Illustrated Key to the Identification of the Anopheline Larvae of India, Ceylon and Malaya, West of Wallace's Line with Practical Notes on their collection, pp. 67.

Derivative : Latin, barba, beard; umbrosus, shady, dark. Presumably

the name refers to the dark scales on female proboscis

as seen in barbirostris.

Type form : Location not known.

Species complex : A member of vanus, subgroup of barbirostris complex.

Sitting posture : Body and mouth parts at an angle to resting surface.

Breeding ecology : Breeds on ground and in rocky pools, stream margins,

ponds, puddles, swamps, margins, shallow wells, springs, ricefields, animal footprints, tree holes, water

pots, etc.

Biting time : Before midnight.

Feeding preference: Man and wild animals.

Flight range : NA

Susceptibility to : NA

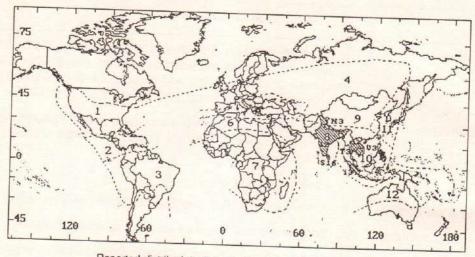
insecticides

Relation to disease: NA

Reported distribution: Distributed in Cambodia, India, Indonesia, Malaysia,

Nepal, South Vietnam, Sri Lanka, and Thailand. In India, reported from Andhra Pradesh, Assam, Bihar, Karnataka, Madhya Pradesh, Maharashtra, Tamil Nadu and West

Bengal.



Reported distribution of An. barbumbrosus in the World.



Reported distribution of An. barbumbrosus in India.

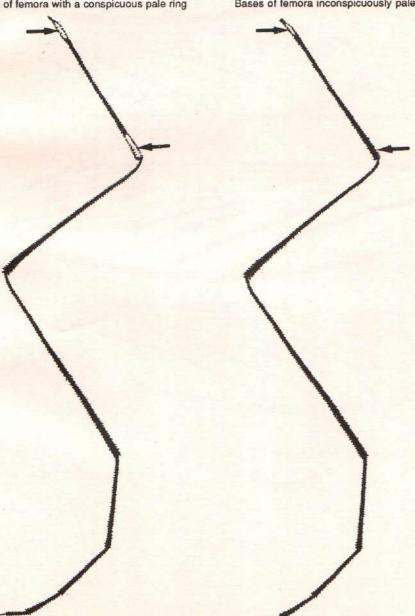
2.7 An. gigas, An. baileyi, An. gigas var. simlensis

Wing with 3 or less pale areas on the costa, subcosta including vein 1(R1)
Palpi dark but with small pale bands at the joints
Hind femur without tuft of dark and pale scales towards apex

Towards apex a large pale spot on dorsal side on mid-leg femur; pale spots on outer half of vein 6(Anal); a pale ring twice in diameter of femur on bases of femora

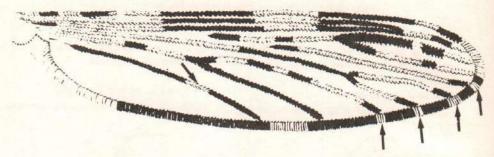
Pale spot on mid-leg
Bases of femora with a conspicuous pale ring

Pale spot on mid-leg absent
Bases of femora inconspicuously pale.



An. gigas, An. baileyi, An. gigas var. simlensis

Pale spot on outer half of vein 6(Anal)

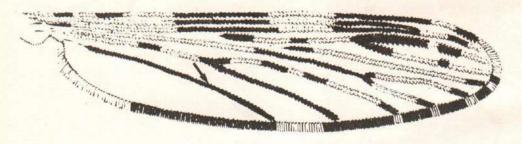


An. gigas

Pale spot on outer half of vein 6(Anal) absent

Other characters

Fringe spot on wing veins 3(R4+5), 4.1(M1), 4.2(M2), and 5.1(Cu1)



An. baileyi, An. gigas var. simlensis

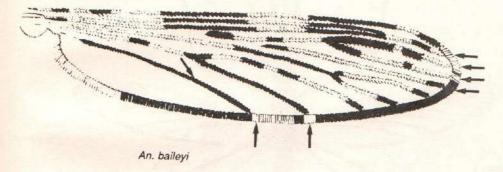
An. baileyi, An. gigas var. simlensis

Fringe spot on vein 3(R4+5)

An. baileyi

Other characters

Fringe spot on wing veins 1(R1), 2.1(R2), 2.2(R3), 5.2(Cu2) and 6(Anal)

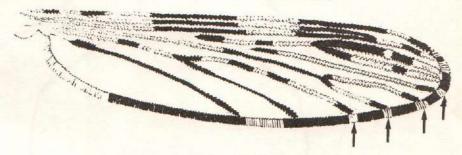


Present

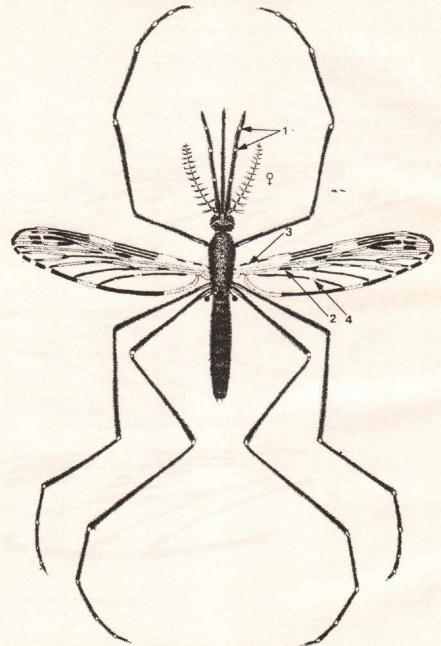
An. gigas var. simlensis

Other characters

Fringe spot on wing veins 3(R4+5), 4.1(M1), 4.2(M2) and 5.1(Cu1) but sometimes on only veins 3(R4+5) and 4.1(M1)



An. gigas var. simlensis



An. gigas Giles 1901, main identification characters

- 1. Palpi completely dark but small bands at joints
- 2. Basal dark area of vein 5 (CU) 1/6 of the stem
- 3. Inner costa interrupted
- 4. Vein 6 (Anal vein) with a pale spot on outer half

Anopheles gigas

Name : An. gigas Giles, 1901. Entomologist's Monthly Magazine, 37: 196–198.

: Latin, gigas, giant, the sons of Earth and Tartarus (Greek.

gigas). Giles found it to be the largest Indian form and

named it accordingly.

Type form : Available at the British Museum of Natural History,

London.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : Generally an outdoor rester, sometimes also rests in

cattlesheds and human dwellings. The type form is apparently attracted by light. All the forms of *An. gigas* are normally recorded from high altitudes, i.e., more than

2000 m.

Breeding ecology : Breeding places are freshwater springs and ponds

with vegetation on the edges, springs and seepages, ponds and small pools along shallow hill streams. The variety *simlensis* breeds in several habitats such as ground pools at the side of streams, ricefields, flooded wheat-fields, shallow swamps, seepage pools, snowwater pools, spring-bed pools, forest pools, rock holes, rainwater collections, and coaltar barrels with clean or

turbid water.

Biting time : Before midnight.

Feeding preference: Wild animals and sometimes man.

Flight range : Reported up to 30.5 km.

Susceptibility to : NA

insecticides

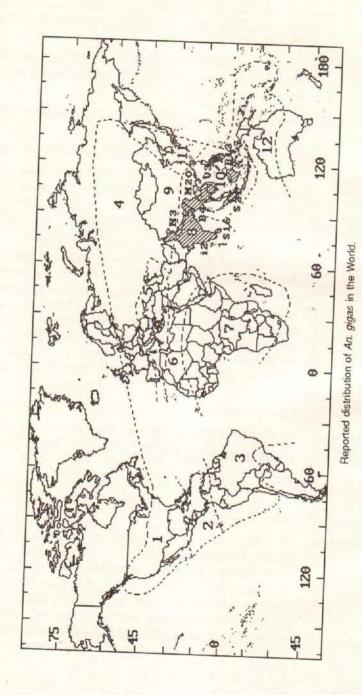
Derivative

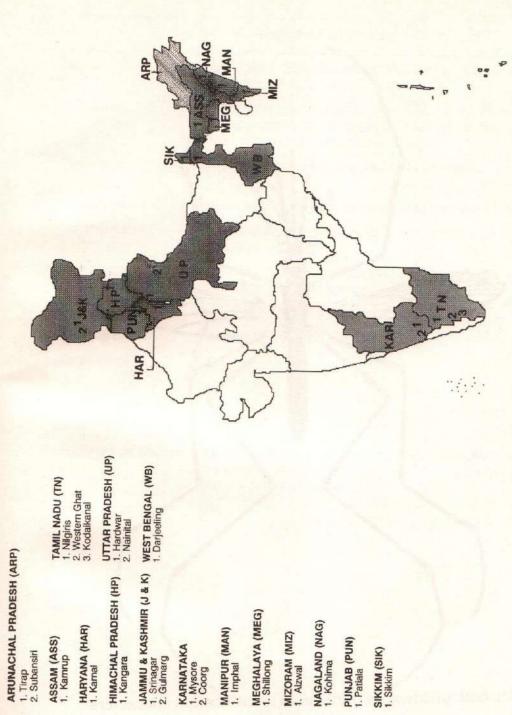
Relation to disease : NA

Reported distribution: The type form is distributed in Bangladesh, Borneo,

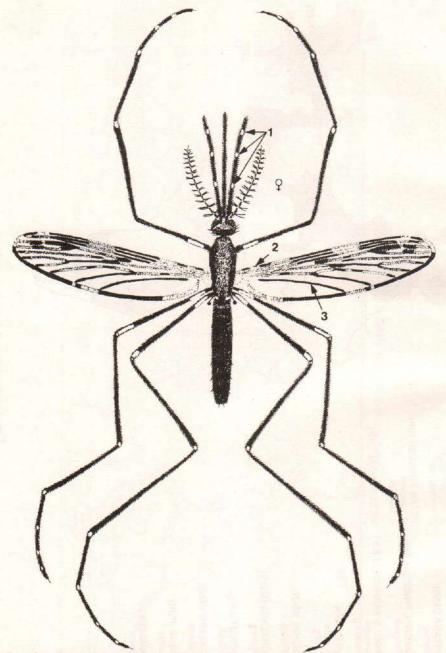
India, Myanmar, Nepal, Sri Lanka, and Sumatra. The variety *simlensis* recorded from India, Nepal, Pakistan, Sri Lanka. In India, the type form and its variety *simlensis* are reported from Arunachal Pradesh, Assam, Delhi, Haryana, Himachal Pradesh, Karnataka, Kashmir, Manipur, Meghalaya, Mizoram, Nagaland, Punjab, Sikkim, Tamil Nadu, Uttar Pradesh, and West

Bengal.





Reported distribution of An. gigas in India.



An. baileyi Edwards 1929, main identification characters

- Palpi dark but with small bands at joints
 Inner costa interrupted
 Outer half of vein 6 (Anal vein) dark.

Anopheles baileyi

Name : An. baileyi Edwards, 1929. Bulletin of Entomological

Research 20: 321-343.

Derivative : Edwards named it after Lt. Col. F.M. Bailey. Previously

it was a variety of An. gigas but recently Harrison et al.

1991 raised it to species level.

Type form : Available at the British Museum of Natural History,

London.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : A wild species but also rests in cattlesheds and human

dwellings.

Breeding ecology : Species breeds throughout the year in deep pools, rocky

pools and perennial springs.

Biting time : NA

Feeding preference: Wild animals.

Flight range : NA

Susceptibility : NA

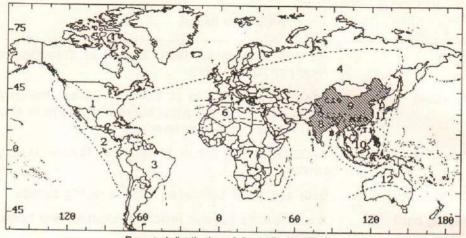
to insecticides

Relation to disease : NA

Reported distribution: Occurs in Bangladesh, Central China, Indochina, India,

Myanmar, Taiwan and Tibet. In India recorded from

Assam and Meghalaya.



Reported distribution of An. baileyi in World.

ASSAM (ASS) 1. Kamrup

MEGHALAYA (MEG) 1. Shillong



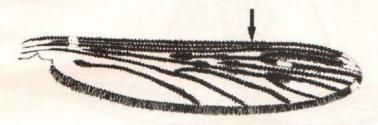
Reported distribution of An. baileyi in India.

2.8. An. annandalei, An. interruptus

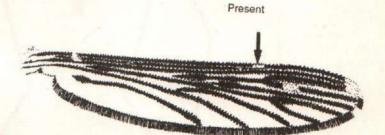
Wing with 3 or less pale areas on the costa, subcosta including vein 1(R1) Palpi dark but with small pale bands at the joints Hind femur with a tuft of dark and pale scales towards apex

Subcostal pale spot on costa

Absent



An. annandalei



An. interruptus

The species can be confirmed by the larval characters Branches of the thoracic pleural hairs

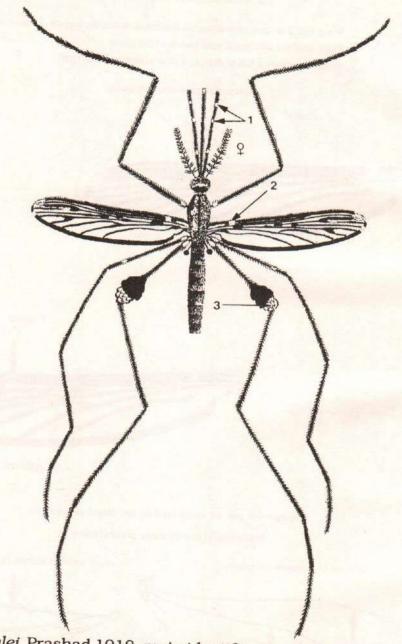
Without any branch (simple)

Very small branches (barb-like)

An. annandalei

a the same of the

An. interruptus



An. annandalei Prashad 1918, main identification characters

- 1. Palpi completely dark but small bands at joints
- 2. Inner costa interrupted
- 3. Apex of the hind femur with prominant tuft of white and black scales

Further identification is based on larval characters

Anopheles annandalei

Name : An. annandalei Prasad 1918. Records of the Indian

Museum, 15: 123-127.

Derivative : Prasad named it after Dr. N. Annandale, Director,

Zoological Survey of India.

Type form : Available at the Zoological Survey of India, Calcutta,

India.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : A forest dweller, rests outdoors.

Breeding ecology : Breeds in tree holes in deep forests or wooded country.

Biting time : Bites before midnight.

Feeding preference: Wild animals.

Flight range : NA

Susceptibility to :

insecticides

: NA

Relation to disease : NA

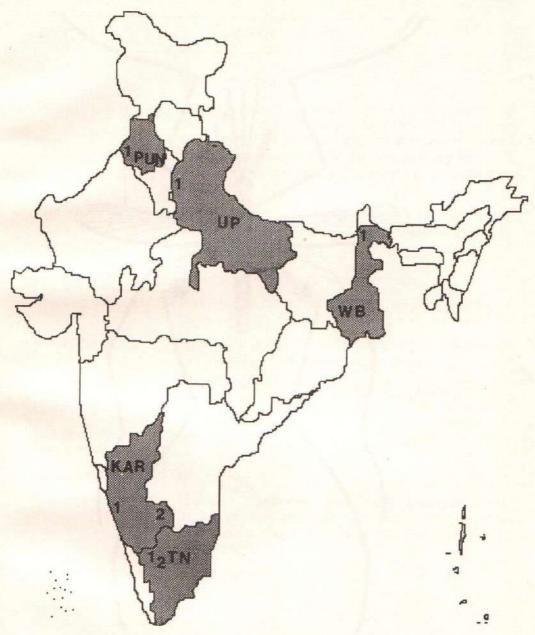
Reported distribution: Occurs in India, Indonesia (Java), and Sri Lanka. In

India, recorded from Karnataka, Punjab, Tamil Nadu,

Uttar Pradesh and West Bengal.



Reported distribution of An. annandalei in the World.



KARNATAKA (KAR) 1. Shimoga 2. Hassan

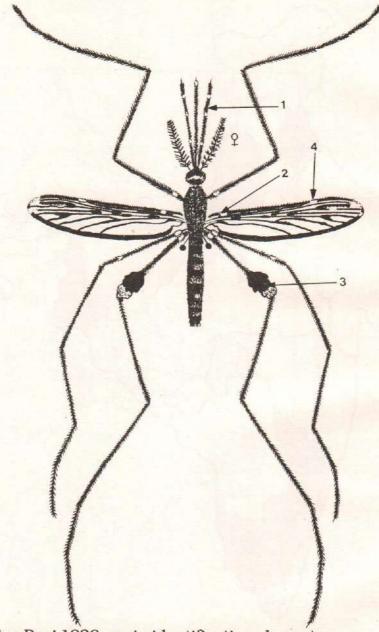
PUNJAB (PUN) 1. Ferozepur

TAMIL NADU (TN) 1. Nilgiris 2. Western Ghats

UTTAR PRADESH (UP) 1. Dehra Dun

WEST BENGAL (WB)
1. Darjeeling

Reported distribution of An. annandalei in India.



An. interruptus Puri 1929, main identification characters

- 1. Palpi completely dark but small bands at joints
- 2. Inner costa interrupted
- 3. Apex of the hind femur with prominant tuft of white and dark scales
- 4. Sub costal pale spot on costa

Further identification is based on larval characters

Anopheles interruptus

Name : An. interruptus Puri, 1929. The Indian Journal of Medical

Research, 17: 385-395.

Derivative : Latin, interruptus, interrupted, broken off; (interrumpere,

to interrupt, break off, sever, break between). Puri does not specifically give reasons for choosing the name interruptus, but it refers to the pale spot on the costa of interruptus, this spot being absent in the type form

annandalei.

Type form : Available at the British Museum of Natural History,

London; Zoological Survey of India, Calcutta, India.

Sitting posture : Body and mouth parts at an angle to resting surface.

Resting habit : A rare species, rests outdoors in forests.

Breeding ecology : A tree-hole breeder, collected up to an altitude of 900 m.

Larvae are very dark.

Biting time : Before midnight. In Thailand peak biting time is between

19.00 and 20.00 hrs in deep forest areas.

Feeding preference: Wild animals.

Flight range : NA

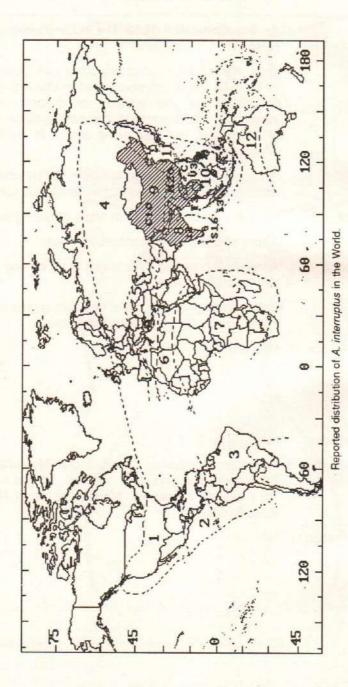
Susceptibility to : NA insecticides

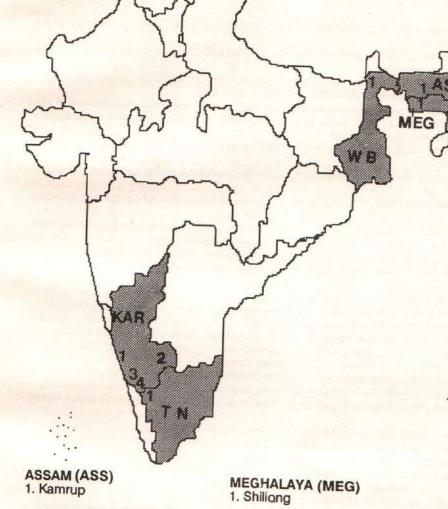
Relation to disease : NA

Reported distribution: Found in Cambodia, China, India, Myanmar, Nepal,

Sri Lanka, Thailand, and Vietnam. In India recorded from Assam, Karnataka, Meghalaya, Tamil Nadu, and

West Bengal.





Reported distribution of A. interruptus in India.

TAMIL NADU (TN)
1. Nilgiris

WEST BENGAL (WB)
1. Darjeeling

KARNATAKA (KAR)
1. Hassan
2. Shimoga
3. Mysore
4. Coorg

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GLOSSARY

Acellular. The term applied to unicellular organisms which are complex in structure and function.

Adaptation. The condition of showing fitness for a particular environment, as applied to characteristics of a structure, function, or entire organism; also the process by which such fitness is acquired.

Affinity. Relationship. Sometimes misleadingly used as synonym for phenetic similarity.

Allochronic species. Species which do not occur at the same time level.

Allopatric. Of populations or species, occupying mutually exclusive (but usually adjacent) geographical areas.

Allopatric speciation. Species formation during geographical isolation.

Allotype. A paratype of the opposite sex to the holotype.

Analogous. The similar function but not of similar origin.

Alpha taxonomy. The level of taxonomy concerned with the characterization and naming of species.

Anatomy. The science of internal morphology, as revealed by dissection.

Anopheles. Genus (or subgenus) of mosquito of the subfamily Anophelinae.

Anopheline. Subfamily of the Culicidae which includes the genus Anopheles.

Anophelism. Presence of anophelines in a locality, irrespective of the presence of malaria.

Anthropophilic. Showing a preference for feeding on man even when non-human hosts are available.

A relative term requiring qualification so as to indicate the extent of this preference.

Artificial classification. Classification based on convenient and conspicuous diagnostic characters without attention to characters indicating relationship; often a classification based on single, arbitrarily chosen character instead of an evaluation of the totality of characters.

Atlas. In taxonomy, a method of presenting taxonomic materials primarily by means of comparative illustrations rather than by comparative descriptions (e.g. monograph).

Beta taxonomy. The level of taxonomy concerned with the arranging of species into a natural system of lower and higher taxa (e.g. Alpha taxonomy, Gamma taxonomy).

BHC (See also HCH). Abbreviation and common name of benzene hexachloride, a chlorinated hydrocarbon insecticide comprising essentially a mixture of isomers of 1,2,3,4,5,6-hexachlorocyclohexane in the form of whitish to light-brown granules, flakes, or powder. The gamma-isomer is the most active constituent. The grades of BHC commercially available are distinguished according to their gamma-isomer content as follows: technical BHC: 12%-16%, refined BHC: 16.1-98.9%, lindane: 99%-100%.

Bilateral symmetry. Symmetry about a single plane, so that there is one way only in which the body can be divided into two similar halves.

Binary. Refers to designations consisting of two kinds of names.

Binomial nomenclature. The system of nomenclature first standardized by Linnaeus and now generally referred to as binominal nomenclature.

Biological classification. The arranging of organisms into taxa on the basis of inferences concerning their genetic relationship.

Biological races. Non-interbreeding sympatric populations, which differ in biology but not, or scarcely, in morphology; supposedly prevented from interbreeding by preference for different food plants or other hosts.

Biological species concept. A concept of the species category stressing reproductive isolation, and the possession of a genetic programme effecting such isolation.

Biota. The flora and fauna of a region.

Biting-capture. Collection of mosquitoes caught in the act of feeding on a human or an animal host.

Biting cycle. Regular variations in the amount of blood-feeding activity exhibited by populations of mosquito species during each 24-hour day and night period.

Cladism. A taxonomic theory by which organisms are ordered and ranked entirely on the basis of "recency of common descent," that is, on the basis of the most recent branching point of the inferred phylogeny.

Classification. The delimitation, ordering, and ranking of taxa.

Clone. All the offspring derived by asexual reproduction from a single sexually produced individual.

Clustering methods. Methods of grouping related or similar species into species groups or higher taxa.

Complex. A neutral term for a number of related taxonomic units, most commonly involving units in which the taxonomy is difficult or confusing.

Compound eye. Eye composed of numerous separate image-forming elements. Found only in insects and crustaceans.

Congeneric. A term applied to species of the same genus.

Conspecific. A term applied to individuals or populations of the same species.

Continuity. In nomenclature, the principle that continuity of usage should take precedence over priority of publication in determining which of two or more competing scientific names should be adopted for a particular taxon.

Continuous variation. Variation in which individuals differ from each other by infinite steps, as variation in quality of expression of a character or group of characters.

Convergence. Morphological similarity in but distantly related forms.

Cotype. Syntype (q.v.).

Cryptic species. Sibling species (q.v.).

DDT. Abbreviation and common name of dichlorodiphenyl-trichloroethane, a chlorinated hydrocarbon insecticide comprising essentially 1,1,1-trichloro-2,2-di-(p-chlorophenyl)ethane in the form of white or cream-coloured granules, flakes or powder. Technical DDT should contain a minimum of 70% by weight of p, p'-isomer having a minimum melting point of 104°C.

Density, anopheline. Number of female anophelines in relation to the number of specified shelters or hosts (e.g. per room, per trap, or per person) or to a given time period (e.g. overnight or per hour), specifying method of collection.

Dieldrin. Chlorinated hydrocarbon insecticide, comprising essentially 1,2,3,4,10,10-hexachloro-6, 7-epoxy-1,4,4a,5,6,7,8,8a-octahydro-endo-1,4-exo-5,8-dimethanonaphthalene (HEOD). Technical dieldrin should contain at least 76.5% by weight of HEOD.

Diploblastic. Having the body organised from only two embryonic cell layers (ectoderm and endoderm).

Direct development. Development without a larval form.

Discontinuous variation. Variation in which the individuals of a sample fall into definite classes which do not grade into each other.

Distal. Situated away from the point of origin or attachment.

Distribution. Geographical, seasonal, ecological or topographical range of an organism.

Ecological isolation. A condition in which interbreeding between two or more otherwise sympatric populations is believed to be prevented by mating in different ecologic niches.

Ecological race. A local race that owes its most conspicuous attributes to the selective effect of a specific environment.

Ecology. The study of the interactions between organisms and their environment.

Ecospecies. "A group of population so related that they are able to exchange genes freely without loss of fertility or vigour in the offspring" (Turesson).

Ecotype. A descriptive term applied to plant races of varying degrees of distinctness which owe their most conspicuous characters to the selective effects of local environments.

Emendation. In nomenclature, an intentional modification of the spelling of a previously published scientific name.

Endophagy. Tendency of mosquitoes to feed indoors.

Endophily. Tendency of mosquitoes to rest indoors, whether by day or by night.

Endopterygote insect. Insect with complete metamorphosis, i.e. the juvenile form is a wingless larva unlike adult in both appearance and mode of life. The abrupt change into the adult form takes place during a pupal stage.

Eucaryotes. Organisms with a well-defined nucleus and meiosis. All higher organisms above the level of procaryotes (q.v.).

Excito-repellency. Property in an insecticide of stimulating irritability in insects.

Exophagy. Tendency of mosquitoes to feed outdoors.

Exophily. Tendency of mosquitoes to rest outdoors, whether by day or by night.

Exopterygote insect. Insect with incomplete metamorphosis, i.e. the juvenile form is a nymph whose wings develop gradually by small increments at each moult.

Evolution. The process of very slow and gradual change which occurs in the descendent generations of populations of organisms by a process of natural selection.

Family. A taxonomic category including one genus or a group of genera or tribes of common phylogenetic origin, which is separated from related similar units (families) by a decided gap, the size of the gap being in inverse ratio to the size of the family.

Family name. The scientific designation of a taxon of family rank, recognized by the termination idae, which termination may not be used in names of other taxa.

Fauna. The animal life of a region.

Faunal work. A publication in which taxa are included on the basis of their occurrence in a specified area rather than on the basis of relationship.

Flight, migratory. Directional flight of a group of mosquitoes, not subject to the ordinary laws of dispersion.

Flight, pre-hibernation. Dispersion or migration of mosquitoes occurring before and in relation to hibernation, often to greater-than-normal distances.

Flight range, effective. Distance from a breeding place that the females of a given species of mosquito travel in numbers sufficient to maintain endemic malaria or to cause an epidemic.

Flora. The plant life of a region.

Gamma taxonomy. The level of taxonomy dealing with various biological aspects of taxa, ranging from the study of intraspecific populations to studies of speciation and of evolutionary rates and trends.

Genotype. The genetic constitution of an individual or taxon (cf. Phenotype). Use of this term in nomenclature for the type-species of a genus is confusing and contrary to the terminology of the Code.

Genus. A category for a taxon including one species or a group of species, presumably of common phylogenetic origin, which is separated from a related, similar unit (genera) by a decided gap, the size of the gap being in inverse ratio to the size of the unit (genus).

Geographic isolate. A population that is separated by geographic barriers from the main body of the species.

Geographical race. Subspecies (q.v.).

Geographical reconnaissance. In malaria eradication terminology, the operation which provides the basis for the choice of field centres and depots, for detailed schedules and itineraries of spraying and surveillance personnel, for the final deployment of transport, and for numerical control of the completeness of work accomplished. It includes collection of information on the number, type, location and means of access of all houses and field shelters, as well as on communications, health units, vehicle-repair facilities, population movements and other relevant factors.

Gynandromorph. An individual in which one part of the body is masculine, the other feminine; most frequent are bilateral gynandromorphs, in which the left and right halves are of different sex.

HCH. HCH (Hexachlorocyclohexane), also known as BHC belongs to Organochlorine. Technical-grade HCH consists of 65-70% alpha-HCH, 7-10% beta-HCH, 14-15% gamma-HCH, and approximately 10% of other isomers and compounds. Lindane contains > 99% gamma-HCH. It is a solid, with a low vapour pressure, and is poorly soluble in water but very soluble in organic solvents, such as acetone, and in aromatic and chlorinated solvents. The n-octanol/water partition coefficient (log P) is 3.2-3.7.

Holistic. Looking at wholes as more than the sums of their parts.

Holotype. The single specimen designated or indicated as "the type" by the original author at the time of the publication of the original description.

Homologous. A feature in two or more taxa which can be traced back to the same feature in the common ancestor of these taxa.

Homonym. In nomenclature, one of two or more identical but independently proposed names for the same or different taxa.

Host races. Different genetic races of the same species in oligophagous food specialists or parasites occurring on different hosts.

Hypodigm. The entire material of a species that is available to a taxonomist.

Identification. The determination of the taxonomic identity of an individual.

Infraspecific. Within the species; usually applied to categories (subspecies) and phena (varieties). Infrasubspecific form. Individual and seasonal variants in a single interbreeding population.

Infrasubspecific name. A name given to an infrasubspecific form.

Insecticide. Product that kills insects either in their immature stages ("ovicide, larvicide") or in their adult stage ("immediate imagicide" or "residual imagicide").

Instar. Stage between successive ecdyses during the larval development of an insect.

International Code of Zoological Nomenclature. The official set of regulations dealing with zoological nomenclature.

Intersex. An individual more or less intermediate in phenotype between male and female.

Junior homonym. The more recently published of two or more identical names for the same or different taxa.

Junior synonym. The more recently published of two or more available synonyms for the same taxon.

Key, taxonomic. List of distinguishing morphological characters so arranged as to facilitate identification of species. The commonest form is the dichotomous key, in which the characters are displayed as pairs of alternatives ("couplets").

Lectotype. One of a series of syntypes which, subsequent to the publication of the original description, is selected and designated through publication to serve as "the type".

Lumper. A taxonomist who emphasizes the demonstration of relationship in the delimitation of taxa and who tends to recognize large taxa.

Macrotaxonomy. The classification of higher taxa.

Malathion. An organophosphorus insecticide. Technical malathion should contain O,O-dimethyl S-[1,2-di-(ethoxycarbonyl) ethyl] phosphorodithioate in proportion by weight of not less than 95%.

Metamorphosis. The series of changes which take place as a larva transforms into the adult form.

Monograph. In taxonomy, an exhaustive treatment of a higher taxon in terms of all available information pertinent to taxonomic interpretation; usually involving full systematic treatment of

information pertinent to taxonomic interpretation; usually involving full systematic treatment of the comparative anatomy, biology, ecology, and detailed distributional analyses of all included taxa.

Monotypic. The taxon containing only one immediately subordinate taxon, as a genus containing one species, or a species containing but one (the nominate) subspecies.

Morphospecies. A typological species recognized merely on the basis of morphological differences. Multivariate analysis. The simultaneous analysis of several variable characters.

Natural selection. The unequal contribution of genotypes to the gene pool of the next generation, through differential mortality and differences in reproductive success, caused by components of the environment.

Neoteny. Attainment of sexual maturity in an immature or larval stage.

Neotype. A specimen selected as type subsequent to the original description in cases where the original types are known to be destroyed or were suppressed by the Commission.

New name. A replacement name for a preoccupied name.

Niche (ecological). The precise constellation of environmental factors into which a species fits or which is required by a species.

Nomenclator. A book containing a list of scientific names assembled for nomenclatural, rather than taxonomic purposes.

Nomenclature. A system of names.

Nomen dubium. The name of a nominal species for which available evidence is insufficient to permit recognition of the zoological species to which it was applied.

Nomen oblitum. A name losing its validity under the statute of limitation.

Nominal taxon (species, genus etc.). A named taxon, objectively defined by its type.

Non-dimensional species. The species concept represented by the non-interbreeding of species at a given place and time.

Nymph. Juvenile stage of exopterygote insect, resembling adult in many ways, but lacking wings.

Oligogenic character. A character determined by only a few genes.

Ontogeny. The developmental history of an individual organism from egg to adult.

Original description. A statement of characters accompanying the proposal of a name for a new taxon in conformity with Arts.

Parasite. Organism living in or on another living organism (host) from which it derives its food.

Paratype. A specimen other than the holotype which was before the author at the time of preparation of the original description and was so designated or indicated by the original author.

Patronymic. In nomenclature, dedicatory name, a name based on that of a person or persons.

Phylogency. The study of the history of the lines of evolution in a group of organisms; the origin and evolution of higher taxa.

Polytypic. A taxon containing two or more taxa in the immediately subordinate category, as a genus with several species or a species with several subspecies.

Pre-Linnaean name. A name published prior to 1 January, 1758, the starting point of zoological nomenclature.

Primary homonym. Each of two or more identical species-group names which, at the time of original publication, were proposed in combination with the generic name (e.g. X-us albus Smith, 1910, and X-us albus Jones, 1920).

Primary zoological literature. Literature dealing with animals or zoological phenomena, not merely a listing of names.

Priority. The principle that of two competing names for the same taxon (below the rank of an infraorder) ordinarily that is valid which was published first.

Proboscis. Insect mouth parts specialised for sucking up fluid. Trunk of elephant.

Procaryotes. Microorganisms (virus, bacteria, blue-green algae) that lack well defined nuclei and

Pupa. Stage between the larva and the imago in the development of the mosquito, during which it is aquatic and active but does not feed. When the larva undergoes metamorphosis into a pupa ("pupation") the fourth and last larval skin is cast.

Race. Genetically distinct mating group within a species. To be distinguished from a subspecies, which is geographically isolated as well as being genetically distinct.

Race. Subspecies (q.v).

Rate, anopheline infection. Percentage of female anophelines of a given species found, by dissection within 24 hours of capture, to contain malaria parasites either as sporozoites in the salivary glands or as occysts on the midgut wall.

Rate, biting. Average number of mosquito bites received by a host in unit time, specified according to host and mosquito species.

Rate, oocyst. Percentage of female Anopheles caught in nature and found, on dissection within 24 hours of capture, to contain oocysts in the midgut.

Rate, sporozoite. Percentage of female Anopheles caught in nature and found, on dissection within 24 hours of capture, to contain sporozoites in the salivary glands.

Resistance. 1. Ability of a parasite strain to multiply or to survive in the presence of concentrations of a drug that normally destroy parasites of the same species or prevent their multiplication. Such resistance may be relative (yielding to increased doses of the drug tolerated by the host).

2. Ability in a population of insects to tolerate doses of an insecticide which would prove lethal to the majority of individuals in a normal population of the same species; developed as a result of selection pressure by the insecticide. Simultaneous resistance to one or more insecticides of two different groups is known as double insecticide resistance (not to be confused with insecticide cross-resistance).

Scientific name. The binominal or trinominal designation of an animal; the formal nomenclatural designation of a taxon.

Secondary homonym. Each of two or more identical specific names which, at the time of original publication were proposed in combination with different generic names but which through subsequent transference, reclassification or combination of genera have come to bear the same (or an identical) combination of a generic and specific name.

Semispecies. The component species of superspecies (Mayr); also, populations that have acquired some, but not yet all, attributes of species rank; borderline cases between species and

subspecies.

Senior homonym. The earliest published of two or more identical names for the same or different

Senior synonym. The earliest published of two or more available synonyms for the same taxon.

Sibling species. Pairs or groups of closely related species which are reproductively isolated but morphologically identical or nearly so.

Species. Group of organisms capable of exchanging genetic material with one another and incapable, by reason of their genetic constitution, of exchanging such material with any other group of organisms. The limits of species are indicated by the comparative study of morphological and other characters.

Species complex. Group of closely related organisms, the exact specific status of which is uncertain although they resemble some well-recognized type species.

Species eradication. Complete elimination from a territory of one or more species of malaria vectors. Species-group. An assemblage of coordinate categories, viz. species and subspecies.

Species. Groups of actually (or potentially) interbreeding natural populations which are reproductively isolated from other such groups.

Subfamily. A category of the family-group subordinate to the family; and individual taxon ranked in the category subfamily.

Subgeneric name. The name of an optional category between the genus and the species enclosed in parentheses when cited in connection with a binominal or trinominal combination and therefore excluded from consideration when determining the number of words of which a specific or subspecific name is composed (e.g., X-us (Y-us) albus rufus is a trinominal).

Subjective synonym. Each of two or more synonyms based on different types, but regarded as referring to the same taxon by those zoologists who hold them to be synonyms.

Subspecies. A geographically defined aggregate of local populations which differs taxonomically from other such subdivision of the species.

Substitute name. A name proposed to replace a preoccupied name and automatically taking the same type and type-locality (= New name).

Superfamily. The taxonomic category immediately above the family and below the order; an individual taxon ranked in this category.

Superspecies. A monophyletic group of entirely or largely allopatric species.

Supraspecific. A term applied to a category or evolutionary phenomenon above the species level.

Susceptibility. 1. Liability in a population of insects to be killed by a particular insecticide. The average susceptibility of a species or population of mosquitoes is usually measured in terms of the median lethal concentration (LC50). 2. Liability of a species of mosquito to become infected when fed on a person known to be infectious; usually measured in relation to the liability of infection of another species fed at the same time on the same person. 3. Liability of a person to become infected. The lower the immunity, the higher the susceptibility.

Swarming. Nuptial flight or dance in which males of some species of mosquito gather together on the wing, usually at dusk or in dim light, in the open, moving up and down without horizontal progression; the females fly into the swarm of males, and copulation takes place in the air.

Sympatric speciation. Speciation without geographic isolation; the acquisition of isolation mechanisms within a deme.

Sympatry. The occurrence of two or more populations in the same area; more precisely, the existence of a population in breeding condition within the cruising range of individuals of another population.

Synchronic species. Species which occur at the same time level.

Synonym. In nomenclature, each of two or more different names for the same taxon.

Synonymy. A chronological list of the scientific names which have been applied to a given taxon, including the dates of publication and the authors of the names.

Synopsis. In taxonomy, a brief summary of current knowledge of a group.

Syntype. Every specimen in a type-series in which no holotype was designated.

Systematics. The science dealing with the diversity of organisms.

Taxon. A taxonomic group that is sufficiently distinct to be worthy of being distinguished by name and to be ranked in a definite category.

Taxonomic category. Designates rank or level in a hierarchic classification. It is a class, the members of which are all taxa assigned a given rank.

Taxonomic character. Any attribute of a member of a taxon by which it differs or may differ from a member of a different taxon.

Taxonomy. The theory and practice of classifying organisms.

Topotype. A specimen collected at the type-locality.

Tribe. A taxonomic category intermediate between the genus and the subfamily.

Trinominal nomenclature. An extension of the binominal system of nomenclature to permit the designation of subspecies by a three-word name.

Triploblastic. Having the body organised from three embryonic layers of cells; ectoderm, mesoderm and endoderm.

Type. A zoological object which serves as the base for the name of a taxon.

Type designation. Determination of the type of a genus under Articles 67-69 of the Code.

Type-locality. The locality at which a holotype, lectotype, or neotype was collected.

Type-species. The species which was designated as type of a nominal genus.

Uninominal nomenclature. The designation of a taxon by a scientific name consisting of a single word; required for taxa above species rank.

Univariate analysis. A biometric analysis of a single character.

Valid name. An available name that is not preoccupied by a valid senior synonym or homonym.

Variation, ecophenotypic. Variation caused by non-genetic response of the phenotype to local conditions of habitats, climate, etc.

Variety. An ambiguous term of classical (Linnaean) taxonomy for a heterogeneous group of phenomena including non-genetic variations of the phenotype, morphs, domestic breeds, and geographic races.

Vector. In malaria, any species of mosquito in which the *Plasmodium* completes its sexual cycle in nature and which is thus able to transmit the disease.

Vector, conditional. Species of mosquito known to transmit malaria but presumed to be incapable of maintaining endemic malaria in the absence of more efficient vectors.

Vector, principal. Species mainly responsible for transmitting malaria in any particular circumstances. Principal vectors may overlap or may alternate seasonally.

Vector, secondary. Species thought to play a minor role in transmission in association with a principal vector and to be capable of maintaining malaria at a reduced level in the absence of the latter.

Vector control. Measures of any kind directed against a vector of disease and intended to limit its ability to transmit the disease.

Vector efficiency. Ability of a mosquito species, in comparison with another species in a similar climatic environment, to transmit malaria in nature. A rough estimate of relative efficiency may be made by comparison of sporozoite rates taken in comparable conditions.

Vernacular name. The colloquial designation of a taxon.

Vertical classification. Classification which stresses common descent and tends to unite ancestral and descendant groups of a phyletic line in the single higher taxon, separating them from contemporaneous taxa having reached a similar grade of evolutionary change.

Zone. In malaria eradication terminology, a territorial and administrative operational unit responsible for the operation, evaluation and administration of the field programme in its areas, which often coincides with an administrative division of the country. The zone office is normally under the direction of a senior professional officer, responsible for execution of all operations and routine evaluation of results in the zone.

Zoological Nomenclature. International Code of rules established by the International Commission on Zoological Nomenclature and periodically revised.

Zoonosis. Disease or infection common to man and other vertebrates and naturally transmitted between these hosts.

Zoophilic. Term applied to mosquitoes showing a relative preference for non-human blood even when human hosts are readily available.



Indian Anophelines is the first book of its kind on the fauna of anopheline mosquitoes from India. The following aspects of mosquito systematics, biology and distribution are described:

Worldwide distribution of anophelines in 12 malaria epidemiological zones; Reported distribution of anophelines in India; Bio-ecology and behaviour of mosquitoes; Updated vector status; Pictorial keys; Bibliography; and Glossary.

Indian Anophelines assume special importance because of the deteriorating malaria situation in India, complicated by vector resistance to insecticides, ecological succession of mosquitoes, invasion of mosquitoes to new areas, as also their disappearance from certain areas. As a result mosquito fauna has undergone major changes and this precise knowledge at the local level in endemic regions is invariably lacking. Often the identification is made difficult due to variations in many appendages. For each anopheline species the book provides names, derivatives, type form availability, resting habits. breeding ecology, biting time, flight range, susceptibility to insecticides, relation to disease. reported distribution in India and the world, and results of vector incrimination studies. Using this book it is easy to identify specimens correctly up to species level. Taxonomic description of each species is supported by high quality illustrations giving distinguishing features of each species and their variations. The style of presentation is lucid and simple. This book is intended as a reference material for students of mosquito systematics. The book therefore would be a valuable addition in the libraries and a source of knowledge for the students engaged in research and those interested in the study of the bioecology and control of mosquitoes.

Dr. B.N. Nagpal did M.Sc. Zoology (entomology) in 1979 from Allahabad University, and joined the Malaria Research Centre as Research Fellow. He obtained Ph.D. from Berhampur University in 1986 for his thesis on "Mosquitoes of Orissa" under the guidance of Dr. V.P. Sharma. He has extensively surveyed large endemic areas of the country for the collection of mosquitoes and has studied in depth their bioecology under natural conditions. Based on his field-collected specimens a museum on mosquitoes has been established at the MRC. The museum contains valuable mosquito specimens of type form and variations found in the field. This book is an outcome of his 15 years of field work supported by laboratory investigations on mosquito systematics.

Dr. V.P. Sharma is D.Phil. and D.Sc. from Allahabad University. He is fellow of learned societies, FNA, FNASc, FAMS, FRAS, and has published more than 200 research and review papers in national and international journals. The first director of the Malaria Research Centre, Dr. Sharma has raised it to the status of an important national laboratory in less than a decade. Dr. Sharma is well known for the development of bioenvironmental malaria control strategy—he has developed a strategy which has found wide application in managing malaria in the field. He has received several awards—Inventions award of Government of India (1971), B.K. Srivastava Oration award (1981), Om Prakash Bhasin award (1985), R.V. Rajam Oration award (1987), M.L. Gupta Trust award (1989), B.N. Singh Oration award (1990), Ranbaxy award (1990), and MOT lyengar award (1990). He was honoured as Padma Shri in 1992. Dr. Sharma is Chairman, Panel of Experts on Environmental Management for Vector Control (PEEM) of the World Health Organization (WHO); Council Member of the International Congress of Entomology; Consultant to WHO on several occasions; and Board Member of the Malaria Foundation Incorporated, New York.

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